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Washington, D.C. 20231

Attorney Docket No. 16930-0010-22

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By: *James Kane*

Transmitted herewith for filing under 37 CFR 1.53(b) is the
☐ patent application of
☐ continuation patent application of
☒ divisional patent application of
☐ continuation-in-part patent application of

Inventor(s)/Applicant Identifier: Douglas Antelman, Richard J. Gregory and Kenneth N. Wills

For: **METHODS OF TREATING HYPERPROLIFERATIVE DISORDERS USING RETINOBLASTOMA FUSION PROTEINS**

☒ This application claims priority from each of the following Application Nos./filing dates:
08/801,092 filed February 14, 1997; 08/751,517 filed November 15, 1996

the disclosure(s) of which is (are) incorporated by reference.

☒ Please amend this application by adding the following before the first sentence: "This application is a division of and claims the benefit of U.S. Application No. 08/801,092 filed February 14, 1997, which is a continuation of and claims the benefit of U.S. Application No. 08/751,517 filed November 15, 1996, the disclosures of which are incorporated by reference."

Enclosed are:

- ☒ 31 page(s) of specification
☒ 4 page(s) of claims
☒ 1 page of Abstract
☒ 51 sheet(s) of ☐ formal ☒ informal drawing(s).
☒ An assignment of the invention to Canji, Inc. was recorded in the prior application
☒ A ☒ signed ☐ unsigned Declaration & Power of Attorney (copy from prior application)
☒ A Preliminary Amendment.
☒ Sequence Listing pages 32-62

Claims after Entry of any Amendments, Less any Canceled Claims

	(Col. 1)	(Col. 2)
FOR:	NO. FILED	NO. EXTRA
BASIC FEE		
TOTAL CLAIMS	22 - 20	= *2
INDEP. CLAIMS	1 - 3	= *0
<input type="checkbox"/> MULTIPLE DEPENDENT CLAIM PRESENTED		

* If the difference in Col. 1 is less than 0, enter "0" in Col. 2.

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+ \$130.00 =	
TOTAL	

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OTHER THAN SMALL ENTITY

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Respectfully submitted,
TOWNSEND and TOWNSEND and CREW LLP

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By: 

PATENT
Attorney Docket No. 16930-001022US

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re application of:

Douglas Antelman *et al.*

Serial No.: not yet assigned

Filed: May 19, 1999

For: METHODS OF TREATING
HYPERPROLIFERATIVE
DISORDERS USING
RETINOBLASTOMA FUSION
PROTEINS

Examiner: Lin Sun-Hoffman, Ph.D.
(parent)

Art Unit: 1642 (parent)

PRELIMINARY AMENDMENT

Assistant Commissioner of Patents and Trademarks
Washington, D.C. 20231

Sir:

Applicant respectfully requests entry of the following amendments prior to
examination of the application.

In the Title:

Please delete the title and insert therefor: --METHODS OF TREATING
HYPERPROLIFERATIVE DISORDERS USING RETINOBLASTOMA FUSION PROTEINS--.

In the Specification:

At page 2, line 5, please delete "518-521" and insert --518-522--.

At page 3, line 17, after "Figure 1A" insert --(SEQ ID NO:1)--.

At page 3, line 19, after "Figure 1B" insert --(SEQ ID NO:2)--.

At page 3, line 21, after "Figure 2A" insert --(SEQ ID NO:3)--.

At page 3, line 23, after "Figure 2B" insert --(SEQ ID NO:4)--.

At page 3, line 26, after "Figure 4" insert --(SEQ ID NOS:5-18)--.

At page 3, line 32, after "Figure 8" insert --(SEQ ID NOS:33-46)--.

At page 3, line 19, after "Figure 1A" insert --(SEQ ID NO:2)--.

At page 10, line 9, please delete "Hartzoglou" and insert --Hatzoglou--.

At page 11, at line 18, please delete "1098" and insert --1089--. At line 19, please delete "Casper" and insert --Kasper--. At line 24, please delete "Tanura" and insert --Tamura--.

At page 19, line 12, please delete "11" and insert --111--. At line 20, please delete "Willart" and insert --Willard--. At line 21, please delete "1995" and insert --1994--.

At page 27, line 6, please delete "Thimmappaaya" and insert --Thimmappaya--.

After page 31, please insert the enclosed paper copy of the sequence listing, which consists of pages 32-62. Please renumber the following pages accordingly.

In the Claims:

Please cancel claims 1-15.

Please amend claims 16, 17, 22-26 and 32-33 as follows:

1 16. (Once amended) A method for treatment of a hyperproliferative disorder in a
2 patient, the method comprising administering to a patient a therapeutically effective dose of a
3 fusion polypeptide that comprises [comprising a fusion of] a DNA binding domain of a
4 transcription factor and a functional growth suppression domain of a [, the transcription factor
5 comprising a DNA binding domain, and a] retinoblastoma (RB) polypeptide [, the RB
6 polypeptide comprising a growth suppression domain].

1 17. (Once amended) The method of claim 16, wherein the fusion polypeptide
2 [protein] is encoded by a nucleic acid delivered to the patient.

1 18. (Reiterated) The method of claim 16, wherein the transcription factor is E2F.

1 19. (Reiterated) The method of claim 18, wherein the cyclin A binding domain
2 of the E2F is deleted or nonfunctional.

1 20. (Reiterated) The method of claim 16, wherein the RB is RB56.

1 21. (Reiterated) The method of claim 16, wherein the RB is wild type RB56.

1 22. (Once amended) The method of claim 16, wherein the functional growth
2 suppression domain of the RB polypeptide comprises from about amino acid residue 379 to
3 about amino acid residue 928 (SEQ ID NO:4).

1 23. (Once amended) The method of claim 16, wherein the functional growth
2 suppression domain of the RB polypeptide comprises at least one substitution of amino acid
3 residues selected from the group consisting of 2, 608, 612, 788, 807, and 811.

1 24. (Once amended) The method of claim 18, wherein the E2F polypeptide
2 comprises about amino acid residues 95 to about 286 (SEQ ID NO:1).

1 25. (Once amended) The method of claim 18, wherein the E2F polypeptide
2 comprises about amino acid residues 95 to about 194 (SEQ ID NO:1).

1 26. (Once amended) The method of claim 16, wherein the fusion polypeptide
2 comprises EF2 amino acid residues from about 95 to about 194 (SEQ ID NO:1) operatively
3 linked to RB amino acid residues from about 379 to about 928 (SEQ ID NO:4).

1 27. (Reiterated) The method of claim 18, wherein the E2F-RB fusion
2 polypeptide is expressed under the control of a tissue-specific promoter.

1 28. (Reiterated) The method of claim 27, wherein the tissue specific promoter is
2 a smooth muscle actin promoter.

1 29. (Reiterated) The method of claim 16, wherein the hyperproliferative
2 disorder is cancer.

1 30. (Reiterated) The method of claim 29, wherein the cancer is bladder cancer.

1 31. (Reiterated) The method of claim 29, wherein the hyperproliferative
2 disorder is restenosis.

1 32. (Once amended) The method of claim 31, wherein the [E2F-RB] fusion
2 polypeptide is administered after angioplasty.

1 33. (Once amended) The method of claim 32, wherein the [E2F-RB] fusion
2 polypeptide is administered as a coating on an angioplasty device.

1 34. (Reiterated) The method of claim 17, wherein the nucleic acid is
2 administered after angioplasty.

1 35. (Reiterated) The method of claim 17, wherein the nucleic acid is
2 administered as a coating on an angioplasty device.

1 36. (Reiterated) The method of claim 17, wherein the nucleic acid is inserted in
2 an adenovirus vector.

Please add the following new claim 37.

1 37. The method of claim 16, wherein the fusion polypeptide lacks a functional
2 cyclin A-kinase binding domain of the transcription factor.

REMARKS

Status of the Application

Claims 16-37 are pending with entry of this amendment, with claims 16-36 previously in the application and entry of new claim 37 respectfully requested.

The Amendments

The amendments to page 3 of the specification provide the sequence ID numbers for the various amino acid and nucleotide sequences. The remaining amendments to the specification correct various errors in the citations of the listed references.

The Sequence Listing

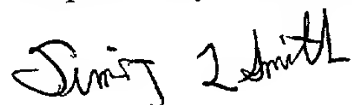
The paper copy of the Sequence Listing in this application is identical to the computer readable copy of the Sequence Listing filed in Application No. 08/801,092, filed February 14, 1997. In accordance with 37 CFR § 1.821(e), please use the only computer readable form filed in that application (filed May 12, 1997) as the computer readable form for the instant application. It is understood that the Patent and Trademark Office will make the necessary change in application number and filing date for the instant application. A paper copy of the Sequence Listing is enclosed herewith for incorporation into the specification. Applicants attest that the information contained in the Sequence Listing introduces no new matter and that the computer-readable form submitted herewith is the same as the paper copy of the Sequence Listing.

CONCLUSION

In view of the foregoing, Applicants believe that all claims now pending in this application are in condition for allowance. The issuance of a formal Notice of Allowance at an early date is respectfully requested.

If a telephone conference would expedite prosecution of this application, the Examiner is invited to telephone the undersigned attorney at (415) 576-0200.

Respectfully submitted,



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PATENT APPLICATION
FOR
TISSUE SPECIFIC EXPRESSION OF RETINOBLASTOMA
PROTEIN

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5 **TISSUE SPECIFIC EXPRESSION OF RETINOBLASTOMA
 PROTEIN**

BACKGROUND OF THE INVENTION

10 Both the retinoblastoma gene (RB) and transcription
factor E2F play a critical role in cell growth control (for a
review, see Adams, P. & Kaelin, W. Seminars in Cancer Biology
6:99-108 (1995)). The RB locus is frequently inactivated in a
variety of human tumor cells. Reintroduction of a wild-type
15 RB gene (e.g., Bookstein et al. Science 247:712-715 (1990)) or
RB protein (pRB) (e.g., Antelman et al. Oncogene 10:697-
704(1995)) into RBneg/RBmut cells can suppress growth in
culture and tumorigenicity *in vivo*.

20 While E2F serves to activate transcription of S-
phase genes, its activity is kept in check by RB. RB arrests
cells by blocking exit from G into S-phase (for example, Dowdy
et al. Cell 73:499-511 (1993)) but the precise pathway of the
arrest remains unclear.

25 Although E2F forms complexes with RB, complex
formation is more efficient if an E2F-related protein, DP-1,
is present. E2F-1 and DP-1 form stable heterodimers which
bind to DNA (for example, Qin et al. Genes and Dev. 6-:953-964
(1992)). DP-1-E2F complexes serve to cooperatively activate
transcription of E2F-dependent genes. Such transcription can
30 be repressed by pRB in the same manner as E2F-1 or DP-1
activated transcription.

35 Transcriptional repression of genes by RB in some
instances can be achieved by tethering pRB to a promoter. For
example, GAL4-pRB fusions bind to GAL4 DNA binding domains and
repress transcription from p53, Sp-1 or AP-1 elements (Adnane,
et al. J. Biol. Chem. 270:8837-8843 (1995); Weintraub, et al.

Nature 358:259-261 (1995)). Sellers, et al. (Proc. Natl. Acad. Sci. 92:11544-11548 (1995)) disclosed fusions of amino acid residues 1-368 of E2F with amino acids 379-792 or 379-928 of RB.

5 Chang, et al. (Science 267:518-521 (1995)) disclosed the use of a replication-defective adenovirus-RB construct in the reduction of neointima formation in two animal models of restenosis, a hyperproliferative disorders.

10 SUMMARY OF THE INVENTION

The instant invention provides the surprising result that a fusion of an E2F polypeptide with an RB polypeptide is more efficient in repressing transcription of the E2F promoter than RB alone, and that such fusions can cause cell cycle
15 arrest in a variety of cell types. Such fusions can thus address the urgent need for therapy of hyperproliferative disorders, including cancer.

One aspect of the invention is a polypeptide comprising a fusion of a transcription factor, the
20 transcription factor comprising a DNA binding domain, and a retinoblastoma (RB) polypeptide, the RB polypeptide comprising a growth suppression domain. Another aspect of the invention is DNA encoding such a fusion polypeptide. The DNA can be inserted in an adenovirus vector.

25 In some embodiments of the invention, the transcription factor is E2F. The cyclin A binding domain of the E2F can be deleted or nonfunctional. The E2F can comprise amino acid residues about 95 to about 194 or about 95 to about 286 in some embodiments.

30 The retinoblastoma polypeptide can be wild-type RB, RB56, or a variant or fragment thereof. In some embodiments, the retinoblastoma polypeptide comprises amino acid residues of about 379 to about 928. Preferred amino acid substitutions of the RB polypeptide include residues 2, 608, 788, 807, and
35 811.

Another aspect of the invention is an expression vector comprising DNA encoding a polypeptide, the polypeptide comprising a fusion of a transcription factor, the

transcription factor comprising a DNA binding domain, and a retinoblastoma (RB) polypeptide, the RB polypeptide comprising a growth suppression domain. In some embodiments a tissue-specific promoter is operatively linked to DNA encoding the fusion polypeptide. The tissue-specific promoter can be a smooth muscle alpha actin promoter.

Another aspect of the invention is a method for treatment of hyperproliferative disorders comprising administering to a patient a therapeutically effective dose of an E2F-RB fusion polypeptide. The hyperproliferative disorder can be cancer. In some embodiments the hyperproliferative disorder is restenosis. The fusion polypeptide and nucleic acid encoding the fusion polypeptide can be used to coat devices used for angioplasty.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1A depicts the predicted amino acid sequence of E2F.

Figure 1B depicts the nucleotide sequence of transcription factor E2F.

Figure 2A depicts the nucleotide sequence of pRB as disclosed by Lee, et al. (Nature 329:642-645 (1987)).

Figure 2B depicts the predicted amino acid sequence of pRB.

Figure 3 is a diagrammatic representation of pCTM.

Figure 4 depicts the nucleotide sequence of plasmid pCTM.

Figure 5 is a diagrammatic representation of pCTMI.

Figure 6 depicts the nucleotide sequence of pCTMI.

Figure 7 is a diagrammatic representation of plasmid pCTMIE.

Figure 8 depicts the nucleotide sequence of pCTMIE.

Figure 9 is a diagram depicting E2F-RB fusion constructs used in the examples. All E2F constructs commenced at amino acid 95 and lacked part of the cyclin A binding domain. E2F-437 contained the DNA binding domain (black), heterodimerization domain (white), and the transactivation domain (stippled). E2F-194 contained solely the DNA binding

domain. E2F-286 contained the DNA binding domain and the DP-1 heterodimerization domain. To generate E2F-194-RB56-5s and E2F-286-RB56-5s, the E2F constructs were fused in-frame to codon 379 of RB. C706F is an inactivating point mutation.

Figure 10 is a diagram depicting transcriptional repression by E2F-RB fusion constructs.

Figure 11 (A-D) depicts expression of E2F-RB fusion proteins in mammalian cell lines. Extracts were prepared from cells used in E2-CAT reporter assays or in FACS assays and analyzed with an anti-RB monoclonal antibody. In panel A, the results are shown from C33A cells transfected with (3) RB56-H209, (4) RB56 wild-type, (5) RB56-5s, (6) E2F286-5s, (7) E2F194-5s, (8) E2F194, (9) E2F286, (10) E2F437. Lane (1) is an RB56 protein standard. Lane (2) is a mock transfection. In panel B, results are shown for transfection of Saos-2 cells with (1) RB56, (2,3) E2F194-5s, and (4,5) E2F286-5s. In panel C, results are shown for transfection of 5637 cells with (2,3) RB56 wild-type, (4,5) RB56-5s; (6,7) E2F194-5s; (7,8) E2F286-5s. Lane (1) is an RB56 protein standard. In panel D, results are shown for NIH-3T3 transfected (3) RB56, (4) E2F286-5s, (5) E2F194-5s. Lane (1) is an RB56 standard; lane (2) is an RB110 standard.

Figure 12 depicts histogram analyses of flow cytometry of RB-expressing NIH-3T3 cells.

Figure 13, panel A, depicts a comparison of the effects of a CMV-driven recombinant adenovirus (ACN56) with two isolates of a human smooth muscle alpha actin-driven E2F-p56 fusion construct consisting of amino acids 95 through 286 of E2F linked directly and in-frame to p56 (amino acids 379-928 of RB cDNA), vs. a control virus (ACN) in a ³H-thymidine uptake assay in the rat smooth muscle cell line A7R5. Panel (B) depicts the effects of the same constructs in the rat smooth muscle cell line A10.

Figure 14 depicts a comparison of the effects of the viruses described in Fig. 13 in non-muscle cells. Panel (A) depicts results in the breast carcinoma cell line MDA MB468. Panel (B) depicts results in the non-small cell lung cell carcinoma line H358.

Figure 15, top panel, depicts the relative infectivity by adenovirus of different cell lines as judged by the level of β -galactosidase (β -gal) staining following infection with equal amounts of a recombinant adenovirus expressing β -gal driven by a CMV promoter. H358 is non-small lung cell carcinoma cell line; MB468 is a breast carcinoma cell line; A7R5 and A10 are smooth muscle cell lines. The lower portion of the figure depicts the relative levels of p56 protein expressed in the same cells when infected with the recombinant adenovirus ACN56, in which the p56 cDNA is driven by the non-tissue specific CMV promoter.

Figure 16 depicts relative protein levels in cells infected with the smooth muscle alpha actin promoter-driven E2F-p56 fusion construct (ASN286-56). UN denoted uninfected; 50, 100, 250, and 500 refer to multiplicities of infection (MOI).

Figure 17 is a bar graph depicting the ratio of intima to media area (as a measurement of the inhibition of neointima formation) from cross-sections (n=9) of rat carotid arteries which were injured and treated with recombinant adenoviruses expressing either β -gal, RB (ACNRB) or p56 (ACN56), all under the control of the CMV promoter.

Figure 18 is a series of three photographs depicting restenosis in a rat angioplasty model. The panel on the left depicts data from a normal animal; the central panel depicts data from an animal injured and then treated with a β -gal expressing recombinant virus; the panel on the right depicts data from an animal injured and then treated with a recombinant adenovirus expressing p56 (ACN56).

Figure 19 depicts tissue-specificity of the smooth muscle alpha actin promoter, as demonstrated by its selective ability to express the β -gal transgene in muscle cells but not non-muscle cells. The panels on the left compare β -gal expression in the breast cell carcinoma line MB468 infected with either an MOI=1 with a CMV-driven β -gal (ACNBGAL) vs an MOI= 100 with the smooth muscle promoter construct (ASNBGAL). The panels on the right show β -gal expression of the rat smooth muscle cell line A7R5 infected with either an MOI=1 of

ACNBGAL or an MOI=50 of ASNBGAL. Expression from ASNBGAL is seen in the muscle cell line, but is absent in the non-muscle cell line, despite the higher degree of infectivity of the cells.

Figure 20 depicts the ability of recombinant adenovirus expressing RB to transduce rat carotid arteries. recombinant adenovirus-treated arteries (1×10^9 pfu) were harvested two days following balloon injury and infection. Cross sections were fixed and an RB specific antibody was used to detect the presence of RB protein in the tissue. The control virus used was ACN. RB protein staining was evident in the ACNRB treated sample, especially at higher magnifications.

Figure 21 depicts a comparison of the effects of a CMV-driven p56 recombinant adenovirus (ACN56E4) vs a human smooth muscle alpha-actin promoter-driven E2F-p56 fusion construct (ASN286-56) vs control adenoviral constructs containing either the CMV or smooth muscle alpha-actin promoters without a downstream transgene (ACNE3 or ASBE3-2 isolates shown, respectively). Assays were ^3H -thymidine uptake either in a smooth muscle cell line (A7R5) or a non-muscle cell line (MDA-MB468, breast carcinoma). Results demonstrated muscle tissue specificity using the smooth muscle alpha-actin promoter and specific inhibition by both the p56 and E2F-p56 transgenes relative to their respective controls.

DESCRIPTION OF THE PREFERRED EMBODIMENT

The instant invention provides RB fusion constructs including fusion polypeptides and vectors encoding them, and methods for the use of such constructs in the treatment of hyperproliferative diseases. In some preferred embodiments of the invention, an RB polypeptide is fused to an E2F polypeptide. Any E2F species can be used, typically E2F-1, -2, -3, -3, or -5 (see, e.g., Wu et al. Mol. Cell. Biol. 15:2536-2546 (1995); Ivey-Hoyle et al. Mol. Cell. Biol. 13:7802 (1993); Vairo et al. Genes and Dev. 9:869 (1995); Beijersbergen et al. Genes and Dev. 8:2680 (1994)); Ginsberg

et al. Genes and Dev. 8:2665 (1994); Buck et al. Oncogene 11:31 (1995)), more typically E2F-1. Typically, the EF2 polypeptide comprises at least the DNA binding domain of E2F, and may optionally include the cyclin A binding domain, the heterodimerization domain, and/or the transactivation domain. Preferably, the cyclin A binding domain is not functional. The nucleotide and amino acid sequence of E2F referred to herein are those of Genbank HUME2F, shown in Figure 1A and 1B. Nucleic acid, preferably DNA, encoding such an EF2 polypeptide is fused in reading frame to an RB polypeptide. The RB polypeptide can be any RB polypeptide, including conservative amino acid variants, allelic variants, amino acid substitution, deletion, or insertion mutants, or fragments thereof. Preferably, the growth suppression domain, i.e., amino acids residues 379-928, of the RB polypeptide is functional (Hiebert, et al. MCB 13:3384-3391 (1993); Qin, et al. Genes and Dev. 6:953-964 (1992)). In some embodiments, wild-type pRB110 is used. More preferably, a truncated version of RB, RB56, is used. RB56 comprises amino acid residues 379-928 of pRB110 (Hiebert, et al. MCB 13:3384-3391 (1993); Qin, et al. Genes and Dev. 6:953-964 (1992)). In some embodiments, amino acid variants of RB at positions 2, 608, 612, 788, 807, or 811, are used singly or in combination. The variant RB56-5s comprises wild-type RB56 having alanine substitutions at 608, 612, 788, 807, and 811. Numbering of RB amino acids and nucleotides is according to the RB sequence disclosed by Lee, et al. (Nature 329:642-645 (1987)), hereby incorporated by reference in its entirety for all purposes. (Figure 2).

Nucleic acids encoding the polypeptides of the invention can be DNA or RNA. The phrase "nucleic acid sequence encoding" refers to a nucleic acid which directs the expression of a specific protein or peptide. The nucleic acid sequences include both the DNA strand sequence that is transcribed into RNA and the RNA sequence that is translated into protein. The nucleic acid sequences include both the full length nucleic acid sequences as well as non-full length

sequences derived from the full length protein. It is further understood that the sequence includes the degenerate codons of the native sequence or sequences which may be introduced to provide codon preference in a specific host cell.

5 The term "vector" as used herein refers to viral expression systems, autonomous self-replicating circular DNA (plasmids), and includes both expression and nonexpression plasmids. Where a recombinant microorganism or cell culture is described as hosting an "expression vector," this includes
10 both extrachromosomal circular DNA and DNA that has been incorporated into the host chromosome(s). Where a vector is being maintained by a host cell, the vector may either be stably replicated by the cells during mitosis as an autonomous structure, or is incorporated within the host's genome. A
15 vector contains multiple genetic elements positionally and sequentially oriented, i.e., operatively linked with other necessary elements such that nucleic acid in the vector encoding the constructs of the invention can be transcribed, and when necessary, translated in transfected cells.

20 The term "gene" as used herein is intended to refer to a nucleic acid sequence which encodes a polypeptide. This definition includes various sequence polymorphisms, mutations, and/or sequence variants wherein such alterations do not affect the function of the gene product. The term "gene" is
25 intended to include not only coding sequences but also regulatory regions such as promoters, enhancers, and termination regions. The term further includes all introns and other DNA sequences spliced from the mRNA transcript, along with variants resulting from alternative splice sites.

30 The term "plasmid" refers to an autonomous circular DNA molecule capable of replication in a cell, and includes both the expression and nonexpression types. Where a recombinant microorganism or cell culture is described as hosting an "expression plasmid", this includes both
35 extrachromosomal circular DNA molecules and DNA that has been incorporated into the host chromosome(s). Where a plasmid is being maintained by a host cell, the plasmid is either being

stably replicated by the cells during mitosis as an autonomous structure or is incorporated within the host's genome.

The phrase "recombinant protein" or "recombinantly produced protein" refers to a peptide or protein produced using non-native cells that do not have an endogenous copy of DNA able to express the protein. The cells produce the protein because they have been genetically altered by the introduction of the appropriate nucleic acid sequence. The recombinant protein will not be found in association with proteins and other subcellular components normally associated with the cells producing the protein. The terms "protein" and "polypeptide" are used interchangeably herein.

In general, a construct of the invention is provided in an expression vector comprising the following elements linked sequentially at appropriate distances for functional expression: a tissue-specific promoter, an initiation site for transcription, a 3' untranslated region, a 5' mRNA leader sequence, a nucleic acid sequence encoding a polypeptide of the invention, and a polyadenylation signal. Such linkage is termed "operatively linked." Enhancer sequences and other sequences aiding expression and/or secretion can also be included in the expression vector. Additional genes, such as those encoding drug resistance, can be included to allow selection or screening for the presence of the recombinant vector. Such additional genes can include, for example, genes encoding neomycin resistance, multi-drug resistance, thymidine kinase, beta-galactosidase, dihydrofolate reductase (DHFR), and chloramphenicol acetyl transferase.

In the instant invention, tissue-specific expression of the RB constructs of the invention is preferably accomplished by the use of a promoter preferentially used by a tissue of interest. Examples of tissue-specific promoters include the promoter for creatine kinase, which has been used to direct the expression of dystrophin cDNA expression in muscle and cardiac tissue (Cox, et al. Nature 364:725-729 (1993)) and immunoglobulin heavy or light chain promoters for the expression of suicide genes in B cells (Maxwell, et al. Cancer Res. 51:4299-4304 (1991)). An endothelial cell-

specific regulatory region has also been characterized
(Jahroudi, et al. Mol. Cell. Biol. 14:999-1008 (1994)).

Amphotrophic retroviral vectors have been constructed carrying
a herpes simplex virus thymidine kinase gene under the control
of either the albumin or alpha-fetoprotein promoters (Huber,
et al. Proc. Natl. Acad. Sci. U.S.A. 88:8039-8043 (1991)) to
target cells of liver lineage and hepatoma cells,
respectively. Such tissue specific promoters can be used in
retroviral vectors (Hartzoglou, et al. J. Biol. Chem.
265:17285-17293 (1990)) and adenovirus vectors (Friedman, et
al. Mol. Cell. Biol. 6:3791-3797 (1986); Wills et al. Cancer
Gene Therapy 3:191-197 (1995)) and still retain their tissue
specificity.

In the instant invention, a preferred promoter for
tissue-specific expression of exogenous genes is the human
smooth muscle alpha-actin promoter. Reddy, et al. (J. Cell
Biology 265:1683-1687 (1990)) disclosed the isolation and
nucleotide sequence of this promoter, while Nakano, et al.
(Gene 99:285-289 (1991)) disclosed transcriptional regulatory
elements in the 5' upstream and the first intron regions of
the human smooth muscle (aortic type) alpha-actin gene.

Petropoulos, et al. (J. Virol. 66:3391-3397 (1992))
disclosed a comparison of expression of bacterial
chloramphenicol transferase (CAT) operatively linked to either
the chicken skeletal muscle alpha actin promoter or the
cytoplasmic beta-actin promoter. These constructs were
provided in a retroviral vector and used to infect chicken
eggs.

Exemplary tissue-specific expression elements for
the liver include but are not limited to HMG-CoA reductase
promoter (Luskey, Mol. Cell. Biol. 7(5):1881-1893 (1987));
sterol regulatory element 1 (SRE-1; Smith et al. J. Biol.
Chem. 265(4):2306-2310 (1990); phosphoenol pyruvate carboxy
kinase (PEPCK) promoter (Eisenberger et al. Mol. Cell Biol.
12(3):1396-1403 (1992)); human C-reactive protein (CRP)
promoter (Li et al. J. Biol. Chem. 265(7):4136-4142 (1990));

human glucokinase promoter (Tanizawa et al. Mol. Endocrinology 6(7):1070-81 (1992); cholesterol 7-alpha hydroxylase (CYP-7) promoter (Lee et al. J. Biol. Chem. 269(20):14681-9 (1994)); beta-galactosidase alpha-2,6 sialyltransferase promoter (Svensson et al. J. Biol. Chem. 265(34):20863-8 (1990); insulin-like growth factor binding protein (IGFBP-1) promoter (Babajko et al. Biochem Biophys. Res. Comm. 196 (1):480-6 (1993)); aldolase B promoter (Bingle et al. Biochem J. 294(Pt2):473-9 (1993)); human transferrin promoter (Mendelzon et al. Nucl. Acids Res. 18(19):5717-21 (1990); collagen type I promoter (Houglum et al. J. Clin. Invest. 94(2):808-14 (1994)).

Exemplary tissue-specific expression elements for the prostate include but are not limited to the prostatic acid phosphatase (PAP) promoter (Banas et al. Biochim. Biophys. Acta. 1217(2):188-94 (1994); prostatic secretory protein of 94 (PSP 94) promoter (Nolet et al. Biochim. Biophys. ACTA 1098(2):247-9 (1991)); prostate specific antigen complex promoter (Casper et al. J. Steroid Biochem. Mol. Biol. 47 (1-6):127-35 (1993)); human glandular kallikrein gene promoter (hgt-1) (Lilja et al. World J. Urology 11(4):188-91 (1993)).

Exemplary tissue-specific expression elements for gastric tissue include but are not limited to the human H⁺/K⁺-ATPase alpha subunit promoter (Tanura et al. FEBS Letters 298:(2-3):137-41 (1992)).

Exemplary tissue-specific expression elements for the pancreas include but are not limited to pancreatitis associated protein promoter (PAP) (Dusetti et al. J. Biol. Chem. 268(19):14470-5 (1993)); elastase 1 transcriptional enhancer (Kruse et al. Genes and Development 7(5):774-86 (1993)); pancreas specific amylase and elastase enhancer promoter (Wu et al. Mol. Cell. Biol. 11(9):4423-30 (1991); Keller et al. Genes & Dev. 4(8):1316-21 (1990)); pancreatic cholesterol esterase gene promoter (Fontaine et al. Biochemistry 30(28):7008-14 (1991)).

Exemplary tissue-specific expression elements for the endometrium include but are not limited to the uteroglobin promoter (Helftenbein et al. Annal. NY Acad. Sci. 622:69-79 (1991)).

5 Exemplary tissue-specific expression elements for adrenal cells include but are not limited to cholesterol side-chain cleavage (SCC) promoter (Rice et al. J. Biol. Chem. 265:11713-20 (1990)).

10 Exemplary tissue-specific expression elements for the general nervous system include but are not limited to gamma-gamma enolase (neuron-specific enolase, NSE) promoter (Forss-Petter et al. Neuron 5(2):187-97 (1990)).

15 Exemplary tissue-specific expression elements for the brain include but are not limited to the neurofilament heavy chain (NF-H) promoter (Schwartz et al. J. Biol. Chem. 269(18):13444-50 (1994)).

20 Exemplary tissue-specific expression elements for lymphocytes include but are not limited to the human CGL-1/granzyme B promoter (Hanson et al. J. Biol. Chem. 266 (36):24433-8 (1991)); the terminal deoxy transferase (TdT), lambda 5, VpreB, and lck (lymphocyte specific tyrosine protein kinase p56lck) promoter (Lo et al. Mol. Cell. Biol. 11(10):5229-43 (1991)); the humans CD2 promoter and its 3'transcriptional enhancer (Lake et al. EMBO J. 9(10):3129-36 (1990)), and the human NK and T cell specific activation (NKG5) promoter (Houchins et al. Immunogenetics 37(2):102-7 (1993)).

30 Exemplary tissue-specific expression elements for the colon include but are not limited to pp60c-src tyrosine kinase promoter (Talamonti et al. J. Clin. Invest 91(1):53-60 (1993)); organ-specific neoantigens (OSNs), mw 40kDa (p40) promoter (Ilantzis et al. Microbiol. Immunol. 37(2):119-28 (1993)); colon specific antigen-P promoter (Sharkey et al. Cancer 73(3 supp.) 864-77 (1994)).

35 Exemplary tissue-specific expression elements for breast cells include but are not limited to the human alpha-

lactalbumin promoter (Thean et al. British J. Cancer,
61(5):773-5 (1990)).

5 Other elements aiding specificity of expression in a
tissue of interest can include secretion leader sequences,
enhancers, nuclear localization signals, endosmolytic
peptides, etc. Preferably, these elements are derived from
the tissue of interest to aid specificity.

10 Techniques for nucleic acid manipulation of the
nucleic acid sequences of the invention such as subcloning
nucleic acid sequences encoding polypeptides into expression
vectors, labelling probes, DNA hybridization, and the like are
described generally in Sambrook et al., Molecular Cloning - A
Laboratory Manual (2nd Ed.), Vol. 1-3, Cold Spring Harbor
Laboratory, Cold Spring Harbor, New York, (1989), which is
15 incorporated herein by reference. This manual is hereinafter
referred to as "Sambrook et al."

20 Once DNA encoding a sequence of interest is isolated
and cloned, one can express the encoded proteins in a variety
of recombinantly engineered cells. It is expected that those
of skill in the art are knowledgeable in the numerous
expression systems available for expression of DNA encoding.
No attempt to describe in detail the various methods known for
the expression of proteins in prokaryotes or eukaryotes is
made here.

25 In brief summary, the expression of natural or
synthetic nucleic acids encoding a sequence of interest will
typically be achieved by operably linking the DNA or cDNA to a
promoter (which is either constitutive or inducible), followed
by incorporation into an expression vector. The vectors can
30 be suitable for replication and integration in either
prokaryotes or eukaryotes. Typical expression vectors contain
transcription and translation terminators, initiation
sequences, and promoters useful for regulation of the
expression of polynucleotide sequence of interest. To obtain
35 high level expression of a cloned gene, it is desirable to
construct expression plasmids which contain, at the minimum, a
strong promoter to direct transcription, a ribosome binding
site for translational initiation, and a

transcription/translation terminator. The expression vectors may also comprise generic expression cassettes containing at least one independent terminator sequence, sequences permitting replication of the plasmid in both eukaryotes and prokaryotes, *i.e.*, shuttle vectors, and selection markers for both prokaryotic and eukaryotic systems. See Sambrook et al.

The E2F-RB fusion constructs of the invention can be introduced into the tissue of interest *in vivo* or *ex vivo* by a variety of methods. In some embodiments of the invention, the nucleic acid, preferably DNA, is introduced to cells by such methods as microinjection, calcium phosphate precipitation, liposome fusion, or biolistics. In further embodiments, the DNA is taken up directly by the tissue of interest. In other embodiments, the constructs are packaged into a viral vector system to facilitate introduction into cells.

Viral vector systems useful in the practice of the instant invention include adenovirus, herpesvirus, adeno-associated virus, minute virus of mice (MVM), HIV, sindbis virus, and retroviruses such as Rous sarcoma virus, and MoMLV. Typically, the constructs of the instant invention are inserted into such vectors to allow packaging of the E2F-RB expression construct, typically with accompanying viral DNA, infection of a sensitive host cell, and expression of the E2F-RB gene. A particularly advantageous vector is the adenovirus vector disclosed in Wills, et al. Human Gene Therapy 5:1079-1088 (1994).

In still other embodiments of the invention, the recombinant DNA constructs of the invention are conjugated to a cell receptor ligand for facilitated uptake (*e.g.*, invagination of coated pits and internalization of the endosome) through a DNA linking moiety (Wu, et al. J. Biol. Chem. 263:14621-14624 (1988); WO 92/06180). For example, the DNA constructs of the invention can be linked through a polylysine moiety to asialo-oromucoid, which is a ligand for the asialoglycoprotein receptor of hepatocytes.

Similarly, viral envelopes used for packaging the constructs of the invention can be modified by the addition of

receptor ligands or antibodies specific for a receptor to permit receptor-mediated endocytosis into specific cells (e.g., WO 93/20221, WO 93/14188; WO 94/06923). In some embodiments of the invention, the DNA constructs of the invention are linked to viral proteins, such as adenovirus particles, to facilitate endocytosis (Curiel, et al. Proc. Natl. Acad. Sci. U.S.A. 88:8850-8854 (1991)). In other embodiments, molecular conjugates of the instant invention can include microtubule inhibitors (WO 94/06922); synthetic peptides mimicking influenza virus hemagglutinin (Plank, et al. J. Biol. Chem. 269:12918-12924 (1994)); and nuclear localization signals such as SV40 T antigen (WO 93/19768).

In some embodiments of the invention, the RB polypeptides of the invention are administered directly to a patient in need of treatment. A "therapeutically effective" dose is a dose of polypeptide sufficient to prevent or reduce severity of a hyperproliferative disorder. As used herein, the term "hyperproliferative cells" includes but is not limited to cells having the capacity for autonomous growth, i.e., existing and reproducing independently of normal regulatory mechanisms. Hyperproliferative diseases may be categorized as pathologic, i.e., deviating from normal cells, characterizing for constituting disease, or may be categorized as non-pathologic, i.e., deviation from normal but not associated with a disease state. Pathologic hyperproliferative cells are characteristic of the following disease states: restenosis, diabetic retinopathy, thyroid hyperplasia, Grave's disease, psoriasis, benign prostatic hypertrophy, Li-Fraumeni syndrome including breast cancer, sarcomas and other neoplasms, bladder cancer, colon cancer, lung cancer, various leukemias and lymphomas. Examples of non-pathological hyperproliferative cells are found, for instance, in mammary ductal epithelial cells during development of lactation and also in cells associated with wound repair. Pathological hyperproliferative cells characteristically exhibit loss of contact inhibition and a decline in their ability to selectively adhere which implies a further breakdown in intercellular communication. These

changes include stimulation to divide and the ability to secrete proteolytic enzymes.

The constructs of the invention are useful in the therapy of various cancers and other conditions in which the administration of RB is advantageous, including but not limited to peripheral vascular diseases and diabetic retinopathy. Although any tissue can be targeted for which some tissue-specific expression element, such as a promoter, can be identified, of particular interest is the tissue-specific administration of an RB construct for hyperproliferative disorders such as restenosis, for which the smooth muscle actin promoter is preferable.

The compositions of the invention will be formulated for administration by manners known in the art acceptable for administration to a mammalian subject, preferably a human. In some embodiments of the invention, the compositions of the invention can be administered directly into a tissue by injection or into a blood vessel supplying the tissue of interest. In further embodiments of the invention the compositions of the invention are administered "locoregionally", i.e., intravesically, intralesionally, and/or topically. In other embodiments of the invention, the compositions of the invention are administered systemically by injection, inhalation, suppository, transdermal delivery, etc. In further embodiments of the invention, the compositions are administered through catheters or other devices to allow access to a remote tissue of interest, such as an internal organ. The compositions of the invention can also be administered in depot type devices, implants, or encapsulated formulations to allow slow or sustained release of the compositions.

The invention provides compositions for administration which comprise a solution of the compositions of the invention dissolved or suspended in an acceptable carrier, preferably an aqueous carrier. A variety of aqueous carriers may be used, e.g., water, buffered water, 0.8% saline, 0.3% glycine, hyaluronic acid and the like. These compositions may be sterilized by conventional, well known

sterilization techniques, or may be sterile filtered. The resulting aqueous solutions may be packaged for use as is, or lyophilized, the lyophilized preparation being combined with a sterile solution prior to administration. The compositions may contain pharmaceutically acceptable auxiliary substances as required to approximate physiological conditions, such as pH adjusting and buffering agents, tonicity adjusting agents, wetting agents and the like, for example, sodium acetate, sodium lactate, sodium chloride, potassium chloride, calcium chloride, sorbitan monolaurate, triethanolamine oleate, etc.

The concentration of the compositions of the invention in the pharmaceutical formulations can vary widely, i.e., from less than about 0.1%, usually at or at least about 2% to as much as 20% to 50% or more by weight, and will be selected primarily by fluid volumes, viscosities, etc., in accordance with the particular mode of administration selected.

The compositions of the invention may also be administered via liposomes. Liposomes include emulsions, foams, micelles, insoluble monolayers, liquid crystals, phospholipid dispersions, lamellar layers and the like. In these preparations the composition of the invention to be delivered is incorporated as part of a liposome, alone or in conjunction with a molecule which binds to a desired target, such as antibody, or with other therapeutic or immunogenic compositions. Thus, liposomes either filled or decorated with a desired composition of the invention of the invention can delivered systemically, or can be directed to a tissue of interest, where the liposomes then deliver the selected therapeutic/immunogenic peptide compositions.

Liposomes for use in the invention are formed from standard vesicle-forming lipids, which generally include neutral and negatively charged phospholipids and a sterol, such as cholesterol. The selection of lipids is generally guided by consideration of, e.g., liposome size, acid lability and stability of the liposomes in the blood stream. A variety of methods are available for preparing liposomes, as described in, e.g., Szoka et al. Ann. Rev. Biophys. Bioeng. 9:467

(1980), U.S. Patent Nos. 4,235,871, 4,501,728, 4,837,028, and 5,019,369, incorporated herein by reference.

5 A liposome suspension containing a composition of the invention may be administered intravenously, locally, topically, etc. in a dose which varies according to, inter alia, the manner of administration, the composition of the invention being delivered, and the stage of the disease being treated.

10 For solid compositions, conventional nontoxic solid carriers may be used which include, for example, pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharin, talcum, cellulose, glucose, sucrose, magnesium carbonate, and the like. For oral
15 administration, a pharmaceutically acceptable nontoxic composition is formed by incorporating any of the normally employed excipients, such as those carriers previously listed, and generally 10-95% of active ingredient, that is, one or more compositions of the invention of the invention, and more preferably at a concentration of 25%-75%.

20 For aerosol administration, the compositions of the invention are preferably supplied in finely divided form along with a surfactant and propellant. Typical percentages of compositions of the invention are 0.01%-20% by weight, preferably 1%-10%. The surfactant must, of course, be
25 nontoxic, and preferably soluble in the propellant. Representative of such agents are the esters or partial esters of fatty acids containing from 6 to 22 carbon atoms, such as caproic, octanoic, lauric, palmitic, stearic, linoleic, linolenic, olesteric and oleic acids with an aliphatic
30 polyhydric alcohol or its cyclic anhydride. Mixed esters, such as mixed or natural glycerides may be employed. The surfactant may constitute 0.1%-20% by weight of the composition, preferably 0.25-5%. The balance of the composition is ordinarily propellant. A carrier can also be
35 included, as desired, as with, e.g., lecithin for intranasal delivery.

The constructs of the invention can additionally be delivered in a depot-type system, an encapsulated form, or an

implant by techniques well-known in the art. Similarly, the constructs can be delivered via a pump to a tissue of interest.

In some embodiments of the invention, the compositions of the invention are administered ex vivo to cells or tissues explanted from a patient, then returned to the patient. Examples of ex vivo administration of gene therapy constructs include Arteaga et al. Cancer Research 56(5):1098-1103 (1996); Nolita et al. Proc Natl. Acad. Sci. USA 93(6):2414-9 (1996); Koc et al. Seminars in Oncology 23(1):46-65 (1996); Raper et al. Annals of Surgery 223(2):116-26 (1996); Dalesandro et al. J. Thorac. Cardi. Surg. 11(2):416-22 (1996); and Makarov et al. Proc. Natl. Acad. Sci. USA 93(1):402-6 (1996).

In some embodiments of the invention, the constructs of the invention are administered to a cardiac artery after balloon angioplasty to prevent or reduce the severity of restenosis. The constructs of the invention can be used to coat the device used for angioplasty (see, for example, Willart, et al. Circulation 89:2190-2197 (1994); French, et al. Circulation 90:2402-2413 (1995)). In further embodiments, the fusion polypeptides of the invention can be used in the same manner.

The following examples are included for illustrative purposes and should not be considered to limit the present invention.

EXAMPLES

Example I

E2F-RB Fusions

A. Introduction

In this example, expression plasmids which encode different segments of E2F fused to RB56 polypeptide were constructed. RB56 is a subfragment of full length RB which contains the "pocket" domains necessary for growth suppression (Hiebert, et al. MCB 13:3384-3391 (1993); Qin, et al. Genes

and Dev. 6:953-964 (1992)). E2F194 contains E2F amino acids 95-194. This fragment contains only the DNA binding domain of E2F. E2F286 contains the DNA binding domain and the DP-1 heterodimerization domain. Both E2F fragments lack the N-terminal cyclin A-kinase binding domain, which appears to down-regulate the DNA binding activity of E2F (Krek et al. Cell 83:1149-1158 (1995); Krek et al. Cell 78:161-172 (1994)).

B. Construction of Vectors

Plasmid pCTM contains a CMV promoter, a tripartite adenovirus leader flanked by T7 and SP6 promoters, and a multiple cloning site with a bovine growth hormone (BGH) polyadenylation site and a SV-40 poly adenylation site downstream. A diagrammatic representation of pCTM is provided in Figure 3. The DNA sequence for pCTM is provided in Figure 4.

pCTMI was constructed from pCTM by digesting pCTM with Xho I and Not I and subcloning a 180 bp intron XhoI-Not I fragment from a pCMV- β -gal vector (Clonetech). A diagrammatic representation of pCTMI is provided in Figure 5. The DNA sequence is provided in Figure 6.

pCTMIE was constructed by amplifying the SV40 enhancer from SV40 viral DNA in a polymerase chain reaction. The amplified product was digested with BglII and inserted into BamHI-digested pCMTI and ligated in the presence of BamHI. The plasmid is depicted diagrammatically in Figure 7. The DNA sequence is provided in Figure 8.

pCTM-RB was prepared as follows. A 3.2 KB Xba I - Cla I fragment of pETRBc (Huang et al. Nature 350:160-162 (1991)) containing the full length human RB cDNA was ligated to Xba I-Cla I digested pCTM. pCTM-RB56 was prepared by ligating the digested pCTM to a 1.7 KB Xba I -Cla I fragment containing the coding sequence for RB56. pCTMI-RB, pCTMIE-RB, pCTMI-RB56(amino acids 381-928) and pCTMIE-RB56(amino acids 381-928) were all constructed by the same methods.

C. RB-E2F fusion Constructs

Figure 9 depicts the fusion constructs used in these studies. These E2F constructs commenced at amino acid 95 and lacked part of the cyclin A binding domain. E2F437 contained the DNA binding domain (black), heterodimerization domain (white) and transactivation domain (stippled). E2F194 contained solely the DNA binding domain. E2F286 contained the DNA binding domain and DP-1 heterodimerization domain. RB56-5s refers to an RB variant having alanine substitutions at amino acid residues 606, 612, 788, 807 and 811. In E2F194-RB56-5s and E2F286-RB56-5s, the E2F fragments were fused in frame to codon 379 of RB-5s. RB56-C706F contained an inactivating point mutation (Kaye et al. Proc. Natl. Acad. Sci. U.S.A. 87:6922-6926 (1990)).

pCMV-E2F194 and pCMV-E2F437 were constructed as follows. DNA encoding amino acids 95-194 of E2F (containing the DNA binding domain) or amino acids 95-437 was amplified in a polymerase chain reaction, digested with HindII, and ligated into SmaI/HindII digested pCMV-RB56 vectors. pCMVE2F286 was constructed by digesting pCMV-E2F437 with AflIII, treating the ends with DNA pol I (Klenow fragment) and religating in the presence of AflIII. The blunt end ligation created a stop codon at position 287. pCMV-E2F286-5s was constructed by ligating AflIII (blunt)/HindIII digested pE2F437 to a Sal I (blunt)-HindIII fragment containing the RB56-5s coding sequence. pCTMIE-E2F194-5s and pCTMIE-E2F286-RB5s were constructed by ligating EcoRI-EcoRV digested pCTMIE (4.2 KB) to HindIII (blunt)-EcoRI fragments from either pCMV-E2F194-RB5s or pCMV-E2F286-RB5s.

D. Promoter Repression

To measure the effect of the E2F-RB fusion proteins, cervical carcinoma cell line C33A (ATCC # HTB-31) was transfected with equivalent amounts of E2F194-RB56 or E2F RB56 with an E2-CAT reporter plasmid (See, e.g., Weintraub et al. Nature 358:259-261 (1992)).

In the C33A assay, 250,000 C33A cells were seeded into each of well of 6-well tissue culture plates and allowed to adhere overnight. 5 μ g each of pCMV-RB56, pCMV-E2F RB56,

or pCMV-E2F plasmid were cotransfected (calcium phosphate method, MBS transfection kit, Stratagene) with 5 μ g of indicated reporter construct E2-CAT or SVCAT) and 2.5 μ g β -gal plasmid (pCMV- β , Clontech) per well into duplicate wells.

5 Cells were harvested 72 hour after transfection and extracts were prepared.

In the 5637 assay, 250,000 5637 cells were seeded as described above. 1 μ g each of RB or E2F-RB fusion plasmid, E2-CAT or SV-CAT reporter plasmid and pCMV- β -galactosidase

10 were cotransfected using the lipofectin reagent (BRL, Bethesda, Maryland) according to the manufacturer's instructions.

CAT assays were performed using either 20 μ L (C33A) or 50 μ L (5637) of cell extract (Gorman et al. Mol. Cell. Biol. 2:1044 (1982)). TLCs were analyzed on a Phosphorimager SF (Molecular Dynamics). CAT activities were normalized for transfection efficiency according to β -galactosidase activities of each extract. β -galactosidase activities of extracts were assayed as described by Rosenthal et al. (Meth. Enzym. 152:704 (1987)).

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The results of these studies were as follows. Transfection of the E2-CAT reporter alone or in the presence of the nonfunctional control RB56-H209 mutant yielded relatively high CAT activity. Cotransfection of wild-type

25 RB56 or the variant RB56-5s resulted in a 10 to 12 fold repression of CAT activity, indicating that RB56 or RB56-5s are both capable of efficiently repressing E2F-dependent transcription. E2F194-RB5s and E2F286-RB5s repressed transcription approximately 50 fold. Transcriptional

30 repression required both the RB56 and the E2F components of the fusion proteins, as expression of E2F194 and E2F286 did not mediate transcriptional repression. No repression of SV40-CAT transcription occurred with E2F-RB constructs, thus demonstrating the specificity of the transcriptional

35 repression by E2FRB for the E2 promoter. These results are depicted diagrammatically in Figure 10.

E. Cell cycle arrest

The ability of E2F-RB fusion polypeptides to cause G1 arrest in Saos-2 (RB-/- cells) (ATCC # HTB-85) and C33A cells was investigated. Previous studies have shown that RB-mediated E2 promoter repression and G1 arrest are linked in Saos-2 cells but dissociated in C33A (RBmut) cells (Xu, et al. PNAS 92:1357-1361 (1992)). Cells were washed in PBS and were fixed in 1 mL -20°C 70% ethanol for 30 minutes. Cells were collected by centrifugation and resuspended in 0.5 mL 2% serum containing 10 µg/ml RNase A and incubated for 30 minutes at 37°C 0.5 mL of PBS containing propidium iodide (100 µg/ml) was added to each sample, mixed and cells were filtered through a FACS tube capstrainer. FACS analysis was performed on a FACS-Scan (Becton-Dickenson) using doublet discrimination. 5,000-10,000 CD20+ events were analyzed. Percent of cells in G₀/G₁, S, and G₂/M was determined using Modfit modeling software.

The results of this experiment were as follows. Both full length RB110 and the truncated version RB56, but not the control mutant RB-H209, caused G₁ arrest in Saos-2 cells (Table 1). Similarly, the RB56-5s, E2F-194-RB56-5s and E2F286-RB56-5s all were capable of arresting cells in G₀/G₁. Transfection of the DNA binding domain, E2F194, did not block S-phase entry in Saos-2 as previously described for rodent cells (Dobrowolski, et al. Oncogene 9:2605-2612 (1994)). In contrast, RB110, RB56, and E2F-RB fusion proteins were not capable of arresting C33A cell lines indicating that the transcriptional repression observed in these cells does not translate into G₁ arrest.

The ability of the E2F-RB fusion proteins to arrest 5637 cells was also investigated (Table 2). RB56 and RB56-5s both efficiently arrested cells in G₀/G₁ (approximately 90% of cells in G₀-G₁), whereas E2F194-RB56-5s and E2F286-RB56-5s are slightly less efficient (about 80% of cells in G₀/G₁) at promoting G₀/G₁ arrest. Without being limited to any one theory, the less efficient arrest of both Saos-2 and 5637 cells by the E2F-RB fusion proteins appears due to the lower levels of steady-state protein produced in these cells (Figure 11, panels b and c).

Table 1: Cell Cycle Regulation by RB and E2F-RB fusion proteins in RBneg cells

	% Cells		
	CD20 ⁺ G ₀ /G ₁	G ₂ /M	S-phase
H209	52.1	27.1	20.8
p56RB	78.8	14.2	7.0
p110RB	70.9	14.9	14.8
p56RB-5s	84.8	13.2	2.0
p56RB-p5	81.3	11.5	7.3
E2F-194-5s	77.8	14.9	7.3
E2F-286-5s	72.2	15.0	12.8
E2F-194	49.9	28.0	22.1

Table 2: Growth Suppression of 5637 Bladder Cells by RB and E2F-RB fusion proteins

5637/CD20 ⁺	% Cells		
	G ₀ /G ₁	S	G ₂ M
CD20	59.7	16.9	20.6
RB56-C706F	57.4	16.9	24.3
RB56WT	90.7	4.12	4.88
RB56-5s	89.91	3.51	6.1
E2F1 94-5s	80.1	1.31	0
E2F-286-5s	79.21	8.1	0

F. Activity of Fusion Proteins in Functional RB Background

The activity of the E2F-RB fusion proteins in a cellular background containing functional RB was then determined. NIH-3T3 cells were transfected with RB56 or E2F-RB56 fusions and stained with anti-RB monoclonal antibody 3C8 (Wen et al. *J. Immuno. Meth.* 169:231-240 (1994)). FACS analysis was performed of the RB expressing cells. The

results are shown in Figure 12. The non-gated population (g) shows the characteristic cell cycle distribution for NIH-3T3 cells (60% G₀, 28% S, 10% G₂/M). In contrast, in cells transfected with RB56 (a,b) or E2F-RB fusion proteins (c-f), greater than 90% of the RB-expressing cells were arrested in G₀/G₁. These data demonstrate that the ability of RB and E2F-RB56 fusions to arrest cells in G₀/G₁ is not limited to RB negative tumor cells. The relative levels of protein expressed in transfected NIH-3T3 cells was also investigated. RB110 was not expressed efficiently in these cells.

Thus, these data demonstrate that E2F-RB fusion proteins are more efficient transcriptional repressors than either pRB or RB56 alone, and that RB can repress transcription by remaining bound to E2F rather than directly blocking the transactivation domain of E2F. These data support the use of E2F-RB fusions as RB agonists in both RB+ cells and in RB negative or RB mutant cells.

Example II.

Tissue-Specific Expression of E2F-RB Fusions

A. Construction of Recombinant Adenovirus:

In this experiment, recombinant adenoviruses comprising an RB polypeptide under the control of a CMV or smooth muscle alpha actin promoter were generated.

The smooth muscle α -actin promoter (bases -670 through +5, Reddy et al. "Structure of the Human Smooth Muscle α -Actin Gene." J. Biol. Chem. 265:1683-1687 (1990), Nakano, et al. "Transcriptional Regulatory Elements In The 5' Upstream and First Intron Regions of The Human Smooth Muscle (aortic type) α -Actin-Encoding Gene." Gene 99:285-289 (1991) was isolated by PCR from a genomic library with 5' Xho I and Avr II and 3' Xba I, Cla I and Hind III restriction sites added for cloning purposes. The fragment was subcloned as an Xho 1, Hind III fragment into a plasmid for sequencing to verify base composition. A fusion construct 286-56 containing the DNA and heterodimerization domain of E2F-1 (bases 95-286) linked to p56 (amino acids 379-928 of full length RB) was subcloned as

an Xba I, Cla I fragment directly downstream of the smooth muscle α -actin promoter, and this expression cassette was digested out and cloned into the plasmid pAd/ITR/IX- as an Xba I to AvrII, and Cla I fragment to create the plasmid pASN286-56. This plasmid consisted of the adenovirus type 5 inverted terminal repeat (ITR), packaging signals and Ela enhancer, followed by the human smooth muscle α -actin promoter and 286-56 cassette, and then Ad 2 sequence 4021-10462 (which contains the E1b/protein IX poly A signal) in a pBR322 background.

Recombinant adenovirus was produced by standard procedures. The plasmid pASN286-56 was linearized with Ngo MI and co-transfected into 293 cells with the large fragment of Cla I digested rAd34 which has deletions in both the E3 and E4 regions of adenovirus type 5. Ad34 was a serotype 5 derivative with a 1.9 KB deletion in early region 3 resulting from deletion of the Xba I restriction fragment extending from Ad5 coordinates 28593 to 30470 and a 1.4 KB deletion of early region 4 resulting from a Taq I fragment of E4 (coordinates 33055-35573) being replaced with a cDNA containing E4 ORF 6 and 6/7.

Recombinant adenovirus produced by homologous recombination was isolated and identified by restriction digest analysis and further purified by limiting dilution. Additional control recombinant adenoviruses are described elsewhere and include the control virus ACN (CMV promoter, Wills, et al. "Gene Therapy For Hepatocellular Carcinoma: Chemosensitivity Conferred By Adenovirus-Mediated Transfer of The HSV-1 Thymidine Kinase Gene." Cancer Gene Therapy 2:191-197 (1995)), and ACN56 (RB expressed under control of a CMV promoter).

ACN56 was prepared as follows. A plasmid containing p56 cDNA was constructed by replacing the p53 cDNA from the plasmid ACNP53 (Wills et al. Human Gene Therapy 5:1079-1088 (1994)) with a 1.7 KB Xba I- BamHI fragment isolated from plasmid pET 9a-Rb56 (Antelman et al. Oncogene 10:697-704 (1995)) which contains p56 cDNA. The resulting plasmid contained amino acids 381-928 of p56, the Ad5 inverted terminal repeat, viral packaging signals and Ela enhancer,

followed by the human cytomegalovirus immediate early promoter (CMV) and Ad 2 tripartite leader cDNA to drive p56 expression. The p56 cDNA was followed by Ad 2 sequence 4021-10462 in a pBR322 background. This plasmid was linearized with EcoRI and cotransfected with the large fragment of bsp 106 digested DL327 (E3 deleted; Thimmappaaya et al. Cell 31:543-551 (1982)) or h5ile4 (E4 deleted; Hemstrom et al. J. Virol. 62:3258-3264 (1988)). Recombinant viruses were further purified by limiting dilution.

B. Cellular Proliferation

In this experiment, cell lines were infected in culture with recombinant adenovirus RB constructs to ascertain the relative expression of the RB polypeptide and the effect on cell proliferation.

For H358 (ATCC # Crl 5807) and MDA-MB468 (ATCC # HTB 132, breast adenocarcinoma) cells, 5,000 cell/well were plated in normal growth media in a 96 well microtiter plate (Costar) and allowed to incubate overnight at 37°C, 7% CO₂. Viruses were serially diluted in growth media and used to infect cells at the indicated doses for 48 hours. At this point, ³H-thymidine was added (Amersham, 0.5 µCi/well) and the cells were incubated at 37°C for another 3 hours prior to harvest. Both A7r5 (ATCC CRL1444, rat smooth muscle) and A10 (ATCC CRL 1476, rat smooth muscle) cells were seeded at 3,000 cells/well in either DME + 0.5% FCS or DME + 20% FCS respectively. Virus was serially diluted in the seeding media and used to infect the cells at the doses indicated in the Figures. The infection and labelling procedure were the same for A10 cells as with the H358 and MDA-MB468 cells except that 2 µCi/well of label was used. The A7r5 cells were not infected with virus until 48 hours after seeding. Forty eight hours after infection, the serum concentration was raised to 10% FCS and 2 µCi/well of ³H-thymidine was added and incubation continued for an additional 3 hours prior to harvest. All cells were harvested by aspirating media from the wells, trypsinization of the cells, and harvesting using a 96 well GF/C filter with

a Packard Top count cell harvester. Results are plotted as the mean percentage (+/- SD) of media treated control proliferation versus dose of virus in Figures 13 and 14.

Thus, Figure 13 depicts a comparison of the effects of adenovirus p56 constructs on muscle cells A10 and A7R5 cells. The CMV-driven p56 (ACN 56) virus inhibited A10 growth to approximately the same extent as the actin promoter-driven E2F-fusion constructs (ASN586-56 #25,26). In Figure 14, the effects of adenovirus constructs on inhibition of a breast cancer cell line, MDA Mβ468 and a non-small cell lung carcinoma cell line, H358, are depicted. In these experiments, actin promoter-driven E2F-p56 was ineffective, while the CMV promoter-driven p56 was effective in inhibiting growth of non-smooth muscle cells.

To determine whether the non-smooth muscle cells were more infectable with adenovirus than the smooth muscle cell lines used, the four cell lines, H358, MB468, A7R5, and A10 were infected at an MOI of 5 with an adenovirus expressing β-galactosidase (ACβGL; Wills, et al. Human Gene Therapy 5:1079-1088 (1994)) and degree of β-gal staining was examined. As shown in Figure 15 (top), the non-smooth muscle cell lines were significantly more infectable than the smooth muscle cell lines. In a further test, cells were infected at higher multiplicities of infection (50, 100, 250, 500) with ACN56 and the amount of p56 present in the infected cells detected by autoradiography. As can be seen in Figure 15 (bottom), the non-muscle cell lines had significantly more p56 present, since as a result of their greater infectivity, infected cells have a greater viral load and thus more copies of the p56 template driven by the non-tissue specific CMV promoter.

In a further experiment, the specificity of the actin smooth muscle promoter for smooth muscle tissue was ascertained. In this experiment, β-gal expression levels in cells infected with β-gal constructs driven with different promoters were measured. As can be seen in Figure 19, despite the lower infectivity of the smooth muscle cells, expression was only evident in these cells using the smooth muscle alpha actin promoter.

Figure 21 depicts a comparison of the effects of a CMV driven p56 recombinant adenovirus (ACN56E4) vs a human smooth muscle alpha-actin promoter driven E2F-p56 fusion construct (ASN286-56) vs control adenoviral construct containing either the CMV or smooth muscle alpha-actin promoters without a downstream transgene (ACNE3 or ASBE3-2 isolates shown, respectively). Assays were 3H-thymidine uptake either in a smooth muscle cell line (A7R5) or a non-muscle cell line (MDA-MB468, breast carcinoma). Results demonstrated muscle tissue specificity using the smooth muscle alpha-actin promoter and specific inhibition of both the p56 and E2F-p56 transgenes relative to their respective controls.

C. Inhibition of Restenosis

The model of balloon injury was based on that described by Clowes, et al. (Clowes, Lab. Invest. 49:327-333 (1983)). Male Sprague-Dawley rats weighing 400-500g were anesthetized with an intraperitoneal injection of sodium pentobarbital (45 mg/kg. Abbot Laboratories, North Chicago, Illinois). The bifurcation of the left common carotid artery was exposed through a midline incision and the left common, internal, and external carotid arteries were temporarily ligated. A 2F embolectomy catheter (Baxter Edwards Healthcare Corp., Irvine, CA) was introduced into the external carotid and advanced to the distal ligation of the common carotid. The balloon was inflated with saline and drawn towards the arteriotomy site 3 times to produce a distending, deendothelializing injury. the catheter was then withdrawn. Adenovirus (1×10^9 pfu of Ad-RB (ACNRb) or Ad-p56 (ACN56) in a volume of $10\mu\text{l}$ diluted to $100\mu\text{l}$ with 15% (wt/vol) Poloxamer 407 (BASF, Parsippany, N.J.) or Ad- β -Gal (1×10^9 pfu, diluted as above) was injected via a cannula, inserted just proximal to the carotid bifurcation into a temporarily isolated segment of the artery. The adenovirus solution was incubated for 20 minutes after which the viral infusion was withdrawn and the cannula removed. The proximal external carotid artery was then ligated and blood flow was restored to the common carotid artery by release of the ligatures. The experimental protocol

was approved by the Institutional Animal Care and Use Committee and complied with the "Guide for the Care and Use of Laboratory Animals." (NIH Publication No. 86-23, revised 1985).

5 Rats were sacrificed at 14 days following treatment with an intraperitoneal injection of pentobarbital (100 mg/kg.). The initially balloon injured segment of the left common carotid artery, from the proximal edge of the omohyoid muscle to the carotid bifurcation, was perfused with saline and dissected free of the surrounding tissue. The tissue was
10 fixed in 100% methanol until imbedded in paraffin. Several 4- μ m sections were cut from each tissue specimen. One section from each specimen was stained with hematoxylin and eosin and another with Richardson's combination elastic-trichrome stain
15 conventional light microscopic analysis.

Histological images of cross sections of hematoxylin and eosin or elastic-trichrome stained arterial sections were projected onto a digitizing board (Summagraphics) and the intimal, medial and luminal areas were measured by
20 quantitative morphometric analysis using a computerized sketching program (MACMEASURE, version 1.9, National Institute of Mental Health).

Results were expressed as the mean \pm S.E.M. Differences between groups were analyzed using an unpaired
25 two-tailed Student's t test. Statistical significance was assumed when the probability of a null effect was <0.05 .

Results are shown in Figures 17 and 18. In Figure 17, the relative inhibition of neointima formation is depicted graphically, demonstrating the ability of p56 and RB to
30 inhibit neointima formation. Figure 18 provides photographic evidence of the dramatic reduction of neointima in the presence of p56.

Adenovirus-treated carotid arteries were harvested from rats at 2 days following balloon injury and infections.
35 Tissue was fixed in phosphate-buffered formalin until embedded in paraffin. Tissue was cut into 4 μ m cross-sections and dewaxed through xylene and graded alcohols. Endogenous peroxidase was quenched with 1% hydrogen peroxide for 30

minutes. Antigen retrieval was performed in 10mM sodium citrate buffer, pH 6.0 at 95°C for 10 minutes. A monoclonal anti-RB antibody (AB-5, Oncogene Sciences, Uniondale, New York) was applied 10µg/ml in PBS in a humid chamber at 4°C for 24 hours. Secondary antibody was applied from the Unitect Mouse Immunohistochemistry Kit (Oncogene Sciences, Uniondale, New York) according to the manufacturer's instructions. The antibody complexes were visualized using 3,3'-diaminobenzidine (DAB, Vector Laboratories, Burlingame, CA). Slides were thin counterstained with hematoxylin and mounted. The results are depicted in Figure 20.

All references cited herein are hereby incorporated by reference in their entirety for all purposes.

WHAT IS CLAIMED IS:

1 1. A polypeptide comprising a fusion of a
2 transcription factor, the transcription factor comprising a
3 DNA binding domain, and a retinoblastoma (RB) polypeptide, the
4 RB polypeptide comprising a growth suppression domain.

1 2. A nucleic acid encoding the fusion polypeptide
2 of claim 1.

1 3. The nucleic acid of claim 2, wherein the
2 nucleic acid is inserted in an adenovirus vector.

1 4. The polypeptide of claim 1, wherein the
2 transcription factor is E2F.

1 5. The polypeptide of claim 4, wherein the cyclin
2 A binding domain of the E2F is deleted or nonfunctional.

1 6. The polypeptide of claim 1, wherein the
2 retinoblastoma polypeptide is RB56.

1 7. The polypeptide of claim 1, wherein the
2 retinoblastoma polypeptide is wild type RB.

1 8. The polypeptide of claim 1, wherein the
2 retinoblastoma polypeptide comprises from about amino acid
3 residue 379 to about amino acid residue 928 of pRB.

1 9. The polypeptide of claim 1, wherein the
2 retinoblastoma polypeptide comprises at least one substitution
3 of amino acid residues selected from the group consisting of
4 2, 608, 612, 788, 807, and 811 of pRB.

1 10. The polypeptide of claim 5, wherein the E2F
2 comprises about amino acid residues 95 to about 286.

11. The polypeptide of claim 4, wherein the E2F comprises about amino acid residues 95 to about 194.

12. The polypeptide of claim 1, wherein the fusion comprises EF2 amino acid residues from about 95 to about 194 operatively linked to RB amino acid residues from about 379 to about 928.

13. An expression vector comprising DNA encoding a polypeptide, the polypeptide comprising a fusion of a transcription factor, the transcription factor comprising a DNA binding domain, and a retinoblastoma (RB) polypeptide, the RB polypeptide comprising a growth suppression domain.

14. The vector of claim 13, comprising a tissue-specific promoter operatively linked to DNA encoding the fusion.

15. The vector of claim 14, wherein the tissue specific promoter is a smooth muscle actin promoter.

16. A method for treatment of a hyperproliferative disorder in a patient comprising administering to a patient a therapeutically effective dose of a fusion polypeptide comprising a fusion of a transcription factor, the transcription factor comprising a DNA binding domain, and a retinoblastoma (RB) polypeptide, the RB polypeptide comprising a growth suppression domain.

17. The method of claim 16, wherein the fusion protein is encoded by a nucleic acid delivered to the patient.

18. The method of claim 16, wherein the transcription factor is E2F.

19. The method of claim 18, wherein the cyclin A binding domain of the E2F is deleted or nonfunctional.

1 20. The method of claim 16, wherein the RB is RB56.

1 21. The method of claim 16, wherein the RB is wild
2 type RB56.

1 22. The method of claim 16, wherein the RB
2 comprises from about amino acid residue 379 to about amino
3 acid residue 928.

1 23. The method of claim 16, wherein the RB
2 comprises at least one substitution of amino acid residues
3 selected from the group consisting of 2, 608, 612, 788, 807,
4 and 811.

1 24. The method of claim 18, wherein the E2F
2 comprises about amino acid residues 95 to about 286.

1 25. The method of claim 18, wherein the E2F
2 comprises about amino acid residues 95 to about 194.

1 26. The method of claim 16, wherein the fusion
2 comprises EF2 amino acid residues from about 95 to about 194
3 operatively linked to RB amino acid residues from about 379 to
4 about 928.

1 27. The method of claim 18, wherein the E2F -RB
2 fusion polypeptide is expressed under the control of a tissue-
3 specific promoter.

1 28. The method of claim 27, wherein the tissue
2 specific promoter is a smooth muscle actin promoter.

1 29. The method of claim 16, wherein the
2 hyperproliferative disorder is cancer.

1 30. The method of claim 29, wherein the cancer is
2 bladder cancer.

1 31. The method of claim 29, wherein the
2 hyperproliferative disorder is restenosis.

1 32. The method of claim 31, wherein the E2F-RB
2 fusion polypeptide is administered after angioplasty.

1 33. The method of claim 32, wherein the E2F-RB
2 fusion polypeptide is administered as a coating on an
3 angioplasty device.

1 34. The method of claim 17, wherein the nucleic
2 acid is administered after angioplasty.

1 35. The method of claim 17, wherein the nucleic
2 acid is administered as a coating on an angioplasty device.

1 36. The method of claim 17, wherein the nucleic
2 acid is inserted in an adenovirus vector.

TISSUE SPECIFIC EXPRESSION OF RETINOBLASTOMA
PROTEIN

5

ABSTRACT OF THE DISCLOSURE

10

Fusions of the transcription factor E2F and the
retinoblastoma protein RB are provided, along with methods of
treatment of hyperproliferative diseases.

10	20	30	40	50	60
MALAGAPAGG	PCAPALEALL	GAGALRL LDS	SQIVIISAAQ	DASAPPAPTG	PAAPAAGPCD
70	80	90	100	110	120
PDLLLFATPQ	APRPTPSAPR	PALGRPPVKR	RLDLETDHQY	LAESSGPARG	RGRHPGKGVK
130	140	150	160	170	180
SPGEKSRYET	SLNLTTKRFL	ELLSHSADGV	VDLNWAAEVL	KVQKRRIYDI	TNVLEGIQLI
190	200	210	220	230	240
AKKSKNHIQW	LGSHTTVGVG	GRLEGLTQDL	RQLQESEQQL	DHLMNICTTQ	LRLLED TDS
250	260	270	280	290	300
QRLAYVTCQD	LRSIADPAEQ	MVMVIKAPPE	TQLQAVDSSE	NFQISL KSKQ	GPIDVFLCPE
310	320	330	340	350	360
ETVGGISPGK	TPSQEVTSEE	ENRATDSATI	VSPPPSSPPS	SLTTDPSQSL	LSLEQEPLLS
370	380	390	400	410	420
RMGSLRAPVD	EDRLSPLVAA	DSLLEHVRED	FSGLLP EEFI	SLSP PHEALD	YHFGLEE GEG
430	440	450	460	470	480
IRDLFDCDFG	DLTPLDF*

FIG. 1A

10	20	30	40	50	60
GGAATTCCGT	GGCCGGGACT	TTGCAGGCAG	CGGCGGCCCG	GGGCGGAGCG	GGATCGAGCC
70	80	90	100	110	120
CTCGCCGAGG	CCTGCCGCCA	TGGGCCCCGCG	CCGCCGCCCG	CGCCTGTCAC	CCGGGCCGCG
130	140	150	160	170	180
CGGGCCGTGA	GCGTCATGGC	CTTGGCCGGG	CCCCCTGCGG	GCGGCCCATG	CGCGCCGGCG
190	200	210	220	230	240
CTGGAGGCCC	TGCTCGGGGC	CGGCGCGCTG	CGGCTGCTCG	ACTCCTCGCA	GATCGTCATC
250	260	270	280	290	300
ATCTCCGCCG	CGCAGGACGC	CAGCGCCCCG	CCGGCTCCCA	CCGGCCCCCG	GGCGCCCCGCC
310	320	330	340	350	360
GCCGGCCCCCT	GCGACCCTGA	CCTGCTGCTC	TTCGCCACAC	CGCAGGCGCC	CCGGCCCCACA
370	380	390	400	410	420
CCCAGTGC GC	CGCGGCCCGC	GCTCGGCCGC	CCGCCGGTGA	AGCGGAGGCT	GGACCTGGAA
430	440	450	460	470	480
ACTGACCATC	AGTACCTGGC	CGAGAGCAGT	GGGCCAGCTC	GGGGCAGAGG	CCGCCATCCA
490	500	510	520	530	540
GGAAAAGGTG	TGAAATCCCC	GGGGGAGAAG	TCACGCTATG	AGACCTCACT	GAATCTGACC
550	560	570	580	590	600
ACCAAGCGCT	TCCTGGAGCT	GCTGAGCCAC	TCGGCTGACG	GTGTCTGTCG	CCTGAACTGG
610	620	630	640	650	660
GCTGCCGAGG	TGCTGAAGGT	GCAGAAGCGG	CGCATCTATG	ACATCACCAA	CGTCCTTGAG
670	680	690	700	710	720
GGCATCCAGC	TCATTGCCAA	GAAGTCCAAG	AACCACATCC	AGTGGCTGGG	CAGCCACACC
730	740	750	760	770	780
ACAGTGGGCG	TCGGCGGACG	GCTTGAGGGG	TTGACCCAGG	ACCTCCGACA	GCTGCAGGAG
790	800	810	820	830	840
AGCGAGCAGC	AGCTGGACCA	CCTGATGAAT	ATCTGTACTA	CGCAGCTGCG	CCTGCTCTCC
850	860	870	880	890	900
GAGGACACTG	ACAGCCAGCG	CCTGGCCTAC	GTGACGTGTC	AGGACCTTCG	TAGCATTGCA
910	920	930	940	950	960
GACCCTGCAG	AGCAGATGGT	TATGGTGATC	AAAGCCCCCTC	CTGAGACCCA	GCTCCAAGCC
970	980	990	1000	1010	1020
GTGGACTCTT	CGGAGAACTT	TCAGATCTCC	CTTAAGAGCA	AACAAGGCCC	GATCGATGTT
1030	1040	1050	1060	1070	1080
TTCCTGTGCC	CTGAGGAGAC	CGTAGGTGGG	ATCAGCCCTG	GGAAGACCCC	ATCCCAGGAG
1090	1100	1110	1120	1130	1140
GTCACTTCTG	AGGAGGAGAA	CAGGGCCACT	GACTCTGCCA	CCATAGTGTC	ACCACCACCA
1150	1160	1170	1180	1190	1200
TCATCTCCCC	CCTCATCCCT	CACCACAGAT	CCCAGCCAGT	CTCTACTCAG	CCTGGAGCAA
1210	1220	1230	1240	1250	1260
GAACCGCTGT	TGTCCCGGAT	GGGCAGCCTG	CGGGCTCCCG	TGGACGAGGA	CCGCCTGTCC

FIG. 1B

1270	1280	1290	1300	1310	1320
CCGCTGGTGG	CGGCCGACTC	GCTCCTGGAG	CATGTGCGGG	AGGACTTCTC	CGGCCTCCTC
1330	1340	1350	1360	1370	1380
CCTGAGGAGT	TCATCAGCCT	TTCCCCACCC	CACGAGGCCC	TCGACTACCA	CTTCGGCCTC
1390	1400	1410	1420	1430	1440
GAGGAGGGCG	AGGGCATCAG	AGACCTCTTC	GACTGTGACT	TTGGGGACCT	CACCCCCCTG
1450	1460	1470	1480	1490	1500
GATTTCTGAC	AGGGCTTGGA	GGGACCAGGG	TTTCCAGAGT	AGCTCACCTT	GTCTCTGCAG
1510	1520	1530	1540	1550	1560
CCCTGGAGCC	CCCTGTCCCT	GGCCGTCCCTC	CCAGCCTGTT	TGGAAACATT	TAATTTATAC
1570	1580	1590	1600	1610	1620
CCCTCTCCTC	TGTCTCCAGA	AGCTTCTAGC	TCTGGGGTCT	GGCTACCGCT	AGGAGGCTGA
1630	1640	1650	1660	1670	1680
GCAAGCCAGG	AAGGGAAGGA	GTCTGTGTGG	TGTGTATGTG	CATGCAGCCT	ACACCCACAC
1690	1700	1710	1720	1730	1740
GTGTGTACCG	GGGGTGAATG	TGTGTGAGCA	TGTGTGTGTG	CATGTACCGG	GGAATGAAGG
1750	1760	1770	1780	1790	1800
TGAACATACA	CCTCTGTGTG	TGCACTGCAG	ACACGCCCCA	GTGTGTCCAC	ATGTGTGTGC
1810	1820	1830	1840	1850	1860
ATGAGTCCAT	CTCTGCGCGT	GGGGGGGCTC	TAAGTGCCTT	TTGGGCCCTT	TTGCTCGTGG
1870	1880	1890	1900	1910	1920
GGTCCCACAA	GGCCCAGGGC	AGTGCCTGCT	CCCAGAATCT	GGTGCTCTGA	CCAGGCCAGG
1930	1940	1950	1960	1970	1980
TGGGGAGGCT	TTGGCTGGCT	GGGCGTGTAG	GACGGTGAGA	GCACTTCTGT	CTTAAAGGTT
1990	2000	2010	2020	2030	2040
TTTTCTGATT	GAAGCTTTAA	TGGAGCGTTA	TTTATTTATC	GAGGCCTCTT	TGGTGAGCCT
2050	2060	2070	2080	2090	2100
GGGGAATCAG	CAAAAGGGGA	GGAGGGGTGT	GGGGTTGATA	CCCCAACTCC	CTCTACCCTT
2110	2120	2130	2140	2150	2160
GAGCAAGGGC	AGGGGTCCCT	GAGCTGTTCT	TCTGCCCCAT	ACTGAAGGAA	CTGAGGCCTG
2170	2180	2190	2200	2210	2220
GGTGATTTAT	TTATTGGGAA	AGTGAGGGAG	GGAGACAGAC	TGACTGACAG	CCATGGGTGG
2230	2240	2250	2260	2270	2280
TCAGATGGTG	GGGTGGGCCC	TCTCCAGGGG	GCCAGTTCAG	GGCCCAGCTG	CCCCCCAGGA
2290	2300	2310	2320	2330	2340
TGGATATGAG	ATGGGAGAGG	TGAGTGGGGG	ACCTTCACTG	ATGTGGGCAG	GAGGGGTGGT
2350	2360	2370	2380	2390	2400
GAAGGCCTCC	CCCAGCCCAG	ACCCTGTGGT	CCCTCCTGCA	GTGTCTGAAG	CGCCTGCCTC
2410	2420	2430	2440	2450	2460
CCCAGTCTC	TGCCCCACCC	TCCAATCTGC	ACTTTGATTT	GCTTCCTAAC	AGCTCTGTTC
2470	2480	2490	2500	2520	2520
CCTCCTGCTT	TGGTTTAAAT	AAATATTTTG	ATGACGTAA	AAAAAGGAAT	TCGATAT

FIG. 1B
(CONTINUED)

1 ttccgggtttt tctcagggga cgttgaaatt atttttgtaa cgggagtcgg gagaggacgg
 61 ggcgtgcccc gcgtgcgcgc gcgtgcgtct ccccggcgct cctccacagc tcgctggctc
 121 ccgcccgcga aaggcgtcat gccgcccaaa acccccgcga aaacggccgc caccgccgcc
 181 gctgccgcgc cggaaccccc ggcaccgcgc ccgcccgcgc ctcttgagga ggaccagag
 241 caggacagcg gcccgaggga cctgcctctc gtcaggcttg agtttgaaga aacagaagaa
 301 cctgatttta ctgcattatg tcagaaatta aagataccag atcatgtcag agagagagct
 361 tgggttaactt gggagaaagt ttcattctgt gatggagtat tgggagggtta tattcaaaag
 421 aaaaaggaac tgtggggaat ctgtatcttt attgcagcag ttgacctaga tgagatgtcg
 481 ttcactttta ctgagctaca gaaaaacata gaaatcagtg tccataaatt ctttaactta
 541 ctaaaagaaa ttgataccag taccaaagtt gataatgcta tgtcaagact gttgaagaag
 601 tatgatgtat tgtttgcact cttcagcaaa ttggaaagga catgtgaact tatatatattg
 661 acacaaccca gcagttcgat atctactgaa ataaattctg cattgggtgct aaaagtttct
 721 tggatcacat ttttattagc taaaggggaa gtattacaaa tggaagatga tctgggtgatt
 781 tcattttcagt taatgctatg tgtccttgac tattttatta aactctcacc tcccatgttg
 841 ctcaaagaac catataaaac agctgttata cccattaatg gttcacctcg aacacccagg
 901 cgaggtcaga acaggagtgc acggatagca aaacaactag aaaatgatac aagaattatt
 961 gaagttctct gtaaagaaca tgaatgtaat atagatgagg tgaaaaatgt ttatttcaaa
 1021 aattttatac cttttatgaa ttctcttgga cttgtaacat ctaatggact tccagagggt
 1081 gaaaatcttt ctaaacgata cgaagaaatt tatcttaaaa ataaagatct agatgcaaga
 1141 ttatttttgg atcatgataa aactcttcag actgattcta tagacagttt tgaaacacag
 1201 agaacaccac gaaaaagtaa ccttgatgaa gaggtgaatg taattcctcc acacactcca
 1261 gttaggactg ttatgaacac tatccaacaa ttaatgatga ttttaaatcc agcaagtgat
 1321 caaccttcag aaaatctgat ttctatttt aacaactgca cagtgaatcc aaaagaaagt
 1381 atactgaaaa gagtgaagga tataggatac atcttttaaag agaaatttgc taaagctgtg
 1441 ggacagggtt gtgtcgaaat tggatcacag cgatacaaac ttggagttcg cttgtattac
 1501 cgagtaatgg aatccatgct taaatcagaa gaagaacgat tatccattca aaatttttagc
 1561 aaacttctga atgacaacat ttttcatatg tctttattgg cgtgcgctct tgaggttgta
 1621 atggccacat atagcagaag tacatctcag aatcttgatt ctggaacaga tttgtctttc
 1681 ccatggattc tgaatgtgct taatttaaaa gcccttgatt tttacaaagt gatcgaaagt
 1741 tttatcaaag cagaaggcaa cttgacaaga gaaatgataa aacattttaga acgatgtgaa
 1801 catcgaatca tggaaatccct tgcattggctc tcagattcac ctttatttga tcttattaaa
 1861 caatcaaagg accgagaagg accaactgat caccttgaat ctgcttgctc tcttaatctt
 1921 cctctccaga ataatcacac tgcagcagat atgtatcttt ctctgtgaag atctccaaag
 1981 aaaaaagggt caactacgcg tgtaaatctt actgcaaattg cagagacaca agcaacctca
 2041 gccttccaga cccagaagcc attgaaatct acctctcttt cactgtttta taaaaaagtg
 2101 tatcggttag cctatctccg gctaaataca ctttgtgaac gccttctgtc tgagcaccga
 2161 gaattagaac atatcatctg gaccttttcc cagcacaccc tgcagaatga gtatgaactc
 2221 atgagagaca ggcattttgga ccaaattatg atgtgttcca tgtatggcat atgcaaagtg
 2281 aagaatatag accttaaat caaaatcatt gtaacagcat acaaggatct tctcatgct
 2341 gttcaggaga cattcaaacc tgttttgatc aaagaagagg agtatgattc tattatagta
 2401 ttctataact cggctctcat gcagagactg aaaacaaata ttttgcagta tgcttccacc
 2461 aggccccccta ccttgctcac aatacctcac attcctcgaa gcccttacia ccctgaagag tccatataaa
 2521 tcacccttac ggattcctgg agggaacatc tatatttcac gatcaagaat cttagtatca
 2581 atttcagaag gtctgccaac accaacaata atgactccaa taaatcagat ggtatgtaac
 2641 attggtgaat cattcgggac ttctgagaag ttccagaaaa ctcctaaacc actgaaaaaa
 2701 agcgaccgtg tgctcaaaag aagtgtgtaa ggaagcaacc ctctaaaccc cccaggagag
 2761 ctacgctttg atattgaagg atcagatgaa gcagatggaa gtaaacatct gcaaaagcag
 2821 tccaaatttc agcagaaact ggcagaaatg acttctactc gaacacgaat gcaaaagcag
 2881 aaaatgaatg atagcatgga tacctcaaac aaggaagaga aatgaggatc tcaggacctt
 2941 ggtggacact gtgtacacct ctggattcat tgtctctcac agatgtgact gtat

FIG. 2A

"MPPKTPRKTAATAAAAAAEPAPPPPPPEEDPEQDSGPEDLPL
VRLEFEETEEPDFTALCQKLKIPDHVRERAWLTWEKVSSVDGVLGGYIQKKKELWGIC
IFIAAVDLDEMSFTFTTELQKNIEISVHKFFNLLKEIDTSTKVDNAMSRLKKYDVLFA
LFSKLERTCELIYLTQPSSSISTEINSALVLKVSUITFLLAKGEVLQMEDDLVISFQL
MLCVLDYFIKLSPPMLLKEPYKTAVIPINGSRTPRRGQNRSARIAKQLENDTRIIEV
LCKEHECNIDEVKNVYFKNFIPFMNSLGLVTSNGLPEVENLSKRYEEIYLKNKDLDA
LFLDHDKTLQTDSDSFETQRTPRKSNLDEEVNVI PPHTPVRTVMNTIQQLMILNSA
SDQPSENLISYFNNCTVNPKEISILKRVKDIFYIFKEKFAKAVGQGCVEIGSQRYKLG
RLYYRVMESMLKSEEERLSIQNF SKLLNDNIFHMSLLACALEVVMATYSRSTSQNLDS
GTDLSFPWILNVLNLKAFDFYKVIESFIKAEGNLTREMIKHLERCEHRIMESLAWLSD
SPLFDLIKQSKDREGPTDHLESACPLNLPLQNNHTAADMYLSPVRSPKKKGSTTRVNS
TANAETQATSAFQTQKPLKSTSLSLFYKKVYRLAYLRLNTLCERLLSEHPELEHIWT
LFQHTLQNEYELMRDRHLDQIMMCSMYGICKVKNI DLKFKIIVTAYKDLPHAVQETFK
RVLIKEEYDSIIVFYNSVFMQRLKTNILQYASTRPPTLSPIPHIPRSPYKFPSSPLR
IPCGNIYISPLKSPYKISEGLPTPTKMTPRSRI LVSIGESFGTSEKFQKINQMVCNSD
RVLKRSAEGSNPPKPLKKLRFDIEGSDEADGSKHLPGESKFQKLAEMTSTRTRMQKQ
KMNDSDMTSNKEEK"

FIG. 2B

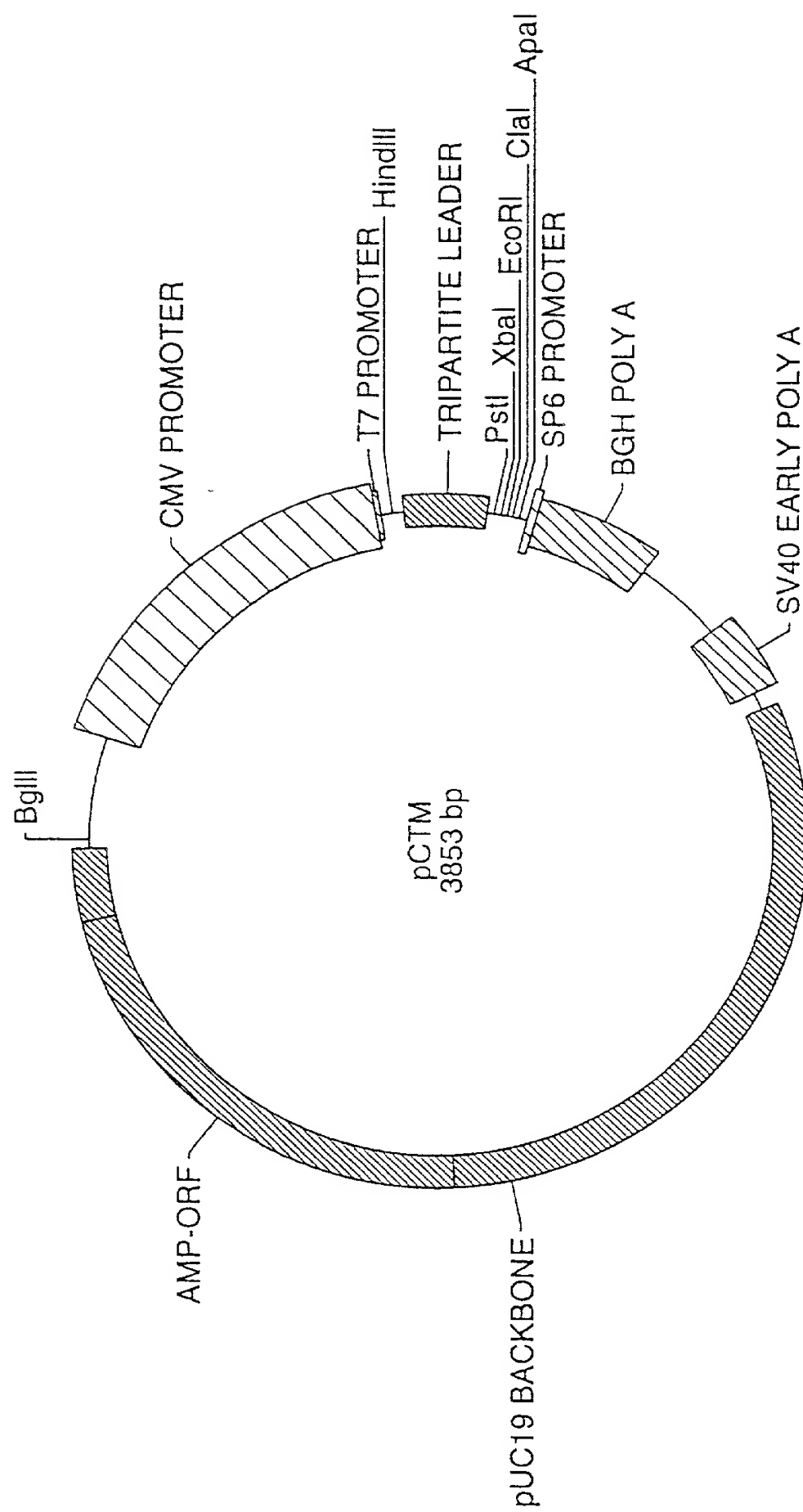


FIG. 3

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```

                                >HincII
                                |
                                >AccI
                                ||
                                >SalI
                                |||
                                |||
      10  | 20 30 40 50 60
      *  *  *  *  *  *
GACGGATCGG GAGATCTCCC GATCCCCTAT GGTCGACTCT CAGTACAATC TGCTCTGATG

                                >AlwNI
                                |
      70  | 80 90 100 110 120
      *  *  *  *  *  *
CCGCATAGTT AAGCCAGTAT CTGCTCCCTG CTTGTGTGTT GGAGGTCGCT GAGTAGTGCG

      >ApoI                                >MfeI
      | 130 140 150 160 | 170 180
      *  *  *  *  *  *  *
CGAGCAAAAT TTAAGCTACA ACAAGGCAAG GCTTGACCGA CAATTGCATG AAGAATCTGC

                                                                >HincII
                                                                |
                                                                >AflIII
                                                                |
                                                                >MluI
                                                                |
      190 200 210 220 230
      *  *  *  *  *
TTAGGGTTAG GCGTTTTGCG CTGCTTCG CGA TGT ACG GGC CAG ATA TAC GCG TTG
      Arg Cys Thr Gly Gln Ile Tyr Ala Leu>
      ___d___d___CMV PROMOTER___d___d___>

      >SpeI      >AseI
      |          |
      240 250 260 270 280
      *  *  *  *  *
ACA TTG ATT ATT GAC TAG TTA TTA ATA GTA ATC AAT TAC GGG GTC ATT
Thr Leu Ile Ile Asp *** Leu Leu Ile Val Ile Asn Tyr Gly Val Ile>
___d___d___d___d___d___d___CMV PROMOTER___d___d___d___d___d___d___>

      290 300 310 320 330
      *  *  *  *  *
AGT TCA TAG CCC ATA TAT GGA GTT CCG CGT TAC ATA ACT TAC GGT AAA
Ser Ser *** Pro Ile Tyr Gly Val Pro Arg Tyr Ile Thr Tyr Gly Lys>
___d___d___d___d___d___d___CMV PROMOTER___d___d___d___d___d___d___>

      >BglI                                >AatII
      |          |
      340 350 360 370
      *  *  *  *
TGG CCC GCC TGG CTG ACC GCC CAA CGA CCC CCG CCC ATT GAC GTC AAT
Trp Pro Ala Trp Leu Thr Ala Gln Arg Pro Pro Pro Ile Asp Val Asn>
___d___d___d___d___d___d___CMV PROMOTER___d___d___d___d___d___d___>

380 390 400 410 420
*  *  *  *  *
AAT GAC GTA TGT TCC CAT AGT AAC GCC AAT AGG GAC TTT CCA TTG ACG
Asn Asp Val Cys Ser His Ser Asn Ala Asn Arg Asp Phe Pro Leu Thr>
___d___d___d___d___d___d___CMV PROMOTER___d___d___d___d___d___d___>

```

FIG. 4

FIG. 4

(CONTINUED)

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```

      >BanII
      |
      >SacI
      |
      >BsiHKAI
      |
      >Ecl136II
      |
      820      830      840      850
      *|*      *      *      *      *
CAG AGC TCT CTG GCT AAC TAG AGA ACC CAC TGC TTA CTG GCT TAT CGA
Gln Ser Ser Leu Ala Asn *** Arg Thr His Cys Leu Leu Ala Tyr Arg>
_d _d _d _d _d _d _d CMV PROMOTER _d _d _d _d _d _d _d >

                                     >KpnI
                                     |
      >AseI      >BsaI      >Acc65I
      |          |          |
      >T7_PROMOTER  >SfcI      >BanI
      |          ||          |
      860      870      880      890      900      910
      *|*      *|*      *|*      *|*      *|*      *|*
AAT T AATACGA CTCACTATAG GGAGACCCAA GCTTCGCGCG GGTACCACTC
Asn Xxx>
_d _>

                                     >PflMI
                                     |
                                     >PvuII
                                     |
      >EarI      >MspAlI      >BanII
      |          |          |
      920      930      940      950      960      970
      *|*      *|*      *|*      *|*      *|*      *|*
TCTTCCGCAT CGCTGTCTGC GAGGGCCAGC TGTGGGCTC GCGGTTGAGG ACAAACCTCTT
_e _ _ _ _ _ TRIPARTITE LEADER SEQUENCE _ _ _ _ _>

      >EarI      >ScaI
      |          |
      980      990      1000      1010      1020      1030
      *|*      *|*      *|*      *|*      *|*      *|*
CGCGGTCTTT CCAGTACTCT TGGATCGGAA ACCCGTCGGC CTCCGAACGG TACTCCGCCA
_e _ _ _ _ _ TRIPARTITE LEADER SEQUENCE _ _ _ _ _>

                                     >SfcI
                                     |
                                     >MspAlI
                                     |
      >XhoI      >BsiEI
      |          |
      >PaeR71      >EaeI
      |          |
      >BsoBI      >NotI
      |          |
      >AvaI      >EagI
      |          |
      1040      1050      1060      1070      1080      1090
      *|*      *|*      *|*      *|*      *|*      *|*
CCGAGGGACC TGAGCGAGTC CGCATCGACC GGATCGGAAA ACCTCTCGAG GCGGCCGCTG
_e _ _ _ _ _ TRIPARTITE LEADER SEQUENCE _ _ _ _ _>

```

FIG. 4
(CONTINUED)

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```

                                >ApaI
                                  |
                    >ClaI    >EcoO109I
                      |        |
                >EcoRV|   >Bsp120I       >SfcI
                  ||      ||         |
          >XbaI  >ApoI           >PstI     >EcoRI   >BsiWI             >BspDI   ||>BanII              >MslI
          ||    |               ||         |            ||         |               | | |
          ||    | 1100         ||         |            ||         | 1120         | 1130         | 1140         |
          || *    *           ||         |            ||         | *         *           ||         |
CAGTCTAGAC GAATTCGCGT ACGATATCGA TGGGCCCTAT T CTA TAG TGT CAC CTA
Leu *** Cys His Leu>
SP6 PROMOTER >

```

```

      >BanII
      |
      >BsiHKAI
      |
      >SacI
      |
  >Ecl136II | >BclI
      |       |
  >BGH_POLY_A |
1150 | | | 1160 | | 1170 | 1180 | 1190 | 1200
    * | | * | | * | * | * | * | * |
AAT G CTAGAGCTCG CTGATCAGCC TCGACTGTGC CTTCTAGTTG CCAGCCATCT
Asn>
>

```

```

      1210      1220      1230      1240      1250      1260
      *      *      *      *      *      *
GTTGTTTGCC CCTCCCCCGT GCCTTCCTTG ACCCTGGAAG GTGCCACTCC CACTGTCCTT

      1270      1280      1290      1300      1310      1320
      *      *      *      *      *      *
TCCTAATAAA ATGAGGAAAT TGCATCGCAT TGTCTGAGTA GGTGTCATTC TATTCTGGGG

```

>BbsI

|

1330 1340 1350 1360 | 1370 1380

* * * * * * * * * *

GGTGGGGGTGG GGCAGGACAG CAAGGGGGGAG GATTGGGAAG ACAATAGCCG AAATGACCGA

```

                                >BssSI
                                |
                                >BspMI
                                |
      1390      1400      1410      1420      1430      1440
      *      *      *      *      *      *
CCAAGCGACG CCCAACCTGC CATCACGAGA TTTCGATTCC ACCGCCGCCT TCTATGAAAG

```

```

                                >NaeI
                                |
                                >BsrFI
                                |
                                >BpmI
                                |
                                |
                                >NgoMI
                                |
                                |
1450      1460      1470      1480      1490      1500
*          *          *          *          *          *
GTTGGGCTTC GGAATCGTTT TCCGGGACGC CGGCTGGATG ATCCTCCAGC GCGGGGATCT

```

FIG. 4
(CONTINUED)

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```

      >BpmI
      |
    >SV40_early_poly_A
      |
1510      1520      1530      1540      1550      1560
*      *      *      *      *      *
CATGCTGGAG TTCTTCGCCC ACCCCAACCTT GTTTATTGCA GCTTATAATG GTTACAAATA

      >ApoI
      |
1570      1580      1590      1600      1610      1620
*      *      *      *      *      *
AAGCAATAGC ATCACAAATT TCACAAATAA AGCATTTTTT TCACTGCATT CTAGTTGTGG

      >HincII
      |
      >Bst1107I  >AccI
      |          ||
      >AccI      >SalI
      ||         |||
1630      1640      1650      1660      1670      1680
*      *      *      *      *      *
TTTGTCCAAA CTCATCAATG TATCTTATCA TGTCTGTATA CCGTCGACCT CTAGCTAGAG
      >BsrBI
      |
1690      1700      1710      1720      1730      1740
*      *      *      *      *      *
CTTGCGGTAA TCATGGTCAT AGCTGTTTCC TGTGTGAAAT TGTTATCCGC TCACAATTCC
_____c_____PUC19 BACKBONE H3 TO AATII_____c_____>

      >BamI
      |
1750      1760      1770      1780      1790      1800
*      *      *      *      *      *
ACACAACATA CGAGCCGGAA GCATAAAGTG TAAAGCCTGG GGTGCCTAAT GAGTGAGCTA
_____c_____PUC19 BACKBONE H3 TO AATII_____c_____>

      >AseI
      |
1810      1820      1830      1840      1850      1860
*      *      *      *      *      *
ACTCACATTA ATTGCGTTGC GCTCACTGCC CGCTTTCCAG TCGGGAAACC TGTCGTGCCA
_____c_____PUC19 BACKBONE H3 TO AATII_____c_____>

>PvuII
|
>MspAlI  >AseI      >EaeI
|          |          |
1870      1880      1890      1900      1910      1920
*      *      *      *      *      *
GCTGCATTAA TGAATCGGCC AACGCGCGGG GAGAGGCGGT TTGCGTATTG GGCGCTCTTC
_____c_____PUC19 BACKBONE H3 TO AATII_____c_____>

>EarI
|
>SapI
|
1930      1940      1950      1960      1970      1980
*      *      *      *      *      *
CGCTTCCTCG CTTCACTGACT CGCTGCGCTC GGTCGTTCGG CTGCGGCGAG CGGTATCAGC
_____c_____PUC19 BACKBONE H3 TO AATII_____c_____>

```

FIG. 4
(CONTINUED)

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```

                                >AflIII
                                |
      1990      2000      2010      2020      2030      2040
      *      *      *      *      *      *
TCACTCAAAG GCGGTAATAC GGTATCCAC AGAATCAGGG GATAACGCAG GAAAGAACAT
_____c_____PUC19 BACKBONE H3 TO AATII_____c_____>

      2050      2060      2070      2080      2090      2100
      *      *      *      *      *      *
GTGAGCAAAA GGCCAGCAAA AGGCCAGGAA CCGTAAAAAG GCCGCGTTGC TGGCGTTTTT
_____c_____PUC19 BACKBONE H3 TO AATII_____c_____>

                                >DrdI
                                |
      2110      2120      2130      2140      2150      2160
      *      *      *      *      *      *
CCATAGGCTC CGCCCCCCTG ACGAGCATCA CAAAATCGA CGCTCAAGTC AGAGGTGGCG
_____c_____PUC19 BACKBONE H3 TO AATII_____c_____>

                                >BssSI
                                |
      2170      2180      2190      2200      2210      2220
      *      *      *      *      *      *
AAACCCGACA GGACTATAAA GATACCAGGC GTTCCCCCT GGAAGCTCCC TCGTGCGCTC
_____c_____PUC19 BACKBONE H3 TO AATII_____c_____>

                                >BsaWl
                                |
      2230      2240      2250      2260      2270      2280
      *      *      *      *      *      *
TCCTGTTCCG ACCCTGCCGC TTACCGGATA CCTGTCCGCC TTTCTCCCTT CGGGAAGCGT
_____c_____PUC19 BACKBONE H3 TO AATII_____c_____>

>HaeII      >SfcI
|            |
| 2290      2300      2310      2320      2330      2340
| *      *      *      *      *      *
GGCGCTTTCT CAATGCTCAC GCTGTAGGTA TCTCAGTTCG GTGTAGGTCG TTCGCTCCAA
_____c_____PUC19 BACKBONE H3 TO AATII_____c_____>

      >BsiHKAI      >MspAI
      |            |
      >ApaLI      >BsiEI      >BsaWI
      |            |            |
      2350      2360      2370      2380      2390      2400
      *      *      *      *      *      *
GCTGGGCTGT GTGCACGAAC CCCCCGTTCA GCCCGACCGC TGCGCCTTAT CCGGTAACATA
_____c_____PUC19 BACKBONE H3 TO AATII_____c_____>

                                >AlwNI
                                |
      2410      2420      2430      2440      2450      2460
      *      *      *      *      *      *
TCGTCTTGAG TCCAACCCGG TAAGACACGA CTTATCGCCA CTGGCAGCAG CCACTGGTAA
_____c_____PUC19 BACKBONE H3 TO AATII_____c_____>

                                >SfcI
                                |
      2470      2480      2490      2500      2510      2520
      *      *      *      *      *      *
CAGGATTAGC AGAGCGAGGT ATGTAGGCGG TGCTACAGAG TTCTTGAAGT GGTGGCCTAA
_____c_____PUC19 BACKBONE H3 TO AATII_____c_____>

```

FIG. 4
(CONTINUED)

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```

      2530      2540      2550      2560      2570      2580
      *      *      *      *      *      *
CTACGGCTAC ACTAGAAGGA CAGTATTTGG TATCTGCGCT CTGCTGAAGC CAGTTACCTT
_____c_____PUC19 BACKBONE H3 TO AATII_____c_____>

>Eco57I
|
| 2590      2600      2610      2620      2630      2640
      *      *      *      *      *      *
CGGAAAAAGA GTTGGTAGCT CTTGATCCGG CAAACAAACC ACCGCTGGTA GCGGTGGTTT
_____c_____PUC19 BACKBONE H3 TO AATII_____c_____>

      2650      2660      2670      2680      2690      2700
      *      *      *      *      *      *
TTTTGTTTGC AAGCAGCAGA TTACGCGCAG AAAAAAGGA TCTCAAGAAG ATCCTTTGAT
_____c_____PUC19 BACKBONE H3 TO AATII_____c_____>

                                                    >BspHI
                                                    |
      2710      2720      2730      2740      2750      2760
      *      *      *      *      *      *
CTTTTCTACG GGGTCTGACG CTCAGTGGAA CGAAAACTCA CGTTAAGGGA TTTTGGTCAT
_____c_____PUC19 BACKBONE H3 TO AATII_____c_____>

                                >DraI
                                |
      2770      2780      2790      2800      2810      2820
      *      *      *      *      *      *
GAGATTATCA AAAAGGATCT TCACCTAGAT CCTTTTAAAT TAAAAATGAA GTTTTAAATC
_____c_____PUC19 BACKBONE H3 TO AATII_____c_____>

                                                    >DraI
                                                    |
      2830      2840      2850      2860      2870      2880
      *      *      *      *      *      *
AATCTAAAGT ATATATGAGT AACTTGGTC TGACAGTTAC CAATGCTTAA TCAGTGAGGC
_____c_____PUC19 BACKBONE H3 TO _____a_____AMP-ORF_____>
_____c_____PUC19 BACKBONE H3 TO AATII_____c_____>

                                                    >BanI
                                                    |
      2830      2840      2850      2860      2870      2880
      *      *      *      *      *      *
AATCTAAAGT ATATATGAGT AACTTGGTC TGACAGTTAC CAATGCTTAA TCAGTGAGGC
_____c_____PUC19 BACKBONE H3 TO _____a_____AMP-ORF_____>
_____c_____PUC19 BACKBONE H3 TO AATII_____c_____>

                                                    >AhdI
                                                    |
      2890      2900      2910      2920      2930      2940
      *      *      *      *      *      *
ACCTATCTCA GCGATCTGTC TATTTGTTTC ATCCATAGTT GCCTGACTCC CCGTCGTGTA
_____a_____a_____AMP-ORF_____a_____a_____>
_____c_____PUC19 BACKBONE H3 TO AATII_____c_____>

                                                    >BsaI
                                                    |
                                                    >BsrDI
                                                    |
                                                    >BpmI
                                                    |
      2950      2960      2970      2980      2990      3000
      *      *      *      *      *      *
GATAACTACG ATACGGGAGG GCTTACCATC TGGCCCCAGT GCTGCAATGA TACCGCGAGA
_____a_____a_____AMP-ORF_____a_____a_____>
_____c_____PUC19 BACKBONE H3 TO AATII_____c_____>

      >BsrFI
      |
      3010      3020      3030      3040      3050      3060
      *      *      *      *      *      *
CCCACGCTCA CCGGCTCCAG ATTTATCAGC AATAAACCAG CCAGCCGGAA GGGCCGAGCG
_____a_____a_____AMP-ORF_____a_____a_____>
_____c_____PUC19 BACKBONE H3 TO AATII_____c_____>

```

FIG. 4
(CONTINUED)

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```

                                >AseI
                                |
      3070      3080      3090      3100      3110      3120
      *      *      *      *      *      *
CAGAAGTGGT CCTGCAACTT TATCCGCCTC CATCCAGTCT ATTAATTGTT GCCGGGAAGC
      a      a      AMP-ORF      a      a      >
      c      PUC19 BACKBONE H3 TO AATII      c      >

                                >PspI406I
                                |
                                >FspI      >BsrDI      >SfiI
      3130      3140      3150      3160      3170      3180
      *      *      *      *      *      *
TAGAGTAAGT AGTTCGCCAG TTAATAGTTT GCGCAACGTT GTTGCCATTG CTACAGGCAT
      a      a      AMP-ORF      a      a      >
      c      PUC19 BACKBONE H3 TO AATII      c      >

>MslI                                >BsaWI
|                                |
      3190      3200      3210      3220      3230      3240
      *      *      *      *      *      *
CGTGGTGTCA CGCTCGTCGT TTGGTATGGC TTCATTCAGC TCCGGTCCC AACGATCAAG
      a      a      AMP-ORF      a      a      >
      c      PUC19 BACKBONE H3 TO AATII      c      >

                                >PvuI
                                |
                                >BsiEI
      3250      3260      3270      3280      3290      3300
      *      *      *      *      *      *
GCGAGTTACA TGATCCCCCA TGTTGTGCAA AAAAGCGGTT AGCTCCTTCG GTCCTCCGAT
      a      a      AMP-ORF      a      a      >
      c      PUC19 BACKBONE H3 TO AATII      c      >

                                >EaeI                                >MslI
                                |                                |
      3310      3320      3330      3340      3350      3360
      *      *      *      *      *      *
CGTTGTCAGA AGTAAGTTGG CCGCAGTGTT ATCACTCATG GTTATGGCAG CACTGCATAA
      a      a      AMP-ORF      a      a      >
      c      PUC19 BACKBONE H3 TO AATII      c      >

                                >ScaI
                                |
      3370      3380      3390      3400      3410      3420
      *      *      *      *      *      *
TTCTCTTACT GTCATGCCAT CCGTAAGATG CTTTCTGTG ACTIGGTGAGT ACTCAACCAA
      a      a      AMP-ORF      a      a      >
      c      PUC19 BACKBONE H3 TO AATII      c      >

                                >BsiEI
                                |
      3430      3440      3450      3460      3470      3480
      *      *      *      *      *      *
GTCATTCTGA GAATAGTGTA TCGGCGGACC GAGTTGCTCT TGCCCGGCGT CAATACGGGA
      a      a      AMP-ORF      a      a      >
      c      PUC19 BACKBONE H3 TO AATII      c      >

```

FIG. 4
(CONTINUED)

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```

                                >XmnI
                                |
                                >DraI   >BsiHKAI   >Psp1406I
                                |       |       |
          3490      3500      3510      3520      3530      3540
          *        *        *        *        *        *
TAATACCGCG CCACATAGCA GAACTTTAAA AGTGCTCATC ATTGGAAAAC GTTCTTCGGG
          a          a          AMP-ORF          a          a
          c          PUC19 BACKBONE H3 TO AATII          c          >

                                >Eco571
                                |
                                >ApaLI
                                |
                                >MspAlI
                                |       >BssSI
          3550      3560      3570      3580      3590      3600
          *        *        *        *        *        *
GCGAAACTC TCAAGGATCT TACCGCTGTT GAGATCCAGT TCGATGTAAC CCACTCGTGC
          a          a          AMP-ORF          a          a
          c          PUC19 BACKBONE H3 TO AATII          c          >

>BsiHKAI
|
|      3610      3620      3630      3640      3650      3660
|      *        *        *        *        *        *
|ACCCAACTGA TCTTCAGCAT CTTTACTTT CACCAGCGTT TCTGGGTGAG CAAAAACAGG
|          a          a          AMP-ORF          a          a
|          c          PUC19 BACKBONE H3 TO AATII          c          >

                                >MslI
                                |
          3670      3680      3690      3700      3710      3720
          *        *        *        *        *        *
AAGGCAAAAT GCCGCAAAAA AGGGAATAAG GGCGACACGG AAATGTTGAA TACTCATACT
          a          a          AMP-ORF          a          a
          c          PUC19 BACKBONE H3 TO AATII          c          >

>EarI   >SspI                                >BspHI   >BsrBI
|       |       |       |       |       |       |       |
| 3730 | 3740 | 3750 | 3760 | 3770 | 3780
| *   * | *   * | *   * | *   * | *   * | *   *
CTTCCTTTT CAATATTATT GAAGCATTTA TCAGGGTTAT TGTCTCATGA GCGGATACAT
          c          PUC19 BACKBONE H3 TO AATII          c          >

          3790      3800      3810      3820      3830      3840
          *        *        *        *        *        *
ATTTGAATGT ATTTAGAAAA ATAAACAAAT AGGGGTTCCG CGCACATTTC CCCGAAAAGT
          c          PUC19 BACKBONE H3 TO AATII          c          >

>HincII
|
>AccI
||
>AatII
||
>SalI
|||
3850 |||
*   * |||
GCCACCTGAC GTC
          c          >

```

FIG. 4
(CONTINUED)

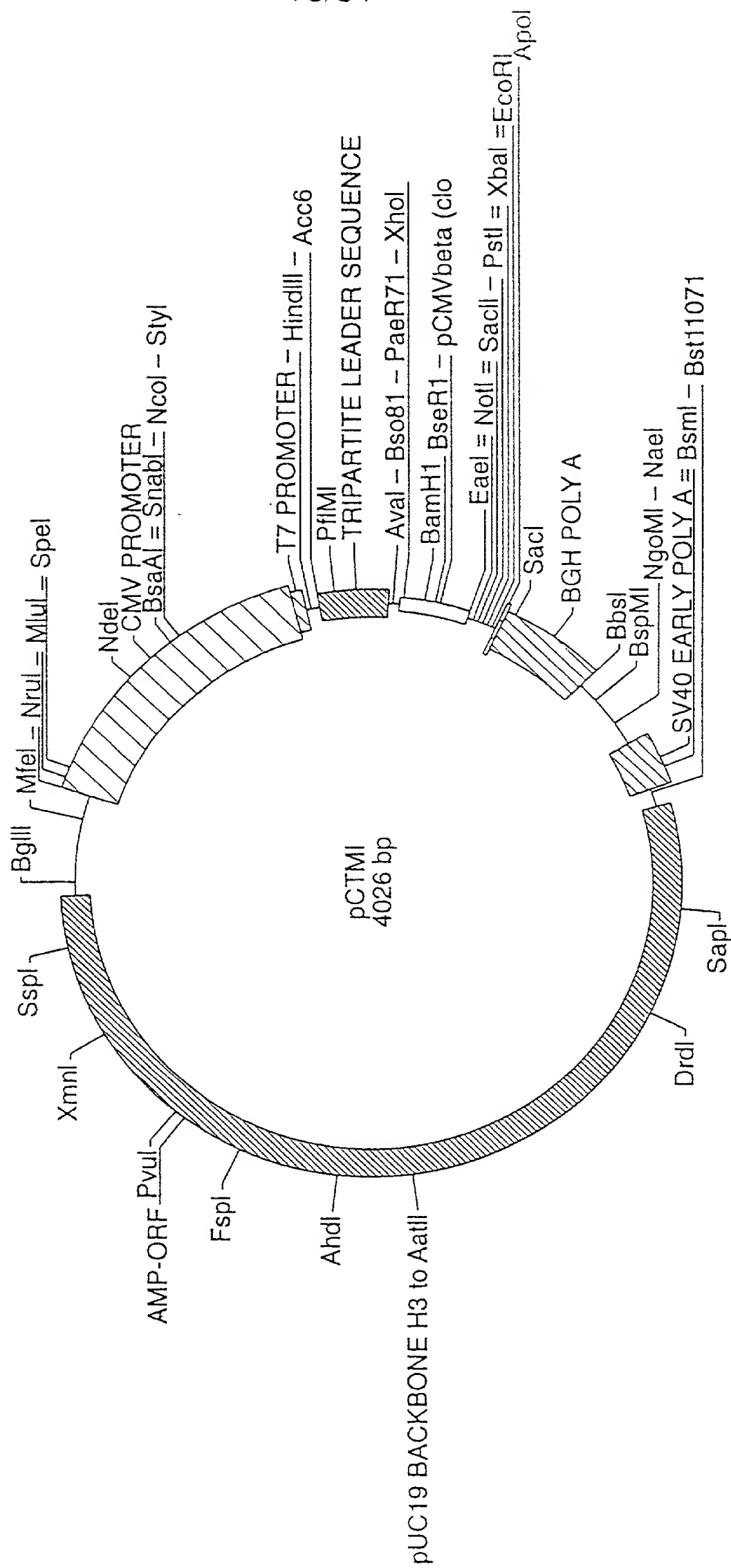


FIG. 5

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```

                                >HincII
                                |
                                >AccI
                                ||
                                >SalI
                                |||
                                |||
    >BglIII
    |
    10 20 30 40 50 60
    *  *  *  *  *  *
GACGGATCGG GAGATCTCCC GATCCCCTAT GGTCGACTCT CAGTACAATC TGCTCTGATG

                                >AlwNI
                                |
    70 80 90 100 110 120
    *  *  *  *  *  *
CCGCATAGTT AAGCCAGTAT CTGCTCCCTG CTTGTGTGTT GGAGGTCGCT GAGTAGTGCG

    >ApoI
    |
    130 140 150 160 170 180
    *  *  *  *  *  *
CGAGCAAAAT TTAAGCTACA ACAAGGCAAG GCTTGACCGA CAATTGCATG AAGAATCTGC

                                >HincII
                                |
                                >AflIII
                                |
                                >MluI
                                |
    190 200 210 220 230
    *  *  *  *  *
TTAGGGTTAG GCGTTTTCG CTGCTTCG CGA TGT ACG GGC CAG ATA TAC GCG TTG
    Arg Cys Thr Gly Gln Ile Tyr Ala Leu>
    ___e___e___CMV PROMOTER___e___e___>

    >SpeI
    |
    240 250 260 270 280
    *  *  *  *  *
ACA TTG ATT ATT GAC TAG TTA TTA ATA GTA ATC AAT TAC GGG GTC ATT
Thr Leu Ile Ile Asp *** Leu Leu Ile Val Ile Asn Tyr Gly Val Ile>
    ___e___e___e___e___e___e___e___CMV PROMOTER___e___e___e___e___e___e___>

    >AseI
    |
    290 300 310 320 330
    *  *  *  *  *
AGT TCA TAG CCC ATA TAT GGA GTT CCG CGT TAC ATA ACT TAC GGT AAA
Ser Ser *** Pro Ile Tyr Gly Val Pro Arg Tyr Ile Thr Tyr Gly Lys>
    ___e___e___e___e___e___e___e___CMV PROMOTER___e___e___e___e___e___e___>

    >BglI
    |
    340 350 360 370
    *  *  *  *
TGG CCC GCC TGG CTG ACC GCC CAA CGA CCC CCG CCC ATT GAC GTC AAT
Trp Pro Ala Trp Leu Thr Ala Gln Arg Pro Pro Pro Ile Asp Val Asn>
    ___e___e___e___e___e___e___e___CMV PROMOTER___e___e___e___e___e___e___>

    >AatII
    |
    380 390 400 410 420
    *  *  *  *  *
AAT GAC GTA TGT TCC CAT AGT AAC GCC AAT AGG GAC TTT CCA TTG ACG
Asn Asp Val Cys Ser His Ser Asn Ala Asn Arg Asp Phe Pro Leu Thr>
    ___e___e___e___e___e___e___e___CMV PROMOTER___e___e___e___e___e___e___>

```

FIG. 6

[illegible][illegible]

FIG. 6

(CONTINUED)

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```

      >HincII
      |
      >HpaI
      |
      1100 | 1110 | 1120 | 1130 | 1140 | 1150
      *   *   *   *   *   *   *
      ACCAGAAAGT TAACTGGTAA GTTTAGTCTT TTTGTCTTTT TATTTTCAGGT CCCGGATCCG
      _____b_____ HYBRID SV40 LATE INTRON _____b_____>

      >BseRI
      |
      1160 | 1170 | 1180 | 1190 | 1200 | 1210
      *   *   *   *   *   *   *
      GTGGTGGTGC AAATCAAAGA ACTGCTCCTC AGTGGATGTT GCCTTTACTT CTAGGCCTGT
      _____b_____ HYBRID SV40 LATE INTRON _____b_____>

      >BsiEI
      |
      >EagI
      |
      >EaeI
      |
      >SacII
      |
      >NotI
      |
      1220 | 1230 | 1240 | 1250 | 1260 | 1270
      *   *   *   *   *   *   *
      ACGGAAGTGT TACTTCTGCT CTAAAAGCTG CGGAATTGTA CCCGCGGCCG CTGCAGTCTA
      _____ HYBRID SV40 LATE INTRON _____b_____>

      >ApaI
      |
      >BspDI
      |
      >EcoO109I
      |
      >ApoI
      |
      >EcoRI
      |
      1280 | 1290 | 1300 | 1310 | 1320
      *   *   *   *   *
      GACGAATTCG CGTACGATAT CGATGGGCCC TATT CTA TAG TGT CAC CTA AAT
      Leu *** Cys His Leu Asn>
      _____c_SP6 PROMOTER_____c_____>

      >SacI
      |
      >BanII
      |
      >BsiHKAI
      |
      >Ecl136II
      |
      >BclI
      |
      >BGH_POLY_A
      |
      1330 | 1340 | 1350 | 1360 | 1370 | 1380
      *   *   *   *   *   *
      GCTAGAGC TCGCTGATCA GCCTCGACTG TGCCTTCTAG TTGCCAGCCA TCTGTTGTTT

      >BanI
      |
      1390 | 1400 | 1410 | 1420 | 1430 | 1440
      *   *   *   *   *   *
      GCCCCTCCCC CGTGCCTTCC TTGACCCTGG AAGGTGCCAC TCCCACTGTC CTTTCCTAAT
  
```

FIG. 6
(CONTINUED)

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```

      1450      1460      1470      1480      1490      1500
      *      *      *      *      *      *
AAAATGAGGA AATTGCATCG CATTGTCTGA GTAGGTGTCA TTCTATTCTG GGGGGTGGGG

                                >BbsI
                                |
      1510      1520      1530      1540      1550      1560
      *      *      *      *      *      *
TGGGGCAGGA CAGCAAGGGG GAGGATTGGG AAGACAATAG CCGAAATGAC CGACCAAGCG

                                >BspMI
                                |
                                >BssSI
                                |
      1570      1580      1590      1600      1610      1620
      *      *      *      *      *      *
ACGCCCCAACC TGCCATCACG AGATTTCGAT TCCACCGCCG CCTTCTATGA AAGGTTGGGC

                                >NaeI
                                |
                                >NgoMI
                                ||
                                >BpmI
                                ||
                                >BsrFI
                                ||
      1630      1640      1650      1660      1670      1680
      *      *      *      *      *      *
TTCGGAATCG TTTTCCGGGA CGCCGGCTGG ATGATCCTCC AGCGCCGGGA TCTCATGCTG

                                >BpmI
                                |
                                >SV40_early_poly_A
                                |
      1690      1700      1710      1720      1730      1740
      *      *      *      *      *      *
GAGTTCTTCG CCCACCCCAA CTTGTTTATT GCAGCTTATA ATGGTTACAA ATAAAGCAAT

                                >ApoI
                                |
                                >BsmI
                                |
      1750      1760      1770      1780      1790      1800
      *      *      *      *      *      *
AGCATCACAA ATTCACAAA TAAAGCATTT TTTTCACTGC ATTCTAGTTG TGGTTTGTCC

                                >HincII
                                |
                                >Bst1107I  >AccI
                                ||
                                >AccI  >SalI
                                |||
      1810      1820      1830      1840      1850      1860
      *      *      *      *      *      *
AAACTCATCA ATGTATCTTA TCATGTCTGT ATACCGTCGA CCTCTAGCTA GAGCTTGGCG
_____>

                                >BsrBI
                                |
      1870      1880      1890      1900      1910      1920
      *      *      *      *      *      *
TAATCATGGT CATAGCTGTT TCCTGTGTGA AATTGTTATC CGCTCACAAT TCCACACAAC
_____d_____d_____PUC19 BACKBONE_____d_____d_____>

```

FIG. 6
(CONTINUED)

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```

                                >BamI
                                |
      1930      1940      1950      1960      1970      1980
      *      *      *      *      *      *
ATACGAGCCG GAAGCATAAA GTGTAAAGCC TGGGGTGCCT AATGAGTGAG CTAAC TCACA
_____d_____d_____PUC19 BACKBONE_____d_____d_____>

                                >AseI
                                |
>AseI                                >PvuII                                |
|                                |                                |
      1990      2000      2010      2020      2030      2040
      *      *      *      *      *      *
TTAATTGCGT TGCCTCACT GCCCGCTTTC CAGTCGGGAA ACCTGTCGTG CCAGCTGCAT
_____d_____d_____PUC19 BACKBONE_____d_____d_____>

                                >EaeI                                >HaeII                                >SapI
                                |                                |                                |
      2050      2060      2070      2080      2090      2100
      *      *      *      *      *      *
TAATGAATCG GCCAACGCGC GGGGAGAGGC GGTTCGCGTA TTGGGCGCTC TTCCGCTTCC
_____d_____d_____PUC19 BACKBONE_____d_____d_____>

                                >BsiEI                                >BsrBI
                                |                                |
      2110      2120      2130      2140      2150      2160
      *      *      *      *      *      *
TCGCTCACTG ACTCGCTGCG CTCGGTCGTT CGGCTGCGGC GAGCGGTATC AGCTCACTCA
_____d_____d_____PUC19 BACKBONE_____d_____d_____>

                                >AflIII
                                |
      2170      2180      2190      2200      2210      2220
      *      *      *      *      *      *
AAGGCGGTAA TACGGTTATC CACAGAATCA GGGGATAACG CAGGAAAGAA CATGTGAGCA
_____d_____d_____PUC19 BACKBONE_____d_____d_____>

      2230      2240      2250      2260      2270      2280
      *      *      *      *      *      *
AAAGGCCAGC AAAAGGCCAG GAACCGTAAA AAGGCCGCGT TGCTGGCGTT TTTCCATAGG
_____d_____d_____PUC19 BACKBONE_____d_____d_____>

                                >DrdI
                                |
      2290      2300      2310      2320      2330      2340
      *      *      *      *      *      *
CTCCGCCCCC CTGACGAGCA TCACAAAAT CGACGCTCAA GTCAGAGGTG GCGAAACCCG
_____d_____d_____PUC19 BACKBONE_____d_____d_____>

                                >BssSI
                                |
      2350      2360      2370      2380      2390      2400
      *      *      *      *      *      *
ACAGGACTAT AAAGATACCA GCGGTTTCCC CCTGGAAGCT CCCTCGTGCG CTCTCCTGTT
_____d_____d_____PUC19 BACKBONE_____d_____d_____>

```

FIG. 6
(CONTINUED)

```

                >BsaWI
                |
      2410      2420      2430      2440      2450      2460
      *        *        *        *        *        *
CCGACCCTGC CGCTTACCGG ATACCTGTCC GCCTTTCTCC CTTCGGGAAG CGTGGCGCTT
      d      d      PUC19 BACKBONE      d      d
>

                >SfcI
                |
      2470      2480      2490      2500      2510      2520
      *        *        *        *        *        *
TCTCAATGCT CACGCTGTAG GTATCTCAGT TCGGTGTAGG TCGTTCGCTC CAAGCTGGGC
      d      d      PUC19 BACKBONE      d      d
>

      >BsiHKAI
      |
      >ApaLI |
      |      |
      |      2530      2540      2550      2560      2570      2580
      |      *        *        *        *        |      *        *
TGTGTGCACG AACCCCCCGT TCAGCCCGAC CGCTGCGCCT TATCCGGTAA CTATCGTCTT
      d      d      PUC19 BACKBONE      d      d
>

                                >BsaWI
                                |
                                2590      2600      2610      2620      2630      2640
                                *        *        *        *        *        *
GAGTCCAACC CGGTAAGACA CGACTTATCG CCACTGGCAG CAGCCACTGG TAACAGGATT
      d      d      PUC19 BACKBONE      d      d
>

                                >AlwNI
                                |
                                2590      2600      2610      2620      2630      2640
                                *        *        *        *        *        *
GAGTCCAACC CGGTAAGACA CGACTTATCG CCACTGGCAG CAGCCACTGG TAACAGGATT
      d      d      PUC19 BACKBONE      d      d
>

                >SfcI
                |
      2650      2660      2670      2680      2690      2700
      *        *        *        *        *        *
AGCAGAGCGA GGTATGTAGG CGGTGCTACA GAGTTCTTGA AGTGGTGGCC TAACTACGGC
      d      d      PUC19 BACKBONE      d      d
>

                                >Eco57I
                                |
                                2710      2720      2730      2740      2750      2760
                                *        *        *        *        *        *
TACACTAGAA GGACAGTATT TGGTATCTGC GCTCTGCTGA AGCCAGTTAC CTTCGGAAAA
      d      d      PUC19 BACKBONE      d      d
>

      2770      2780      2790      2800      2810      2820
      *        *        *        *        *        *
AGAGTTGGTA GCTCTTGATC CGGCAAACAA ACCACCGCTG GTAGCGGTGG TTTTTTTGTT
      d      d      PUC19 BACKBONE      d      d
>

      2830      2840      2850      2860      2870      2880
      *        *        *        *        *        *
TGCAAGCAGC AGATTACGCG CAGAAAAAAA GGATCTCAAG AAGATCCTTT GATCTTTTCT
      d      d      PUC19 BACKBONE      d      d
>

                                >BspHI
                                |
                                2890      2900      2910      2920      2930      2940
                                *        *        *        *        *        *
ACGGGGTCTG ACGCTCAGTG GAACGAAAAC TCACGTAAAG GGATTTTGGT CATGAGATTA
      d      d      PUC19 BACKBONE      d      d
>

```

FIG. 6
(CONTINUED)

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```

                                >DraI
                                |
          2950      2960      2970      2980      2990      3000
          *      *      *      *      *      *
TCAAAAAGGA TCTTCACCTA GATCCTTTTA AATTAAAAAT GAAGTTTTAA ATCAATCTAA
          d          d PUC19 BACKBONE d          d >

                                >BanI
                                |
          3010      3020      3030      3040      3050      3060
          *      *      *      *      *      *
AGTATATATG AGTAAACTTG GTCTGACAGT TACCAATGCT TAATCAGTGA GGCACCTATC
          d          d PUC19 BACKBONE d          d >
                                a AMP-ORF a >

                                >AhdI
                                |
          3070      3080      3090      3100      3110      3120
          *      *      *      *      *      *
TCAGCGATCT GTCTATTTTCG TTCATCCATA GTTGCTGAC TCCCGTCGT GTAGATAACT
          a          a AMP-ORF a          a >
          d          d PUC19 BACKBONE d          d >

                                >BsaI
                                |
                                >BsrDI
                                |
                                >BpmI
                                |
          3130      3140      3150      3160      3170      3180
          *      *      *      *      *      *
ACGATACGGG AGGGCTTACC ATCTGGCCCC AGTGCTGCAA TGATACCGCG AGACCCACGC
          a          a AMP-ORF a          a >
          d          d PUC19 BACKBONE d          d >

>BsrFI
|
          3190      3200      3210      3220      3230      3240
          *      *      *      *      *      *
TCACCGGCTC CAGATTTATC AGCAATAAAC CAGCCAGCCG GAAGGGCCGA GCGCAGAAGT
          a          a AMP-ORF a          a >
          d          d PUC19 BACKBONE d          d >

                                >BglI
                                |

                                >AseI
                                |
          3250      3260      3270      3280      3290      3300
          *      *      *      *      *      *
GGTCCTGCAA CTTTATCCGC CTCCATCCAG TCTATTAATT GTTGCCGGGA AGCTAGAGTA
          a          a AMP-ORF a          a >
          d          d PUC19 BACKBONE d          d >

                                >Psp1406I
                                |
                                >FspI
                                |
                                >BsrDI
                                |
                                >SfcI
                                |
                                >MslI
                                |
          3310      3320      3330      3340      3350      3360
          *      *      *      *      *      *
AGTAGTTCGC CAGTTAATAG TTTGCGCAAC GTTGTTGCCA TTGCTACAGG CATCGTGGTG
          a          a AMP-ORF a          a >
          d          d PUC19 BACKBONE d          d >

```

FIG. 6
(CONTINUED)

25/51

>BsaWI
|

3370	3380	3390	3400	3410	3420
*	*	*	*	*	*
TCACGCTCGT	CGTTTGGTAT	GGCTTCATTC	AGCTCCGGTT	CCCAACGATC	AAGGCGAGTT
a	a	AMP-ORF		a	a
d	d	PUC19 BACKBONE		d	d

>BsiEI
|
>PvuI
|

3430	3440	3450	3460	3470	3480
*	*	*	*	*	*
ACATGATCCC	CCATGTTGTG	CAAAAAAGCG	GTTAGCTCCT	TCGGTCCTCC	GATCGTTGTC
a	a	AMP-ORF		a	a
d	d	PUC19 BACKBONE		d	d

>EaeI
|
>MslI
|

3490	3500	3510	3520	3530	3540
*	*	*	*	*	*
AGAAGTAAGT	TGGCCGCAGT	GTTATCACTC	ATGGTTATGG	CAGCACTGCA	TAATTCTCTT
a	a	AMP-ORF		a	a
d	d	PUC19 BACKBONE		d	d

>ScaI
|

3500	3560	3570	3580	3590	3600
*	*	*	*	*	*
ACTGTCATGC	CATCCGTAAG	ATGCTTTTCT	GTGACTGGTG	AGTACTCAAC	CAAGTCATTC
a	a	AMP-ORF		a	a
d	d	PUC19 BACKBONE		d	d

>BsiEI
|

3610	3620	3630	3640	3650	3660
*	*	*	*	*	*
TGAGAATAGT	GTATGCGGCG	ACCGAGTTGC	TCTTGCCCGG	CGTCAATACG	GGATAATACC
a	a	AMP-ORF		a	a
d	d	PUC19 BACKBONE		d	d

>Psp1406I
|
>DraI >BsiHKAI >XmnI
| | |

3670	3680	3690	3700	3710	3720
*	*	*	*	*	*
GCGCCACATA	GCAGAACTTT	AAAAGTGCTC	ATCATTGGAA	AACGTTCTTC	GGGGCGAAAA
a	a	AMP-ORF		a	a
d	d	PUC19 BACKBONE		d	d

>ApaLI
|
>Eco57I
|
>BssSI >BsiHKAI
| | |

3730	3740	3750	3760	3770	3780
*	*	*	*	*	*
CTCTCAAGGA	TCTTACCGCT	GTTGAGATCC	AGTTCGATGT	AACCCACTCG	TGCACCCAAC
a	a	AMP-ORF		a	a
d	d	PUC19 BACKBONE		d	d

FIG. 6
(CONTINUED)

```

      3790      3800      3810      3820      3830      3840
      *      *      *      *      *      *
TGATCTTCAG CATCTTTTAC TTTCACCAGC GTTTCTGGGT GAGCAAAAAC AGGAAGGCAA
      a      a      AMP-ORF      a      a      >
      d      d      PUC19 BACKBONE      d      d      >

                                >MslI                                >EarI
                                |                                |
      3850      3860      3870      3880      3890      3900
      *      *      *      *      *      *
AATGCCGCAA AAAAGGGAAT AAGGGCGACA CGGAAATGTT GAATACTCAT ACTCTTCCTT
      a      a      AMP-ORF      a      >
      d      d      PUC19 BACKBONE      d      d      >

      >SspI                                >BspHI      >BsrBI
      |                                |      |
      3910      3920      3930      3940      3950      3960
      *      *      *      *      *      *
TTTCAATATT ATTGAAGCAT TTATCAGGGT TATTGTCTCA TGAGCGGATA CATATTTGAA
      d      d      PUC19 BACKBONE      d      d      >

      3970      3980      3990      4000      4010      4020
      *      *      *      *      *      *
TGTATTTAGA AAAATAAACA AATAGGGGTT CCGCGCACAT TTCCCCGAAA AGTGCCACCT
      d      d      PUC19 BACKBONE      d      d      >

>HincII
|
>AatII
||
>AccI
||
>SalI
|||
|*|
GACGTC
_____>

```

FIG. 6
(CONTINUED)

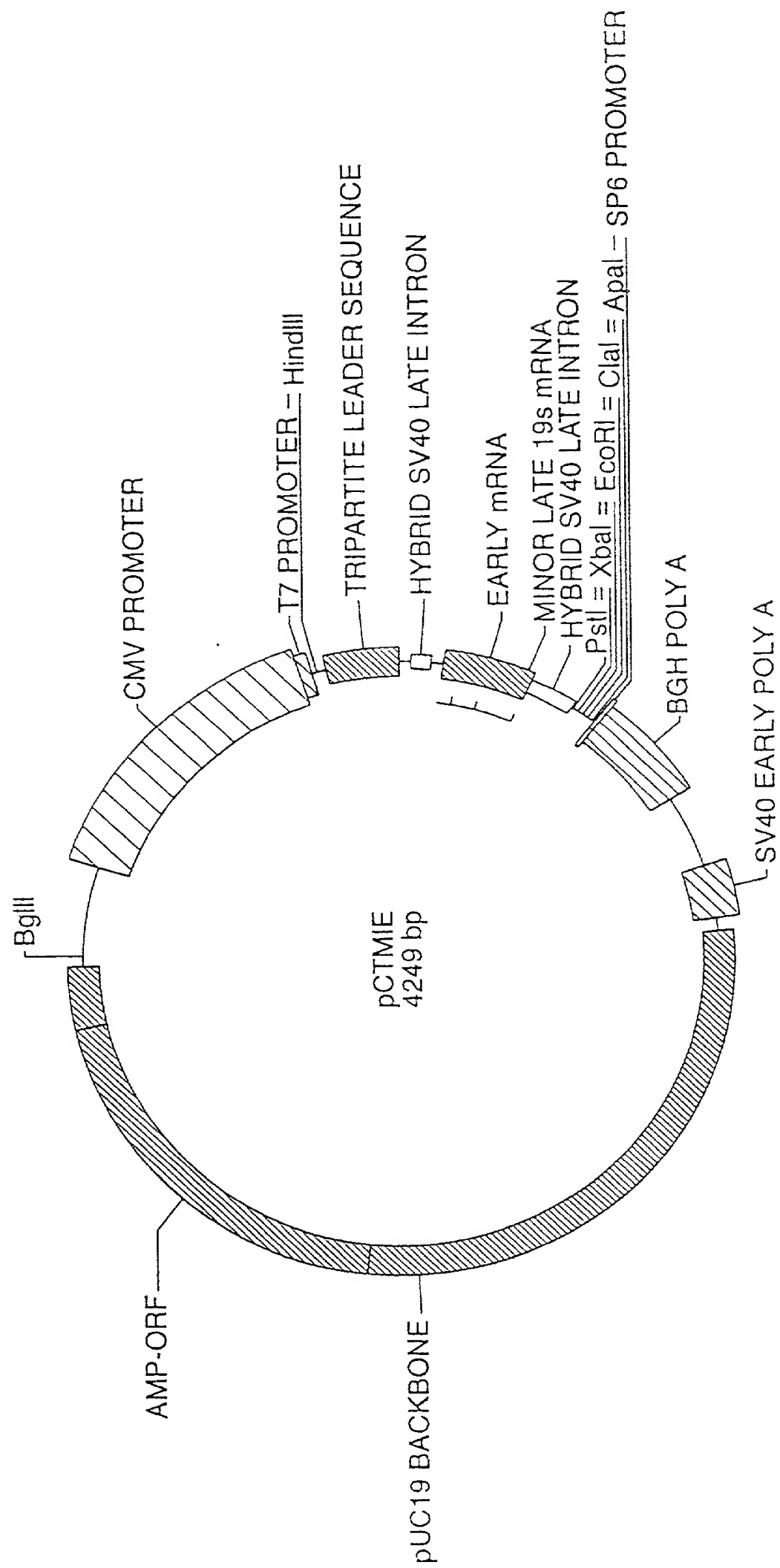


FIG. 7

28/51

```

                                >HincII
                                |
                                >AccI
                                ||
                                >Sali
                                |||
      >BglIII
      |
10  *  |  *  20  *  30  |||  *  40  *  50  *  60  *
GACGGATCGG GAGATCTCCC GATCCCCTAT GGTCGACTCT CAGTACAATC TGCTCTGATG

                                >AlwNI
                                |
70  *  80  *  90  *  100  *  110  *  120  *
CCGCATAGTT AAGCCAGTAT CTGCTCCCTG CTTGTGTGTT GGAGGTCGCT GAGTAGTGCG

                                >ApoI
                                |
130 *  140 *  150 *  160 *  170 *  180 *
CGAGCAAAAT TTAAGCTACA ACAAGGCAAG GCTTGACCGA CAATTGCATG AAGAATCTGC

                                >MfeI
                                |
                                >HincII
                                |
                                >AflIII
                                |
                                >MluI
                                |
190 *  200 *  210 *  220 *  230 *
TTAGGGTTAG GCGTTTTGCG CTGCTTCG CGA TGT ACG GGC CAG ATA TAC GCG TTG
Arg Cys Thr Gly Gln Ile Tyr Ala Leu>
__f__f__CMV PROMOTER__f__f__>

                                >SpeI
                                |
240 *  250 *  260 *  270 *  280 *
ACA TTG ATT ATT GAC TAG TTA TTA ATA GTA ATC AAT TAC GGG GTC ATT
Thr Leu Ile Ile Asp *** Leu Leu Ile Val Ile Asn Tyr Gly Val Ile>
__f__f__f__f__f__f__CMV PROMOTER__f__f__f__f__f__f__>

                                >AseI
                                |
290 *  300 *  310 *  320 *  330 *
AGT TCA TAG CCC ATA TAT GGA GTT CCG CGT TAC ATA ACT TAC GGT AAA
Ser Ser *** Pro Ile Tyr Gly Val Pro Arg Tyr Ile Thr Tyr Gly Lys>
__f__f__f__f__f__f__CMV PROMOTER__f__f__f__f__f__f__>

                                >BglI
                                |
340 *  350 *  360 *  370 *
TGG CCC GCC TGG CTG ACC GCC CAA CGA CCC CCG CCC ATT GAC GTC AAT
Trp Pro Ala Trp Leu Thr Ala Gln Arg Pro Pro Pro Ile Asp Val Asn>
__f__f__f__f__f__f__CMV PROMOTER__f__f__f__f__f__f__>

                                >AatII
                                |
380 *  390 *  400 *  410 *  420 *
AAT GAC GTA TGT TCC CAT AGT AAC GCC AAT AGG GAC TTT CCA TTG ACG
Asn Asp Val Cys Ser His Ser Asn Ala Asn Arg Asp Phe Pro Leu Thr>
__f__f__f__f__f__f__CMV PROMOTER__f__f__f__f__f__f__>

```

FIG. 8

[illegible][illegible]


```

_____TRIPARTITE LEADER SEQUENCE_____g_____>
      |
      |>HpaI
      |>HincII
      |
1100 1110 1120 1130 1140 1150
*   *   *   *   *   *
ACCAGAAAGT TAACTGGTAA GTTTAGTCTT TTTGTCTTTT TATTTTCAGGT CCCGGATCTG
_____b_____HYBRID SV40 LATE INTRON_____b_____>
      |
      |>PpuMI
      |>EcoO109I
      |
1160 1170 1180 1190 1200 1210
*   *   *   *   *   *
AGTTAGGGCG GGACATGGGC GGAGTTAGGG GCGGGACTAT GGTGCTGAC TAATTGAGAT
      |
      |>Ppu10I
      |>21_bp_tandem_repeat_III_[110],[102],[112]
      |
1160 1170 1180 1190 1200 1210
*   *   *   *   *   *
AGTTAGGGCG GGACATGGGC GGAGTTAGGG GCGGGACTAT GGTGCTGAC TAATTGAGAT
      |
      |>SphI
      |>NsiI
      |
      |<72_bp_tandem_repeat_enhancer_sequence_
      |
1220 1230 1240 1250 1260 1270
*   *   *   *   *   *
GCATGCTTTG CATACTTCTG CCTGCTGGGG AGCCTGGGGA CTTTCCACAC CTGGTTGCTG
<_____h_____h_____EARLY MRNA_____h_____h_____>
      |
      |>NsiI
      |>Ppu10I
      |>SphI
      |
1280 1290 1300 1310 1320 1330
*   *   *   *   *   *
ACTAATTGAG ATGCATGCTT TGCATACTTC TGCCTGCTGG GGAGCCTGGG GACTTTCCAC
<_____h_____h_____EARLY MRNA_____h_____h_____>
      |
      |>PvuII
      |>BsaWI
      |>BseRI
      |
<72_bp_tandem_repeat_enhancer_sequence_B_
      |
      |<T_antigen_binding_site_II_
      |
1340 1350 1360 1370 1380 1390
*   *   *   *   *   *
ACCCTAACTG ACACACATTC CACAGCTGGT TCTTTCAGAT CCGGTGGTGG TGCAAATCAA
      |
      |_____HYBRID SV40_____>
<_____h_____EARLY MRNA_____h_____
      |
      |_____MINOR LATE 19S_____>
      |
      |>StuI
      |
1400 1410 1420 1430 1440 1450
*   *   *   *   *   *
AGAACTGCTC CTCAGTGGAT GTTGCCTTTA CTTCTAGGCC TGTACGGAAG TGTTACTTCT
_____c_____HYBRID SV40 LATE INTRON_____c_____>

```

FIG. 8
(CONTINUED)

[illegible][illegible]

```

      >NaeI
      |
    >BpmI
      |
    >BsrFI
      |
    NgoMI
      |
1870  * 1880  * 1890  * 1900  * 1910  * 1920  *
GGACGCCGGC TGGATGATCC TCCAGCGCGG GGATCTCATG CTGGAGTTCT TCGCCCACCC

    >BpmI
    |
>SV40_early_poly_A
    |
1930  * 1940  * 1950  * 1960  * 1970  * 1980  *
CAACTTGTTT ATTGCAGCTT ATAATGGTTA CAAATAAAGC AATAGCATCA CAAATTCAC

      >BsmI
      |
1990  * 2000  * 2010  * 2020  * 2030  * 2040  *
AAATAAAGCA TTTTTCAC TGCATTCTAG TTGTGGTTTG TCCAAACTCA TCAATGTATC

      >HincII
      |
    >Bst1107I  >AccI
      |      |
    >AccI  >SalI
      ||  ||
2050  * 2060  * 2070  * 2080  * 2090  * 2100  *
TTATCATGTC TGTATACCGT CGACCTCTAG CTAGAGCTTG GCGTAATCAT GGTCATAGCT
      PUC19 BACKBONE
      >

      >BsrBI
      |
2110  * 2120  * 2130  * 2140  * 2150  * 2160  *
GTTTCCTGTG TGAAATTGTT ATCCGCTCAC AATTCCACAC AACATACGAG CCGGAAGCAT
      e PUC19 BACKBONE e

      >BanI
      |
2170  * 2180  * 2190  * 2200  * 2210  * 2220  *
AAAGTGTAAG GCCTGGGGTG CCTAATGAGT GAGCTAACTC ACATTAATTG CGTTGCGCTC
      e PUC19 BACKBONE e

      >PvuII  >AseI  >EaeI
      |      |      |
2230  * 2240  * 2250  * 2260  * 2270  * 2280  *
ACTGCCCCGCT TTCCAGTCGG GAAACCTGTC GTGCCAGCTG CATTAATGAA TCGGCCAACG
      e PUC19 BACKBONE e

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FIG. 8
(CONTINUED)

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                                >SapI
                                |
                        >HaeII  >EarI
                        |      |
                2290      2300      2310 | 2320      2330      2340
                *      *      *      * | *      *      *      *
CGCGGGGAGA GGCGGTTTGC GTATTGGGCG CTCTTCCGCT TCCTCGCTCA CTGACTCGCT
_____e_____e_____PUC19 BACKBONE_____e_____e_____>

                >BsiEI                >BsrBI
                |                        |
                2350      2360      | 2370      2380      2390      2400
                *      *      *      * | *      *      *      *
GCGCTCGGTC GTTCGGCTGC GGCGAGCGGT ATCAGCTCAC TCAAAGGCGG TAATACGGTT
_____e_____e_____PUC19 BACKBONE_____e_____e_____>

                                >AflIII
                                |
                2410      2420      2430 | 2440      2450      2460
                *      *      *      * | *      *      *      *
ATCCACAGAA TCAGGGGATA ACGCAGGAAA GAACATGTGA GCAAAAGGCC AGCAAAAGGC
_____e_____e_____PUC19 BACKBONE_____e_____e_____>

                2470      2480      2490      2500      2510      2520
                *      *      *      *      *      *      *
CAGGAACCGT AAAAAGGCCG CGTTGCTGGC GTTTTTCCAT AGGCTCCGCC CCCCTGACGA
_____e_____e_____PUC19 BACKBONE_____e_____e_____>

                                >DrdI
                                |
                2530      2540 | 2550      2560      2570      2580
                *      *      * | *      *      *      *
GCATCACAAA AATCGACGCT CAAGTCAGAG GTGGCGAAAC CCGACAGGAC TATAAAGATA
_____e_____e_____PUC19 BACKBONE_____e_____e_____>

                                >BssSI                >BsaWI
                                |                        |
                2590      2600      | 2610      2620      2630      2640
                *      *      *      * | *      *      *      *
CCAGGCGTTT CCCCCTGGAA GCTCCCTCGT GCGCTCTCCT GTTCCGACCC TGCCGCTTAC
_____e_____e_____PUC19 BACKBONE_____e_____e_____>

                                >HaeII                >SfcI
                                |                        |
                2650      2660      2670      2680 | 2690      2700
                *      *      *      *      * | *      *      *
CGGATACCTG TCCGCCTTTC TCCCTTCGGG AAGCGTGGCG CTTTCTCAAT GCTCAGCTG
_____e_____e_____PUC19 BACKBONE_____e_____e_____>

                                >BsiHKAI
                                |
                                >ApaLI
                                |
                2710      2720      2730      2740      2750 | 2760
                *      *      *      *      *      * | *      *
TAGGTATCTC AGTTCGGTGT AGGTCGTTTC CTCCAAGCTG GGCTGTGTGC ACGAACCCCC
_____e_____e_____PUC19 BACKBONE_____e_____e_____>

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FIG. 8
(CONTINUED)

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      >BsiEI          >BsaWI
      |              |
      2770          2780          2790          2800          2810          2820
      *            *            *            *            *            *
CGTTCAGCCC GACCGCTGCG CCTTATCCGG TAACTATCGT CTTGAGTCCA ACCCGGTAAG
      e            e            PUC19 BACKBONE e            e            >

      >AlwNI
      |
      2830          2840          2850          2860          2870          2880
      *            *            *            *            *            *
ACACGACTTA TCGCCACTGG CAGCAGCCAC TGGTAACAGG ATTAGCAGAG CGAGGTATGT
      e            e            PUC19 BACKBONE e            e            >

      >SfcI
      |
      2890          2900          2910          2920          2930          2940
      *            *            *            *            *            *
AGGCGGTGCT ACAGAGTTCT TGAAGTGGTG GCCTAACTAC GGCTACACTA GAAGGACAGT
      e            e            PUC19 BACKBONE e            e            >

      >Eco57I
      |
      2950          2960          2970          2980          2990          3000
      *            *            *            *            *            *
ATTTGGTATC TGCGCTCTGC TGAAGCCAGT TACCTTCGGA AAAAGAGTTG GTAGCTCTTG
      e            e            PUC19 BACKBONE e            e            >

      3010          3020          3030          3040          3050          3060
      *            *            *            *            *            *
ATCCGGCAAA CAAACCACCG CTGGTAGCGG TGGTTTTTTT GTTTGCAAGC AGCAGATTAC
      e            e            PUC19 BACKBONE e            e            >

      3070          3080          3090          3100          3110          3120
      *            *            *            *            *            *
GCGCAGAAAA AAAGGATCTC AAGAAGATCC TTTGATCTTT TCTACGGGGT CTGACGCTCA
      e            e            PUC19 BACKBONE e            e            >

      >BspHI
      |
      3130          3140          3150          3160          3170          3180
      *            *            *            *            *            *
GTGGAACGAA AACTCACGTT AAGGGATTTT GGTCATGAGA TTATCAAAAA GGATCTTCAC
      e            e            PUC19 BACKBONE e            e            >

      >DraI          >DraI
      |              |
      3190          3200          3210          3220          3230          3240
      *            *            *            *            *            *
CTAGATCCTT TTAAATTAAA AATGAAGTTT TAAATCAATC TAAAGTATAT ATGAGTAAAC
      e            e            PUC19 BACKBONE e            e            >

      >BanI
      |
      3250          3260          3270          3280          3290          3300
      *            *            *            *            *            *
TTGGTCTGAC AGTTACCAAT GCTTAATCAG TGAGGCACCT ATCTCAGCGA TCTGTCTATT
      e            a            a AMP-ORF a            a            >
      e            e            PUC19 BACKBONE e            e            >

```

FIG. 8
(CONTINUED)

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```

>AhdI
      |
3310      3320      3330      3340      3350      3360
*      *      *      *      *      *
TCGTTTCATCC ATAGTTGCCT GACTCCCCGT CGTGTAAGATA ACTACGATAC GGGAGGGCTT
      a      a      AMP-ORF      a      a      >
      e      e      PUC19 BACKBONE      e      e      >

>BsaI
      |
      |>BsrDI      >BpmI      >BsrFI
      |      |      |
3370      3380      3390      3400      3410      3420
*      *      *      *      *      *
ACCATCTGGC CCCAGTGCTG CAATGATACC GCGAGACCCA CGCTCACCGG CTCCAGATTT
      a      a      AMP-ORF      a      a      >
      e      e      PUC19 BACKBONE      e      e      >

>BglI
      |
3430      3440      3450      3460      3470      3480
*      *      *      *      *      *
ATCAGCAATA AACCAGCCAG CCGGAAGGGC CGAGCGCAGA AGTGGTCCTG CAACTTTATC
      a      a      AMP-ORF      a      a      >
      e      e      PUC19 BACKBONE      e      e      >

>AseI
      |
3490      3500      3510      3520      3530      3540
*      *      *      *      *      *
CGCCTCCATC CAGTCTATTA ATTGTTGCCG GGAAGCTAGA GTAAGTAGTT CGCCAGTTAA
      a      a      AMP-ORF      a      a      >
      e      e      PUC19 BACKBONE      e      e      >

>Psp1406I
      |
      |>FspI      >BsrDI      >SfcI      >MslI
      |      |      |      |
3550      3560      3570      3580      3590      3600
*      *      *      *      *      *
TAGTTTGCGC AACGTTGTTG CCATTGCTAC AGGCATCGTG GTGTCACGCT CGTCGTTTGG
      a      a      AMP-ORF      a      a      >
      e      e      PUC19 BACKBONE      e      e      >

>BsaWI
      |
3610      3620      3630      3640      3650      3660
*      *      *      *      *      *
TATGGCTTCA TTCAGCTCCG GTTCCCAACG ATCAAGGCGA GTTACATGAT CCCCCATGTT
      a      a      AMP-ORF      a      a      >
      e      e      PUC19 BACKBONE      e      e      >

>BsiEI
      |
      |>PvuI      >EaeI
      |      |
3670      3680      3690      3700      3710      3720
*      *      *      *      *      *
GTGCAAAAAA GCGGTTAGCT CCTTCGGTCC TCCGATCGTT GTCAGAAGTA AGTTGGCCGC
      a      a      AMP-ORF      a      a      >
      e      e      PUC19 BACKBONE      e      e      >

```

FIG. 8
(CONTINUED)

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                >MslI
                |
      3730      3740      3750      3760      3770      3780
      *      *      *      *      *      *
AGTGTTATCA CTCATGGTTA TGGCAGCACT GCATAATTCT CTTACTGTCA TGCCATCCGT
      a      a      AMP-ORF      a      a      >
      e      e      PUC19 BACKBONE      e      e      >

                >ScaI
                |
      3790      3800      3810      3820      3830      3840
      *      *      *      *      *      *
AAGATGCTTT TCTGTGACTG GTGAGTACTC AACCAAGTCA TTCTGAGAAT AGTGTATGCG
      a      a      AMP-ORF      a      a      >
      e      e      PUC19 BACKBONE      e      e      >

      >BsiEI
      |
      3850      3860      3870      3880      3890      3900
      *      *      *      *      *      *
GCGACCGAGT TGCTCTTGCC CGGCGTCAAT ACGGGATAAT ACCGCGCCAC ATAGCAGAAC
      a      a      AMP-ORF      a      a      >
      e      e      PUC19 BACKBONE      e      e      >

      >Psp1406I
      |
>DraI      >BsiHKAI      >XmnI
      |      |      |
      3910      3920      3930      3940      3950      3960
      *      *      *      *      *      *
TTTAAAAGTG CTCATCATTG GAAAACGTTT TTCGGGGCGA AAACCTCTCA GGATCTTACC
      a      a      AMP-ORF      a      a      >
      e      e      PUC19 BACKBONE      e      e      >

                >Eco57I
                |
                >ApaLI
                |
                >BssSI      >BsiHKAI
                |      |
      3970      3980      3990      4000      4010      4020
      *      *      *      *      *      *
GCTGTTGAGA TCCAGTTCGA TGTAACCCAC TCGTGCACCC AACTGATCTT CAGCATCTTT
      a      a      AMP-ORF      a      a      >
      e      e      PUC19 BACKBONE      e      e      >

      4030      4040      4050      4060      4070      4080
      *      *      *      *      *      *
TACTTTCACC AGCGTTTCTG GGTGAGCAAA AACAGGAAGG CAAAATGCCG CAAAAAAGGG
      a      a      AMP-ORF      a      a      >
      e      e      PUC19 BACKBONE      e      e      >

      >MslI      >EarI      >SspI
      |      |      |
      4090      4100      4110      4120      4130      4140
      *      *      *      *      *      *
AATAAGGGCG ACACGGAAAT GTTGAATACT CATACTCTTC CTTTTTCAAT ATTATTGAAG
      a      a      AMP-ORF      a      a      >
      e      e      PUC19 BACKBONE      e      e      >

```

FIG. 8
(CONTINUED)

```

                >BspHI   >BsrBI
                |       |
      4150      4160 | 4170      4180      4190      4200
      *         *   *   *         *         *         *
CATTTCATCAG GGTTCATTGTC TCATGAGCGG ATACATATTT GAATGTATTT AGAAAAATAA
_____e_____e_____PUC19 BACKBONE_____e_____e_____>

                                >HincII
                                |
                                >AccI
                                ||
                                >AatII
                                ||
                                >SalI
                                |||
      4210      4220      4230      4240
      *         *         *         *         *
ACAAATAGGG GTTCCGCGCA CATTTCCTCG AAAAGTGCCA CCTGACGTC
_____e_____PUC19 BACKBONE_____e_____>

```

FIG. 8
(CONTINUED)

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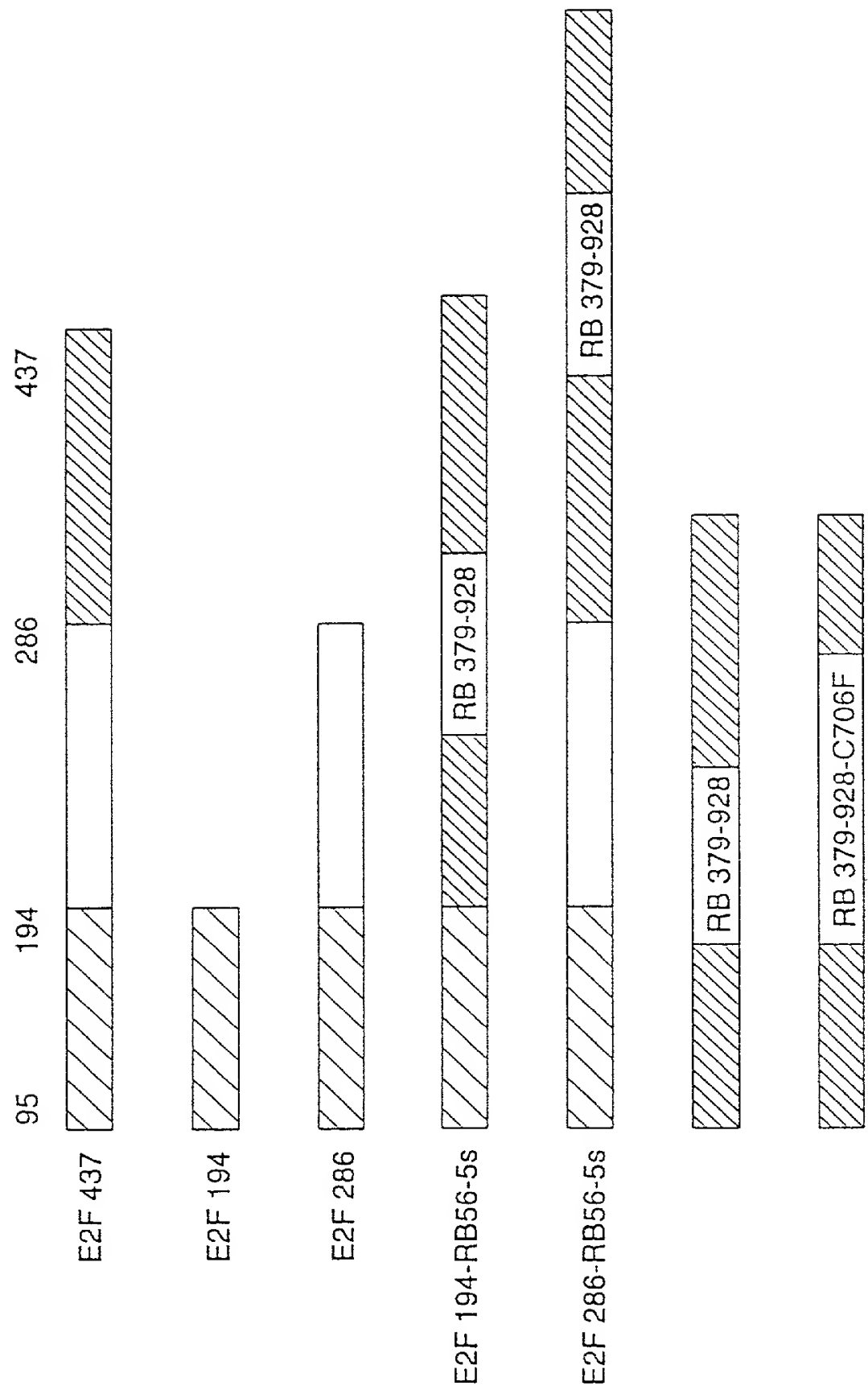


FIG. 9

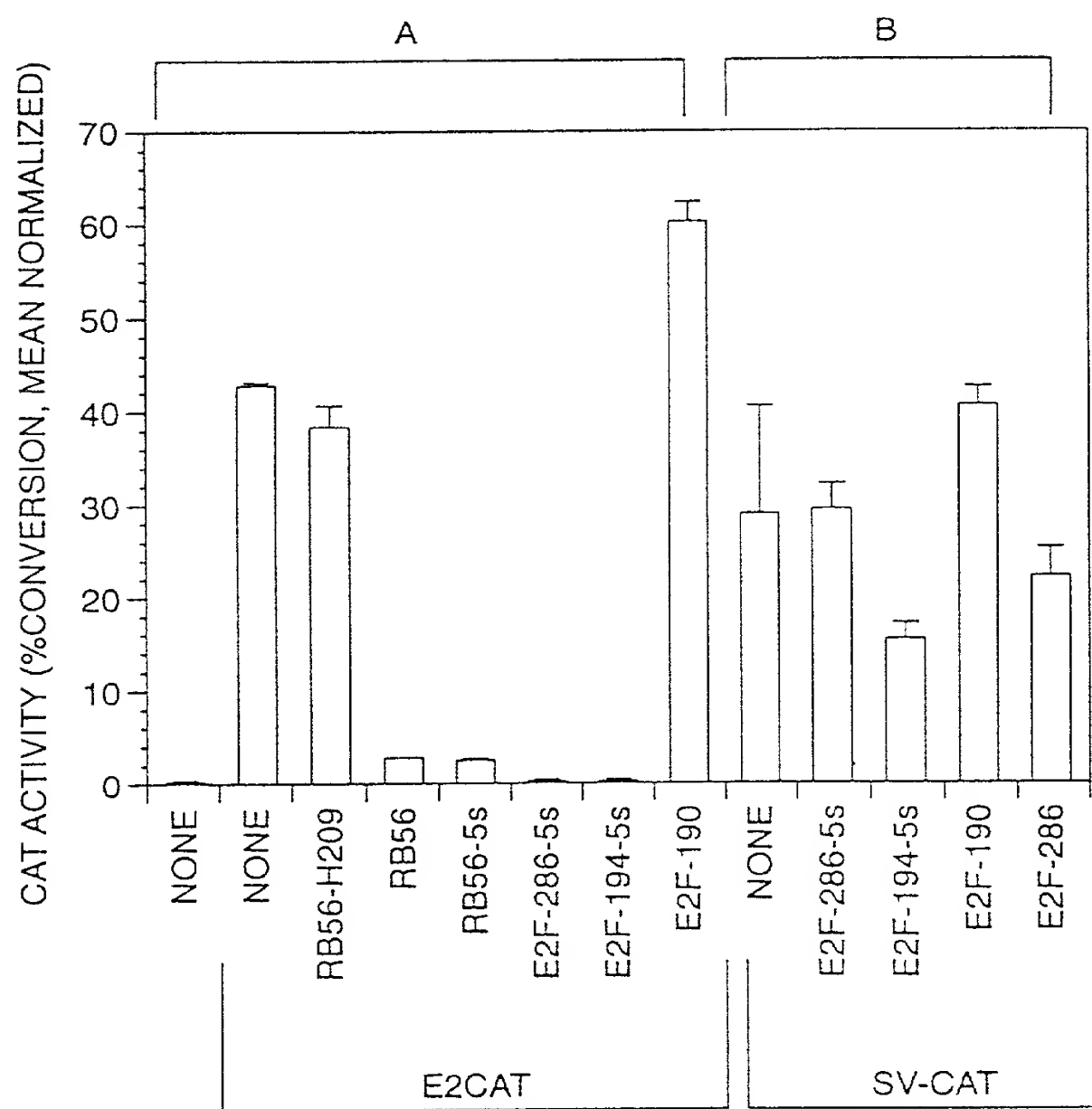


FIG. 10

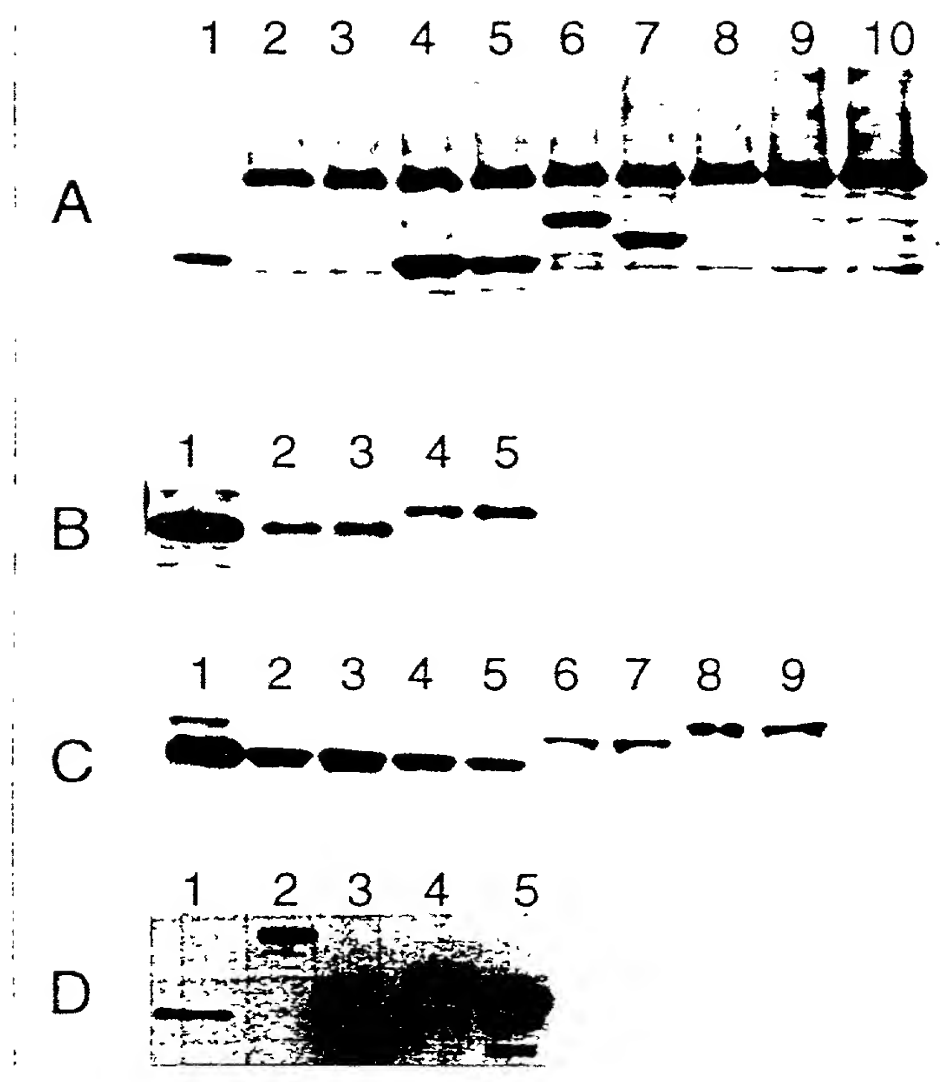


FIG. 11

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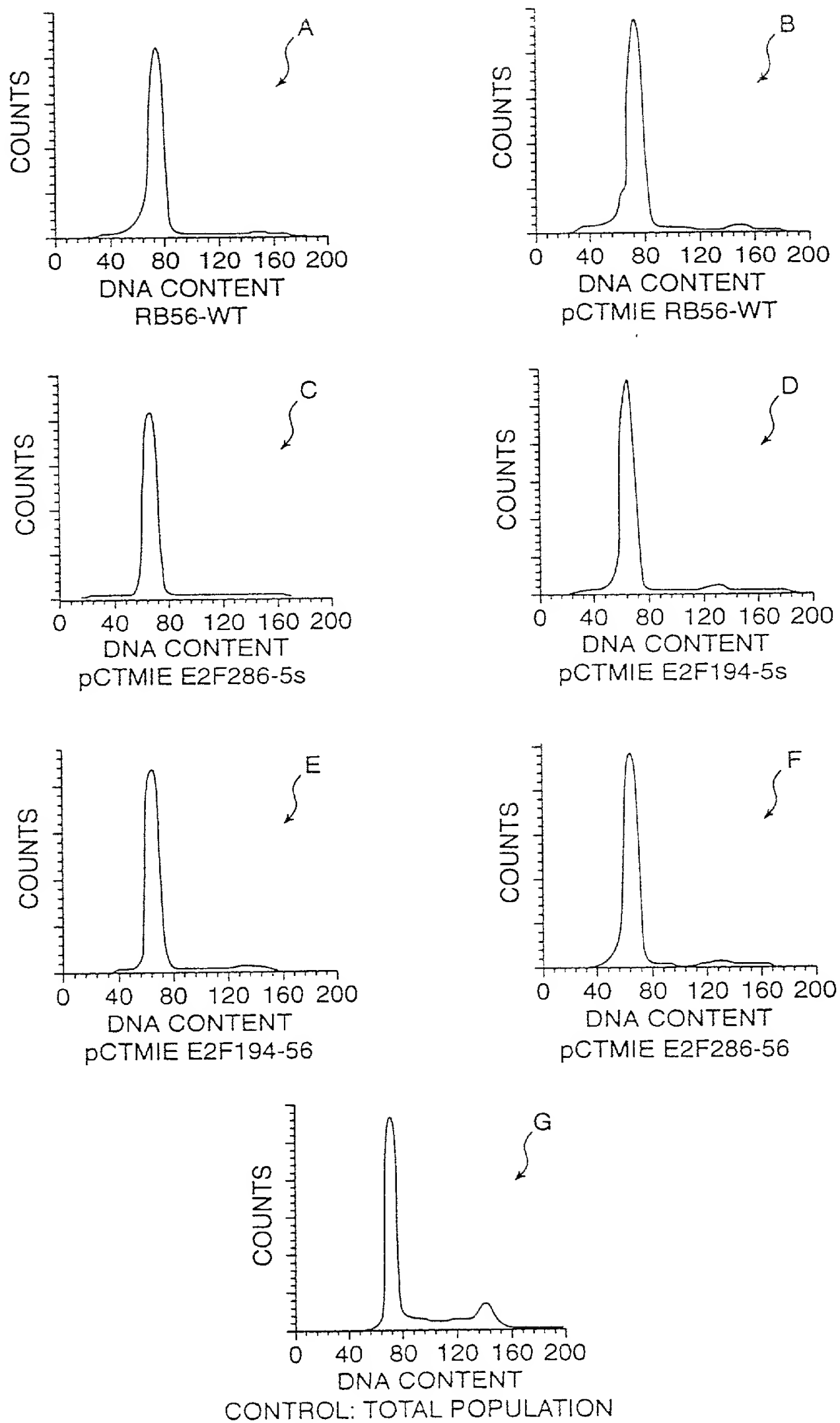


FIG. 12

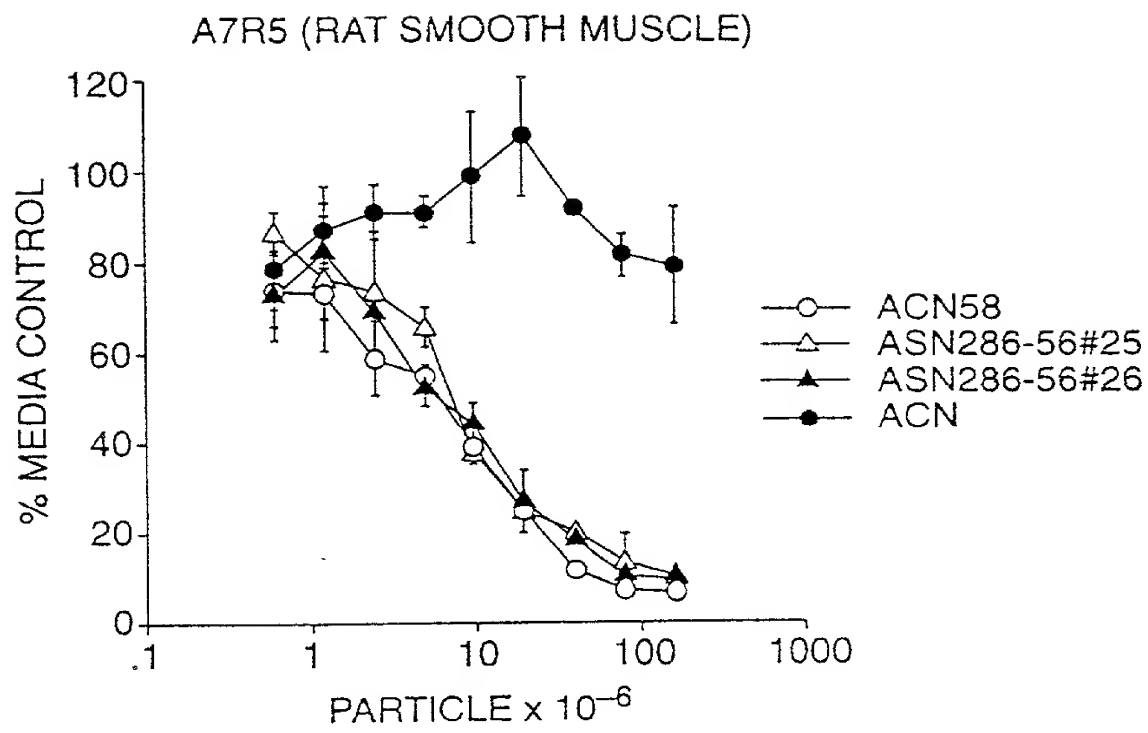


FIG. 13A

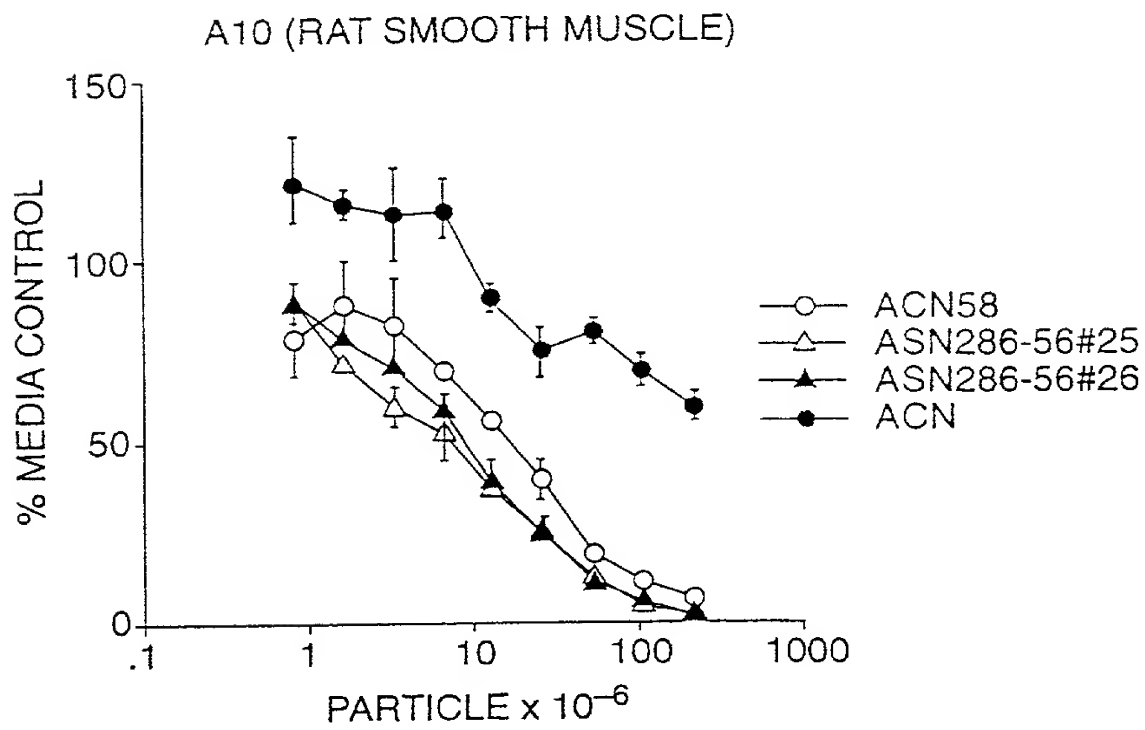


FIG. 13B

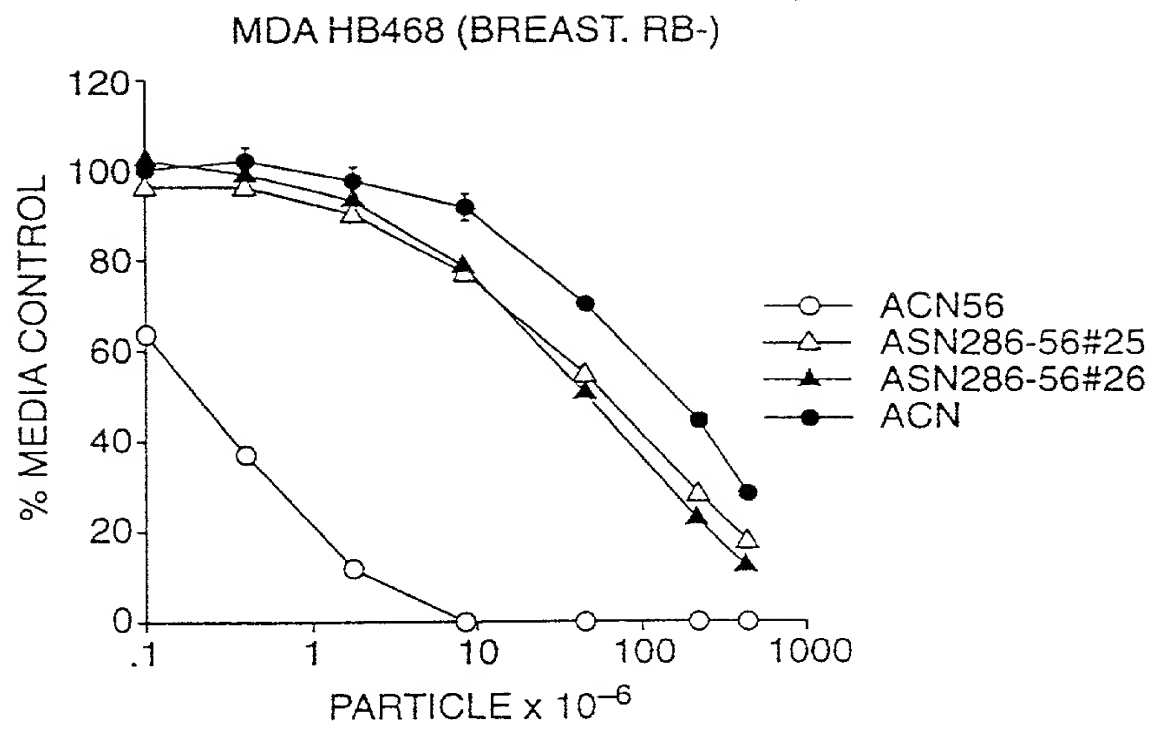


FIG. 14A

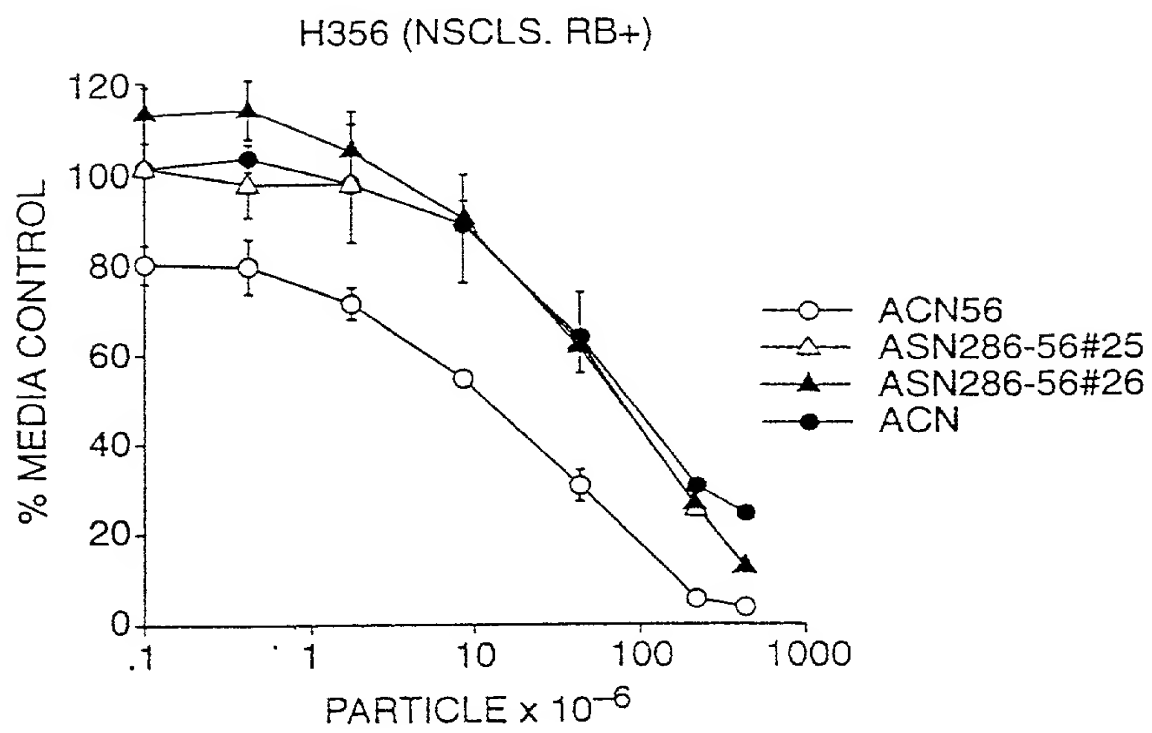
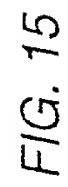
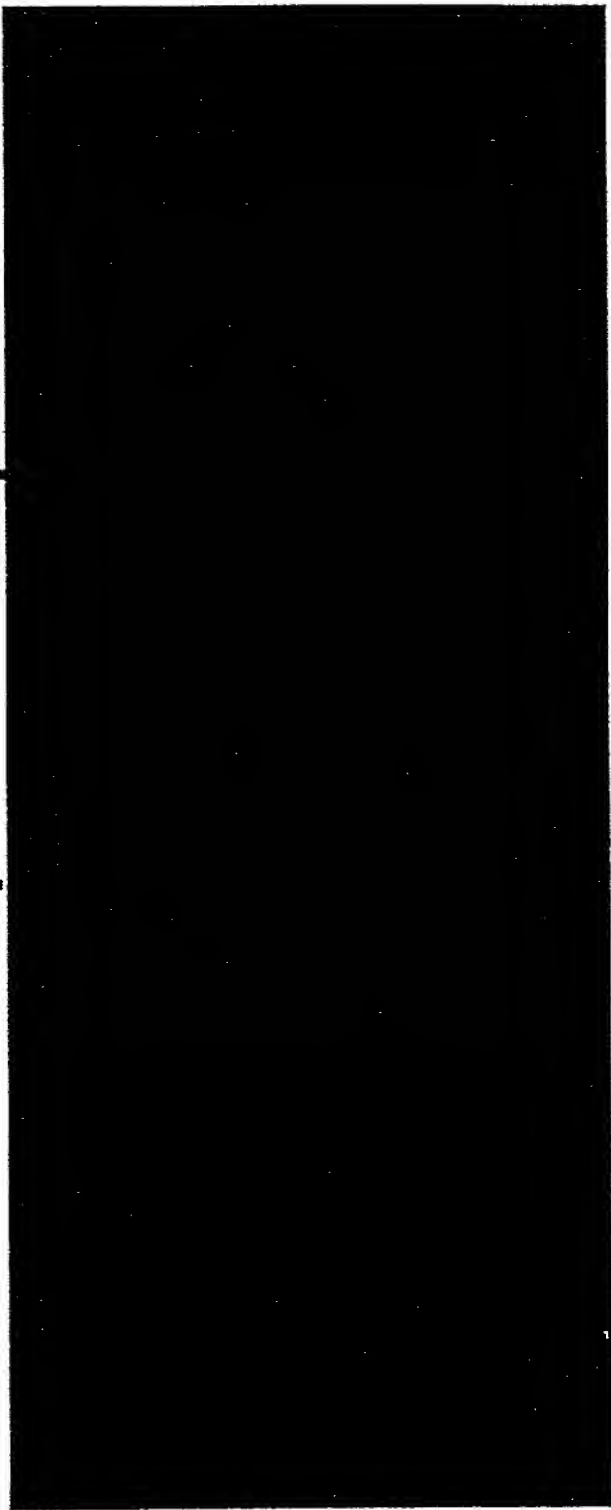


FIG. 14B



MB468 (BREAST) | A7R5 (SM) | A10 (SM)



UN 50 250 500 UN 50 250 UN 100 500

FIG. 16

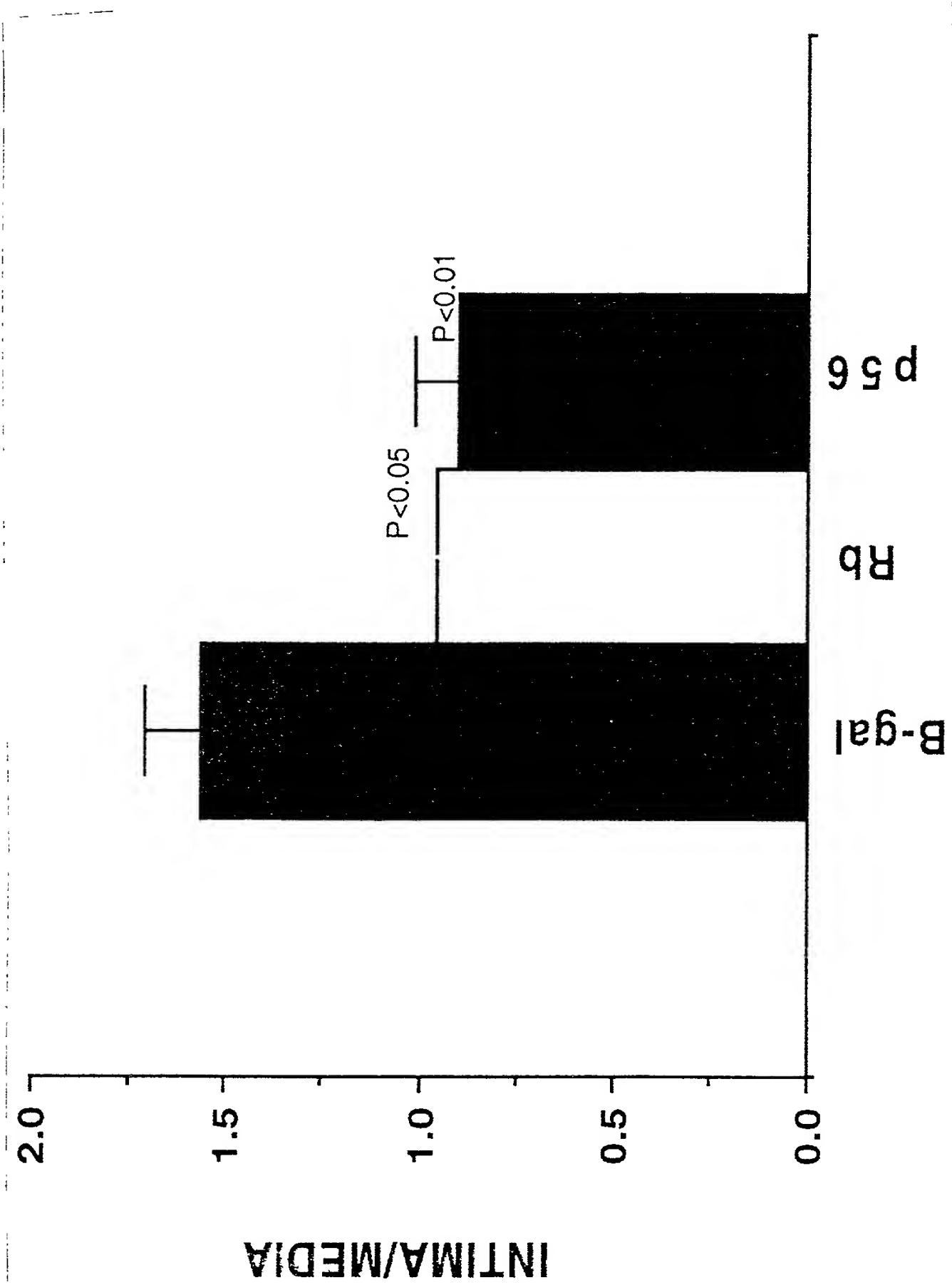
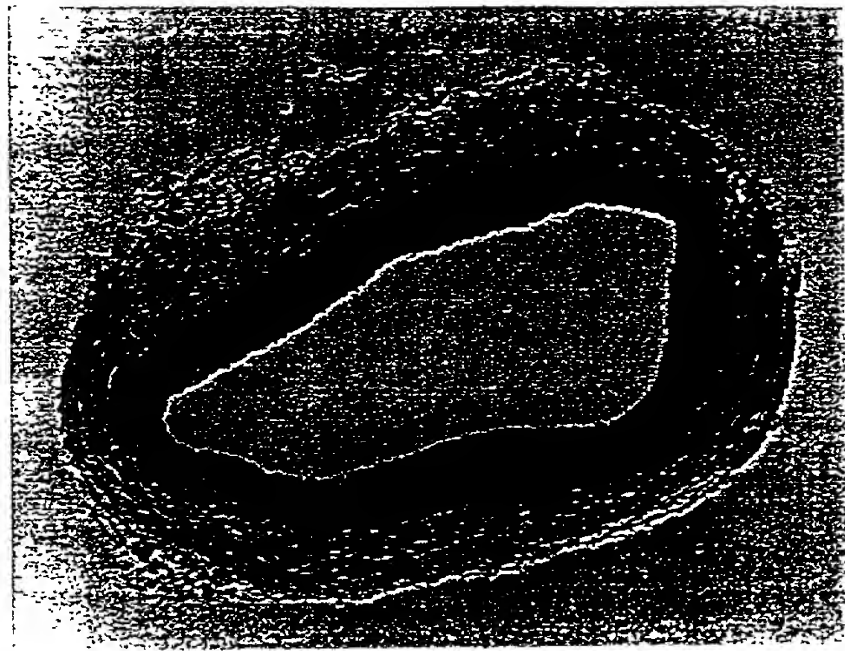
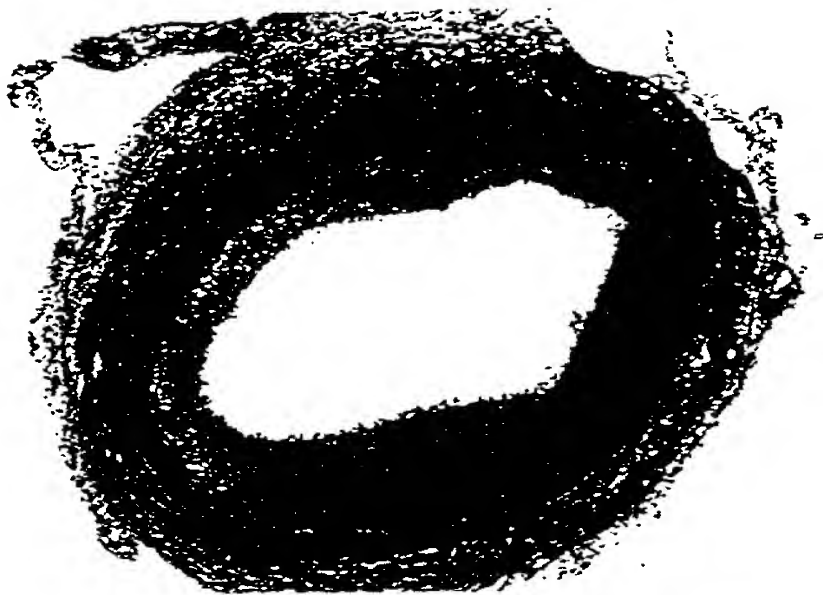


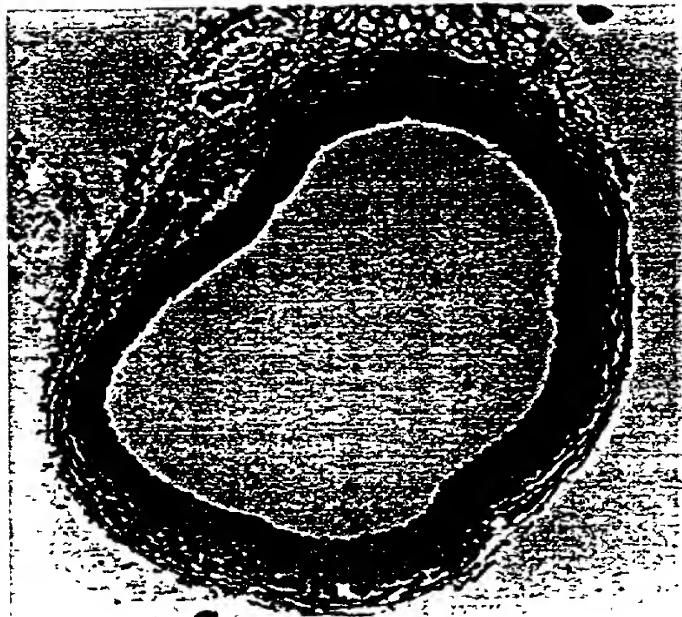
FIG. 17



p56^{RB}-Treated



Restenotic



Normal

FIG. 18

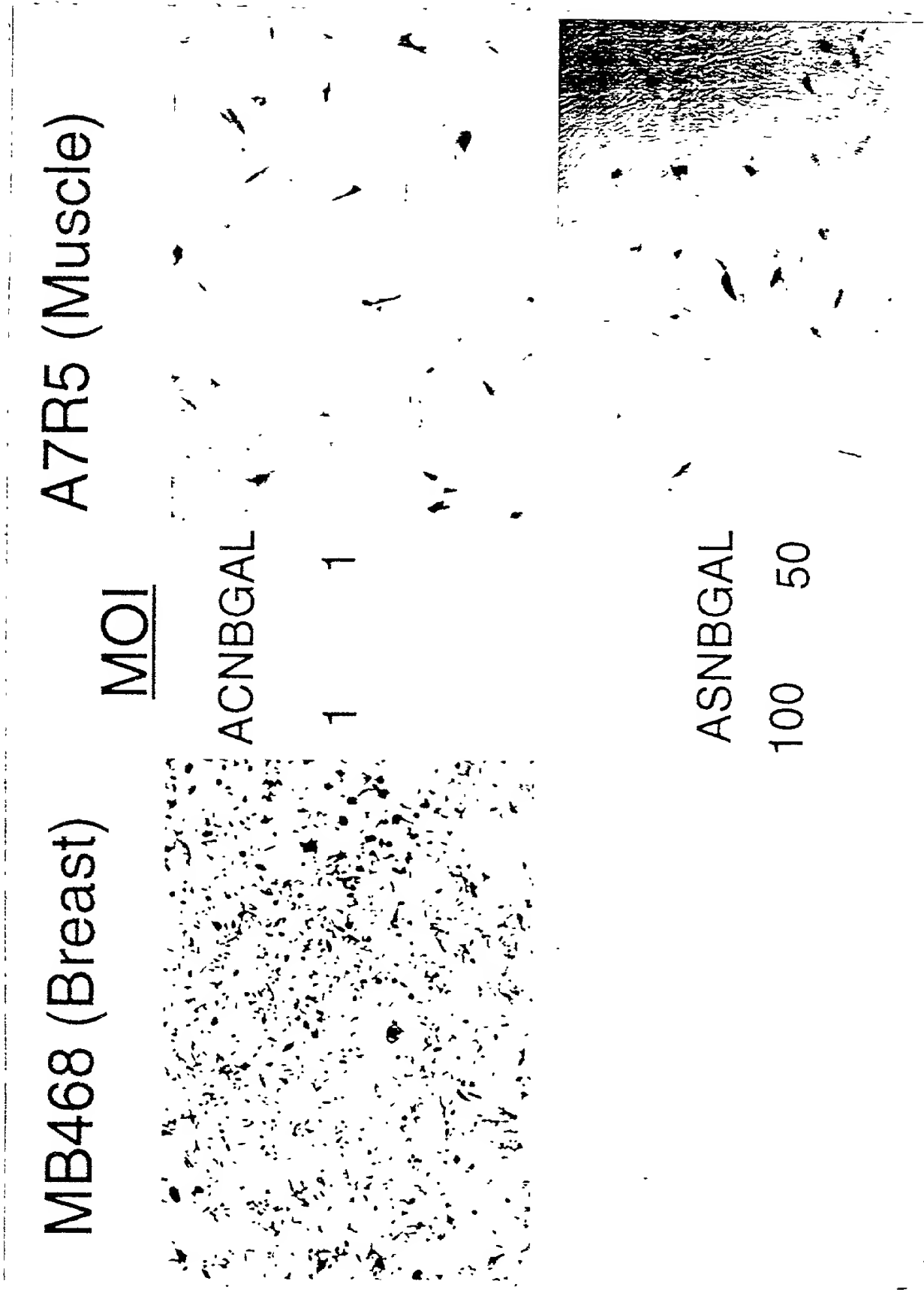


FIG. 19

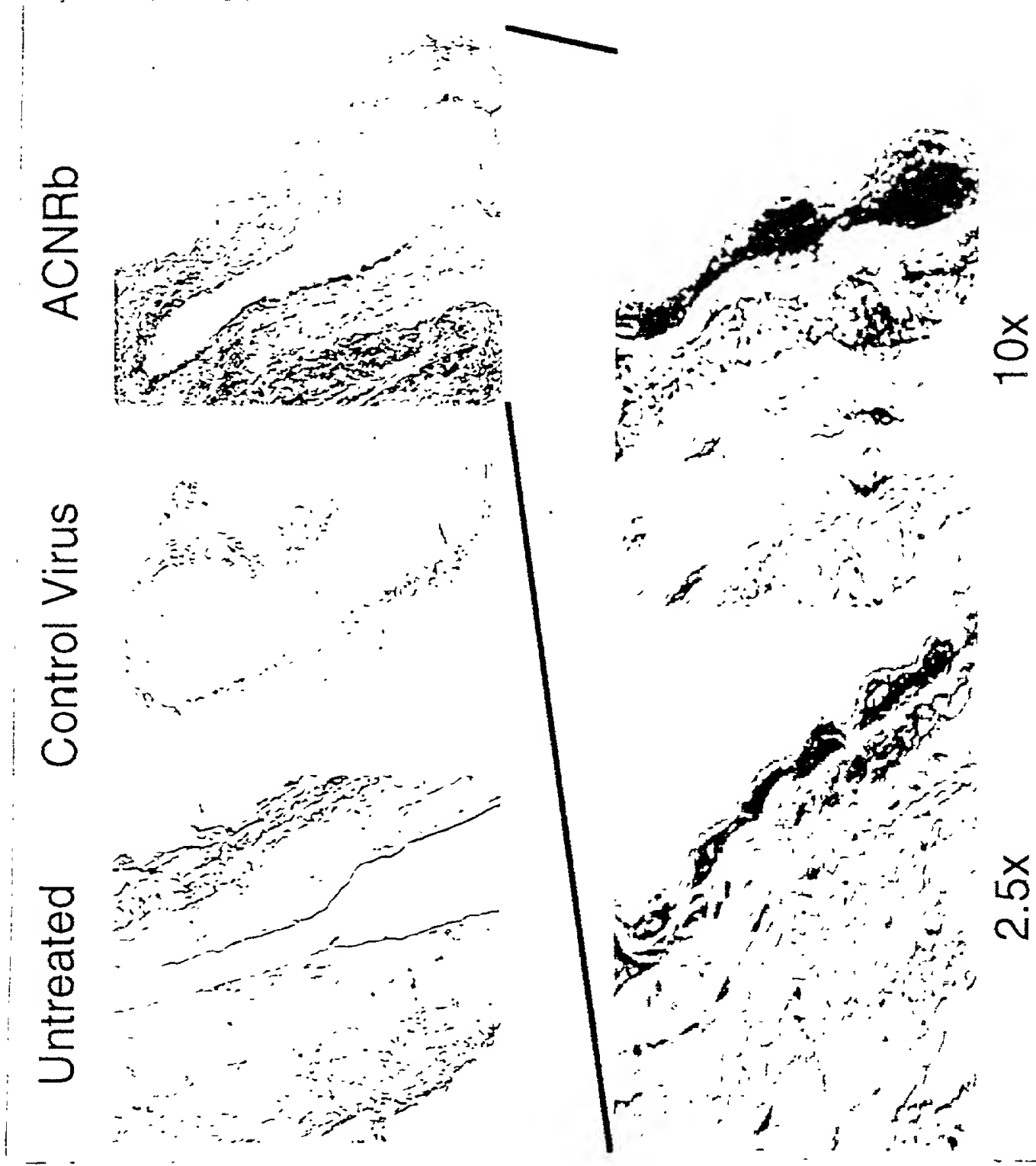


FIG. 20

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A7r5 3H-THYMIDINE

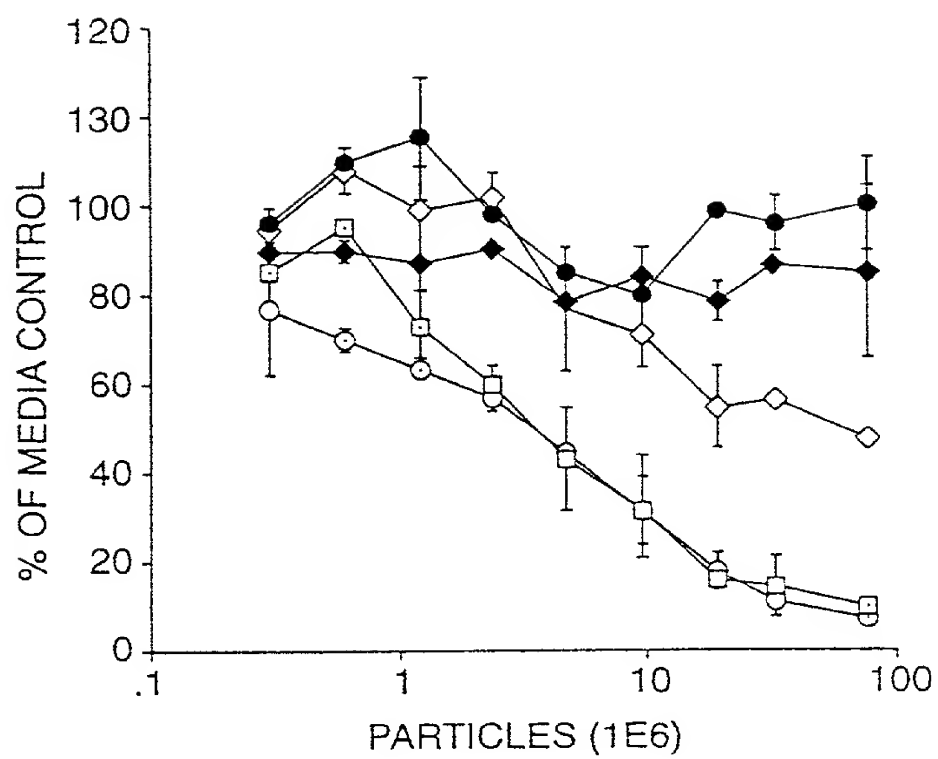


FIG. 21A

MDA468 3H-THYMIDINE

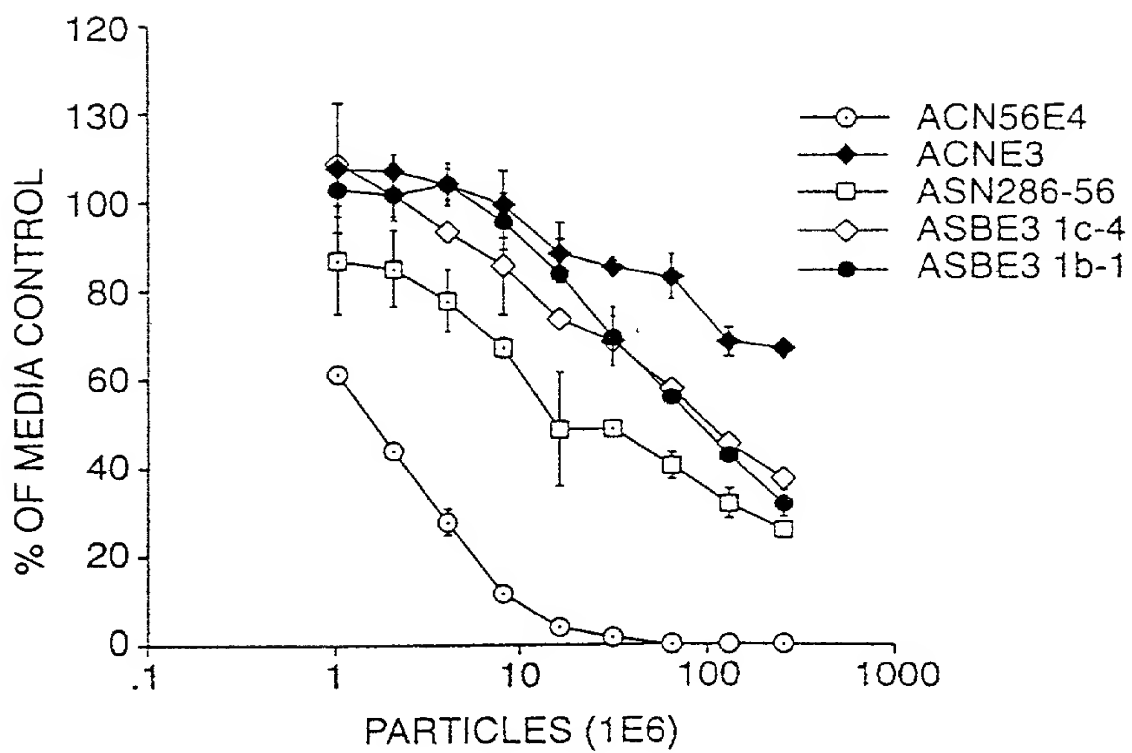


FIG. 21B

DECLARATION AND POWER OF ATTORNEY

As a below named inventor, I declare that:

My residence, post office address and citizenship are as stated below next to my name; I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural inventors are named below) of the subject matter which is claimed and for which a patent is sought on the invention entitled: **TISSUE SPECIFIC EXPRESSION OF RETINOBLASTOMA PROTEIN** the specification of which ___ is attached hereto or X was filed on February 14, 1997 as Application No. 08/801,092 and was amended on ___ (if applicable).

I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above. I acknowledge the duty to disclose information which is material to the examination of this application in accordance with Title 37, Code of Federal Regulations, Section 1.56. I claim foreign priority benefits under Title 35, United States Code, Section 119 of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed.

Prior Foreign Application(s)

Country	Application No.	Date of Filing	Priority Claimed Under 35 USC 119
			Yes _ No _
			Yes _ No _

I hereby claim the benefit under Title 35, United States Code §119(e) of any United States provisional application(s) listed below:

Application No.	Filing Date

I claim the benefit under Title 35, United States Code, Section 120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of Title 35, United States Code, Section 112, I acknowledge the duty to disclose material information as defined in Title 37, Code of Federal Regulations, Section 1.56 which occurred between the filing date of the prior application and the national or PCT international filing date of this application:

Application No.	Date of Filing	Status
08/751,517	11/15/96	_ Patented <u>X</u> Pending _ Abandoned
		_ Patented _ Pending _ Abandoned

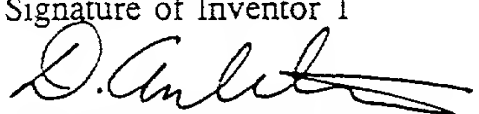
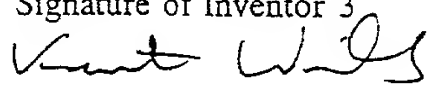
POWER OF ATTORNEY: As a named inventor, I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith.

William M. Smith, Reg. No. 30,223
James M. Heslin, Reg. No. 29,541
Renee A. Fitts, Reg. No. 35,136
Randolph T. Apple, Reg. No. 36,429
Joe Liebeschuetz, Reg. No. 37,505
Karen B. Dow, Reg. No. 29,684

Send Correspondence to: William M. Smith TOWNSEND and TOWNSEND and CREW LLP Two Embarcadero Center, 8th Floor San Francisco, CA 94111-3834	Direct Telephone Calls to: (Name, Reg. No., Telephone No.) Name: William M. Smith Reg. No.: 30,223 Telephone: (415) 326-2400
--	---

Full Name of Inventor 1	Last Name ANTELMAN	First Name DOUGLAS	Middle Name or Initial	
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Full Name of Inventor 2	Last Name GREGORY	First Name RICHARD	Middle Name or Initial J.	
Residence & Citizenship	City WESTFORD	State/Foreign Country MASSACHUSETTS	Country of Citizenship U.S.A.	
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Full Name of Inventor 3	Last Name WILLS	First Name KENNETH	Middle Name or Initial N.	
Residence & Citizenship	City ENCINITAS	State/Foreign Country CALIFORNIA	Country of Citizenship U.S.A.	
Post Office Address	Post Office Address 821 BLUFFCREST LANE	City ENCINITAS	State/Country CA	Zip Code 92024

I further declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

Signature of Inventor 1  DOUGLAS ANTELMAN	Signature of Inventor 2 RICHARD J. GREGORY	Signature of Inventor 3  KENNETH N. WILLS
Date April 25 1997	Date	Date April 25, 1997

DP.MRG 8/96

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I further declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

Signature of Inventor 1	Signature of Inventor 2	Signature of Inventor 3
DOUGLAS ANTELMAN	<i>Richard J. Gregory</i> RICHARD J. GREGORY	KENNETH N. WILLS
Date	Date 4/23/91	Date

DP.MRG 8/96
H:\HOME\RAF\WORK\16930\1020\1020.DEC

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: Antelman, Douglas
Gregory, Richard J.
Wils, Kenneth N.
- (ii) TITLE OF INVENTION: Tissue Specific Expression of
Retinoblastoma Protein
- (iii) NUMBER OF SEQUENCES: 46
- (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: TOWNSEND and TOWNSEND and CREW LLP
 - (B) STREET: Two Embarcadero Center, 8th Floor
 - (C) CITY: San Francisco
 - (D) STATE: CA
 - (E) COUNTRY: USA
 - (F) ZIP: 94111
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER: US 08/801,092
 - (B) FILING DATE: 14-FEB-1997
 - (C) CLASSIFICATION:
- (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: US 08/751,517
 - (B) FILING DATE: 15-NOV-1996
 - (C) CLASSIFICATION:
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Fitts, Renee A.
 - (B) REGISTRATION NUMBER: 35,136
 - (C) REFERENCE/DOCKET NUMBER: 016930-001020
- (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: 415-576-0200
 - (B) TELEFAX: 703-576-0300

(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 437 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: not relevant
 - (D) TOPOLOGY: not relevant
- (ii) MOLECULE TYPE: peptide
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

Met	Ala	Leu	Ala	Gly	Ala	Pro	Ala	Gly	Gly	Pro	Cys	Ala	Pro	Ala	Leu
1				5				10						15	
Glu	Ala	Leu	Leu	Gly	Ala	Gly	Ala	Leu	Arg	Leu	Leu	Asp	Ser	Ser	Gln
		20						25					30		
Ile	Val	Ile	Ile	Ser	Ala	Ala	Gln	Asp	Ala	Ser	Ala	Pro	Pro	Ala	Pro
		35					40					45			
Thr	Gly	Pro	Ala	Ala	Pro	Ala	Ala	Gly	Pro	Cys	Asp	Pro	Asp	Leu	Leu
	50					55					60				
Leu	Phe	Ala	Thr	Pro	Gln	Ala	Pro	Arg	Pro	Thr	Pro	Ser	Ala	Pro	Arg
65					70					75					80
Pro	Ala	Leu	Gly	Arg	Pro	Pro	Val	Lys	Arg	Arg	Leu	Asp	Leu	Glu	Thr
				85					90					95	
Asp	His	Gln	Tyr	Leu	Ala	Glu	Ser	Ser	Gly	Pro	Ala	Arg	Gly	Arg	Gly
			100					105					110		
Arg	His	Pro	Gly	Lys	Gly	Val	Lys	Ser	Pro	Gly	Glu	Lys	Ser	Arg	Tyr
		115					120					125			
Glu	Thr	Ser	Leu	Asn	Leu	Thr	Thr	Lys	Arg	Phe	Leu	Glu	Leu	Leu	Ser
	130					135					140				
His	Ser	Ala	Asp	Gly	Val	Val	Asp	Leu	Asn	Trp	Ala	Ala	Glu	Val	Leu
145					150					155					160
Lys	Val	Gln	Lys	Arg	Arg	Ile	Tyr	Asp	Ile	Thr	Asn	Val	Leu	Glu	Gly
				165					170					175	
Ile	Gln	Leu	Ile	Ala	Lys	Lys	Ser	Lys	Asn	His	Ile	Gln	Trp	Leu	Gly
			180					185					190		
Ser	His	Thr	Thr	Val	Gly	Val	Gly	Gly	Arg	Leu	Glu	Gly	Leu	Thr	Gln
		195					200					205			
Asp	Leu	Arg	Gln	Leu	Gln	Glu	Ser	Glu	Gln	Gln	Leu	Asp	His	Leu	Met
	210					215					220				
Asn	Ile	Cys	Thr	Thr	Gln	Leu	Arg	Leu	Leu	Ser	Glu	Asp	Thr	Asp	Ser
225					230					235					240
Gln	Arg	Leu	Ala	Tyr	Val	Thr	Cys	Gln	Asp	Leu	Arg	Ser	Ile	Ala	Asp
				245					250					255	
Pro	Ala	Glu	Gln	Met	Val	Met	Val	Ile	Lys	Ala	Pro	Pro	Glu	Thr	Gln
			260					265					270		
Leu	Gln	Ala	Val	Asp	Ser	Ser	Glu	Asn	Phe	Gln	Ile	Ser	Leu	Lys	Ser
		275					280					285			
Lys	Gln	Gly	Pro	Ile	Asp	Val	Phe	Leu	Cys	Pro	Glu	Glu	Thr	Val	Gly
	290					295					300				
Gly	Ile	Ser	Pro	Gly	Lys	Thr	Pro	Ser	Gln	Glu	Val	Thr	Ser	Glu	Glu
305					310					315					320
Glu	Asn	Arg	Ala	Thr	Asp	Ser	Ala	Thr	Ile	Val	Ser	Pro	Pro	Pro	Ser
				325					330					335	

Ser Pro Pro Ser Ser Leu Thr Thr Asp Pro Ser Gln Ser Leu Leu Ser
 340 345 350
 Leu Glu Gln Glu Pro Leu Leu Ser Arg Met Gly Ser Leu Arg Ala Pro
 355 360 365
 Val Asp Glu Asp Arg Leu Ser Pro Leu Val Ala Ala Asp Ser Leu Leu
 370 375 380
 Glu His Val Arg Glu Asp Phe Ser Gly Leu Leu Pro Glu Glu Phe Ile
 385 390 395 400
 Ser Leu Ser Pro Pro His Glu Ala Leu Asp Tyr His Phe Gly Leu Glu
 405 410 415
 Glu Gly Glu Gly Ile Arg Asp Leu Phe Asp Cys Asp Phe Gly Asp Leu
 420 425 430
 Thr Pro Leu Asp Phe
 435

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2517 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

GGAATTCCGT GGCCGGGACT TTGCAGGCAG CGGCGGCCGG GGGCGGAGCG GGATCGAGCC	60
CTCGCCGAGG CCTGCCGCCA TGGGCCCGCG CCGCCGCCGC CGCCTGTCAC CCGGGCCGCG	120
CGGGCCGTGA GCGTCATGGC CTTGGCCGGG GCCCCTGCGG GCGGCCCATG CGCGCCGGCG	180
CTGGAGGCCC TGCTCGGGGC CGGCGCGCTG CGGCTGCTCG ACTCCTCGCA GATCGTCATC	240
ATCTCCGCCG CGCAGGACGC CAGCGCCCCG CCGGCTCCCA CCGGCCCCGC GGCGCCCGCC	300
GCCGGCCCCCT GCGACCCTGA CCTGCTGCTC TTCGCCACAC CGCAGGCGCC CCGGCCACAA	360
CCCAGTGCGC CGCGGCCCGC GCTCGGCCGC CCGCCGGTGA AGCGGAGGCT GGACCTGGAA	420
ACTGACCATC AGTACCTGGC CGAGAGCAGT GGGCCAGCTC GGGGCAGAGG CCGCCATCCA	480
GGAAAAGGTG TGAAATCCCC GGGGGAGAAG TCACGCTATG AGACCTCACT GAATCTGACC	540
ACCAAGCGCT TCCTGGAGCT GCTGAGCCAC TCGGCTGACG GTGTCGTCGA CCTGAACTGG	600
GCTGCCGAGG TGCTGAAGGT GCAGAAGCGG CGCATCTATG ACATCACCAA CGTCCTTGAG	660
GGCATCCAGC TCATTGCCAA GAAGTCCAAG AACCACATCC AGTGGCTGGG CAGCCACACC	720
ACAGTGGGCG TCGGCGGACG GCTTGAGGGG TTGACCCAGG ACCTCCGACA GCTGCAGGAG	780
AGCGAGCAGC AGCTGGACCA CCTGATGAAT ATCTGTACTA CGCAGCTGCG CCTGCTCTCC	840

GAGGACACTG	ACAGCCAGCG	CCTGGCCTAC	GTGACGTGTC	AGGACCTTCG	TAGCATTGCA	900
GACCCTGCAG	AGCAGATGGT	TATGGTGATC	AAAGCCCCTC	CTGAGACCCA	GCTCCAAGCC	960
GTGGACTCTT	CGGAGAACTT	TCAGATCTCC	CTTAAGAGCA	AACAAGGCCC	GATCGATGTT	1020
TTCCTGTGCC	CTGAGGAGAC	CGTAGGTGGG	ATCAGCCCTG	GGAAGACCCC	ATCCCAGGAG	1080
GTCACTTCTG	AGGAGGAGAA	CAGGGCCACT	GACTCTGCCA	CCATAGTGTC	ACCACCACCA	1140
TCATCTCCCC	CCTCATCCCT	CACCACAGAT	CCCAGCCAGT	CTCTACTCAG	CCTGGAGCAA	1200
GAACCGCTGT	TGTCCCGGAT	GGGCAGCCTG	CGGGCTCCCG	TGGACGAGGA	CCGCCTGTCC	1260
CCGCTGGTGG	CGGCCGACTC	GCTCCTGGAG	CATGTGCGGG	AGGACTTCTC	CGGCCTCCTC	1320
CCTGAGGAGT	TCATCAGCCT	TTCCCCACCC	CACGAGGCCC	TCGACTACCA	CTTCGGCCTC	1380
GAGGAGGGCG	AGGGCATCAG	AGACCTCTTC	GACTGTGACT	TTGGGGACCT	CACCCCCCTG	1440
GATTTCTGAC	AGGGCTTGGA	GGGACCAGGG	TTTCCAGAGT	AGCTCACCTT	GTCTCTGCAG	1500
CCCTGGAGCC	CCCTGTCCCT	GGCCGTCCTC	CCAGCCTGTT	TGGAAACATT	TAATTTATAC	1560
CCCTCTCCTC	TGTCTCCAGA	AGCTTCTAGC	TCTGGGGTCT	GGCTACCGCT	AGGAGGCTGA	1620
GCAAGCCAGG	AAGGGAAGGA	GTCTGTGTGG	TGTGTATGTG	CATGCAGCCT	ACACCCACAC	1680
GTGTGTACCG	GGGGTGAATG	TGTGTGAGCA	TGTGTGTGTG	CATGTACCGG	GGAATGAAGG	1740
TGAACATACA	CCTCTGTGTG	TGCACTGCAG	ACACGCCCCA	GTGTGTCCAC	ATGTGTGTGC	1800
ATGAGTCCAT	CTCTGCGCGT	GGGGGGGCTC	TAACTGCACT	TTCGGCCCTT	TTGCTCGTGG	1860
GGTCCCACAA	GGCCCAGGGC	AGTGCCTGCT	CCCAGAATCT	GGTGCTCTGA	CCAGGCCAGG	1920
TGGGGAGGCT	TTGGCTGGCT	GGGCGTGTAG	GACGGTGAGA	GCACTTCTGT	CTTAAAGGTT	1980
TTTTCTGATT	GAAGCTTTAA	TGGAGCGTTA	TTTATTTATC	GAGGCCTCTT	TGGTGAGCCT	2040
GGGGAATCAG	CAAAAGGGGA	GGAGGGGTGT	GGGGTTGATA	CCCCAACTCC	CTCTACCCTT	2100
GAGCAAGGGC	AGGGGTCCCT	GAGCTGTTCT	TCTGCCCCAT	ACTGAAGGAA	CTGAGGCCTG	2160
GGTGATTTAT	TTATTGGGAA	AGTGAGGGAG	GGAGACAGAC	TGACTGACAG	CCATGGGTGG	2220
TCAGATGGTG	GGGTGGGCCC	TCTCCAGGGG	GCCAGTTCAG	GGCCCAGCTG	CCCCCCAGGA	2280
TGGATATGAG	ATGGGAGAGG	TGAGTGGGGG	ACCTTCACTG	ATGTGGGCAG	GAGGGGTGGT	2340
GAAGGCCTCC	CCCAGCCCAG	ACCCTGTGGT	CCCTCCTGCA	GTGTCTGAAG	CGCCTGCCTC	2400
CCCACTGCTC	TGCCCCACCC	TCCAATCTGC	ACTTTGATTT	GCTTCCTAAC	AGCTCTGTTC	2460
CCTCCTGCTT	TGGTTTTAAT	AAATATTTTG	ATGACGTTAA	AAAAAGGAAT	TCGATAT	2517

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2994 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

TTCCGGTTTT	TCTCAGGGGA	CGTTGAAATT	ATTTTTGTAA	CGGGAGTCGG	GAGAGGACGG	60
GGCGTGCCCC	GCGTGCGCGC	GCGTCGTCCT	CCCCGGCGCT	CCTCCACAGC	TCGCTGGCTC	120
CCGCCGCGGA	AAGGCGTCAT	GCCGCCCAAA	ACCCCCGAA	AAACGGCCGC	CACCGCCGCC	180
GCTGCCGCCG	CGGAACCCCC	GGCACCGCCG	CCGCCGCCCC	CTCCTGAGGA	GGACCCAGAG	240
CAGGACAGCG	GCCCGGAGGA	CCTGCCTCTC	GTCAGGCTTG	AGTTTGAAGA	AACAGAAGAA	300
CCTGATTTTA	CTGCATTATG	TCAGAAATTA	AAGATAACCAG	ATCATGTCAG	AGAGAGAGCT	360
TGGTTAACTT	GGGAGAAAGT	TTCATCTGTG	GATGGAGTAT	TGGGAGGTTA	TATTCAAAG	420
AAAAAGGAAC	TGTGGGGAAT	CTGTATCTTT	ATTGCAGCAG	TTGACCTAGA	TGAGATGTCG	480
TTCACTTTTA	CTGAGCTACA	GAAAAACATA	GAAATCAGTG	TCCATAAATT	CTTTAACTTA	540
CTAAAAGAAA	TTGATAACCAG	TACCAAAGTT	GATAATGCTA	TGTCAAGACT	GTTGAAGAAG	600
TATGATGTAT	TGTTTGCCT	CTTCAGCAAA	TTGGAAAGGA	CATGTGAACT	TATATATTTG	660
ACACAACCCA	GCAGTTCGAT	ATCTACTGAA	ATAAATTCTG	CATTGGTGCT	AAAAGTTTCT	720
TGGATCACAT	TTTTATTAGC	TAAAGGGGAA	GTATTACAAA	TGGAAGATGA	TCTGGTGATT	780
TCATTTTCAGT	TAATGCTATG	TGTCCTTGAC	TATTTTATTA	AACTCTCACC	TCCCATGTTG	840
CTCAAAGAAC	CATATAAAAC	AGCTGTTATA	CCCATTAATG	GTTACCTCG	AACACCCAGG	900
CGAGGTCAGA	ACAGGAGTGC	ACGGATAGCA	AAACAAC TAG	AAAATGATAC	AAGAATTATT	960
GAAGTTCTCT	GTAAAGAACA	TGAATGTAAT	ATAGATGAGG	TGAAAAATGT	TTATTTCAA	1020
AATTTTATAC	CTTTTATGAA	TTCTCTTGGA	CTTGTAACAT	CTAATGGACT	TCCAGAGGTT	1080
GAAAATCTTT	CTAAACGATA	CGAAGAAATT	TATCTTAAAA	ATAAAGATCT	AGATGCAAGA	1140
TTATTTTGG	ATCATGATAA	AACTCTTCAG	ACTGATTCTA	TAGACAGTTT	TGAAACACAG	1200
AGAACACCAC	GAAAAAGTAA	CCTTGATGAA	GAGGTGAATG	TAATTCCTCC	ACACACTCCA	1260
GTTAGGACTG	TTATGAACAC	TATCCAACAA	TTAATGATGA	TTTTAAATTC	AGCAAGTGAT	1320
CAACCTTCAG	AAAATCTGAT	TTCCTATTTT	AACAAC TGCA	CAGTGAATCC	AAAAGAAAGT	1380
ATACTGAAAA	GAGTGAAGGA	TATAGGATAC	ATCTTTAAAG	AGAAATTTGC	TAAAGCTGTG	1440
GGACAGGGTT	GTGTCGAAAT	TGGATCACAG	CGATACAAAC	TTGGAGTTTC	CTTGTATTAC	1500
CGAGTAATGG	AATCCATGCT	TAAATCAGAA	GAAGAACGAT	TATCCATTCA	AAATTTTAGC	1560
AAACTTCTGA	ATGACAACAT	TTTTCATATG	TCTTTATTGG	CGTGCGCTCT	TGAGGTTGTA	1620

ATGGCCACAT	ATAGCAGAAG	TACATCTCAG	AATCTTGATT	CTGGAACAGA	TTTGTCTTTC	1680
CCATGGATTC	TGAATGTGCT	TAATTTAAAA	GCCTTTGATT	TTTACAAAGT	GATCGAAAGT	1740
TTTATCAAAG	CAGAAGGCAA	CTTGACAAGA	GAAATGATAA	AACATTTAGA	ACGATGTGAA	1800
CATCGAATCA	TGGAATCCCT	TGCATGGCTC	TCAGATTCAC	CTTTATTTGA	TCTTATTAAA	1860
CAATCAAAGG	ACCGAGAAGG	ACCAACTGAT	CACCTTGAAT	CTGCTTGTCC	TCTTAATCTT	1920
CCTCTCCAGA	ATAATCACAC	TGCAGCAGAT	ATGTATCTTT	CTCCTGTAAG	ATCTCCAAAG	1980
AAAAAAGGTT	CAACTACGCG	TGTAAATTCT	ACTGCAAATG	CAGAGACACA	AGCAACCTCA	2040
GCCTTCCAGA	CCCAGAAGCC	ATTGAAATCT	ACCTCTCTTT	CACTGTTTTA	TAAAAAAGTG	2100
TATCGGCTAG	CCTATCTCCG	GCTAAATACA	CTTTGTGAAC	GCCTTCTGTC	TGAGCACCCA	2160
GAATTAGAAC	ATATCATCTG	GACCCTTTTC	CAGCACACCC	TGCAGAATGA	GTATGAACTC	2220
ATGAGAGACA	GGCATTTGGA	CCAAATTATG	ATGTGTTCCA	TGTATGGCAT	ATGCAAAGTG	2280
AAGAATATAG	ACCTTAAATT	CAAAATCATT	GTAACAGCAT	ACAAGGATCT	TCCTCATGCT	2340
G TTCAGGAGA	CATTCAAACG	TGTTTTGATC	AAAGAAGAGG	AGTATGATTC	TATTATAGTA	2400
TTCTATAACT	CGGTCTTCAT	GCAGAGACTG	AAAACAAATA	TTTTGCAGTA	TGCTTCCACC	2460
AGGCCCCCTA	CCTTGTCACC	AATACCTCAC	ATTCCTCGAA	GCCCTTACAA	GTTTCCTAGT	2520
TCACCCTTAC	GGATTCTTGG	AGGGAACATC	TATATTTTAC	CCCTGAAGAG	TCCATATAAA	2580
ATTTCAGAAG	GTCTGCCAAC	ACCAACAAAA	ATGACTCCAA	GATCAAGAAT	CTTAGTATCA	2640
ATTGGTGAAT	CATTCGGGAC	TTCTGAGAAG	TTCCAGAAAA	TAAATCAGAT	GGTATGTAAC	2700
AGCGACCGTG	TGCTCAAAAG	AAGTGCTGAA	GGAAGCAACC	CTCCTAAACC	ACTGAAAAAA	2760
CTACGCTTTG	ATATTGAAGG	ATCAGATGAA	GCAGATGGAA	GTAAACATCT	CCCAGGAGAG	2820
TCCAAATTTT	AGCAGAAACT	GGCAGAAATG	ACTTCTACTC	GAACACGAAT	GCAAAAGCAG	2880
AAAATGAATG	ATAGCATGGA	TACCTCAAAC	AAGGAAGAGA	AATGAGGATC	TCAGGACCTT	2940
GGTGGACACT	GTGTACACCT	CTGGATTTCAT	TGTCTCTCAC	AGATGTGACT	GTAT	2994

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 928 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: not relevant
- (D) TOPOLOGY: not relevant

(ii) MOLECULE TYPE: peptide

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Pro Pro Lys Thr Pro Arg Lys Thr Ala Ala Thr Ala Ala Ala Ala

1	5	10	15
Ala	Ala	Glu	Pro
20	Pro	Ala	Pro
Pro	Pro	Pro	Pro
25	Pro	Pro	Pro
30	Glu	Glu	Asp
Pro	Glu	Gln	Asp
35	Ser	Gly	Pro
40	Glu	Asp	Leu
45	Pro	Leu	Val
50	Arg	Leu	Glu
55	Phe	Thr	Ala
60	Leu	Cys	Gln
65	Lys	Leu	Lys
70	Val	Arg	Glu
75	Glu	Arg	Ala
80	Trp	Leu	Thr
85	Thr	Trp	Glu
90	Lys	Lys	Lys
95	Val	Ser	Ser
100	Val	Asp	Gly
105	Val	Leu	Gly
110	Leu	Glu	Asp
115	Thr	Phe	Thr
120	Glu	Leu	Gln
125	Lys	Asn	Ile
130	Thr	Ser	Thr
135	Lys	Val	Lys
140	Asp	Thr	Ser
145	Tyr	Asp	Val
150	Leu	Phe	Ala
155	Leu	Lys	Lys
160	Leu	Lys	Lys
165	Leu	Arg	Thr
170	Cys	Glu	Leu
175	Ile	Tyr	Leu
180	Val	Leu	Gln
185	Leu	Lys	Met
190	Val	Leu	Gln
195	Val	Leu	Gln
200	Val	Leu	Gln
205	Val	Leu	Gln
210	Val	Leu	Gln
215	Val	Leu	Gln
220	Val	Leu	Gln
225	Val	Leu	Gln
230	Val	Leu	Gln
235	Val	Leu	Gln
240	Val	Leu	Gln
245	Val	Leu	Gln
250	Val	Leu	Gln
255	Val	Leu	Gln
260	Val	Leu	Gln
265	Val	Leu	Gln
270	Val	Leu	Gln
275	Val	Leu	Gln
280	Val	Leu	Gln
285	Val	Leu	Gln
290	Val	Leu	Gln
295	Val	Leu	Gln
300	Val	Leu	Gln
305	Val	Leu	Gln
310	Val	Leu	Gln
315	Val	Leu	Gln
320	Val	Leu	Gln
325	Val	Leu	Gln
330	Val	Leu	Gln
335	Val	Leu	Gln

Leu	Asp	His	Asp	Lys	Thr	Leu	Gln	Thr	Asp	Ser	Ile	Asp	Ser	Phe	Glu
			340					345					350		
Thr	Gln	Arg	Thr	Pro	Arg	Lys	Ser	Asn	Leu	Asp	Glu	Glu	Val	Asn	Val
		355					360					365			
Ile	Pro	Pro	His	Thr	Pro	Val	Arg	Thr	Val	Met	Asn	Thr	Ile	Gln	Gln
	370					375					380				
Leu	Met	Met	Ile	Leu	Asn	Ser	Ala	Ser	Asp	Gln	Pro	Ser	Glu	Asn	Leu
385					390					395					400
Ile	Ser	Tyr	Phe	Asn	Asn	Cys	Thr	Val	Asn	Pro	Lys	Glu	Ser	Ile	Leu
				405					410					415	
Lys	Arg	Val	Lys	Asp	Ile	Gly	Tyr	Ile	Phe	Lys	Glu	Lys	Phe	Ala	Lys
			420					425					430		
Ala	Val	Gly	Gln	Gly	Cys	Val	Glu	Ile	Gly	Ser	Gln	Arg	Tyr	Lys	Leu
		435					440					445			
Gly	Val	Arg	Leu	Tyr	Tyr	Arg	Val	Met	Glu	Ser	Met	Leu	Lys	Ser	Glu
	450					455					460				
Glu	Glu	Arg	Leu	Ser	Ile	Gln	Asn	Phe	Ser	Lys	Leu	Leu	Asn	Asp	Asn
465					470					475					480
Ile	Phe	His	Met	Ser	Leu	Leu	Ala	Cys	Ala	Leu	Glu	Val	Val	Met	Ala
			485						490					495	
Thr	Tyr	Ser	Arg	Ser	Thr	Ser	Gln	Asn	Leu	Asp	Ser	Gly	Thr	Asp	Leu
			500					505					510		
Ser	Phe	Pro	Trp	Ile	Leu	Asn	Val	Leu	Asn	Leu	Lys	Ala	Phe	Asp	Phe
		515					520					525			
Tyr	Lys	Val	Ile	Glu	Ser	Phe	Ile	Lys	Ala	Glu	Gly	Asn	Leu	Thr	Arg
	530					535					540				
Glu	Met	Ile	Lys	His	Leu	Glu	Arg	Cys	Glu	His	Arg	Ile	Met	Glu	Ser
545					550					555					560
Leu	Ala	Trp	Leu	Ser	Asp	Ser	Pro	Leu	Phe	Asp	Leu	Ile	Lys	Gln	Ser
				565					570					575	
Lys	Asp	Arg	Glu	Gly	Pro	Thr	Asp	His	Leu	Glu	Ser	Ala	Cys	Pro	Leu
			580					585					590		
Asn	Leu	Pro	Leu	Gln	Asn	Asn	His	Thr	Ala	Ala	Asp	Met	Tyr	Leu	Ser
		595					600					605			
Pro	Val	Arg	Ser	Pro	Lys	Lys	Lys	Gly	Ser	Thr	Thr	Arg	Val	Asn	Ser
	610					615						620			
Thr	Ala	Asn	Ala	Glu	Thr	Gln	Ala	Thr	Ser	Ala	Phe	Gln	Thr	Gln	Lys
625					630					635					640
Pro	Leu	Lys	Ser	Thr	Ser	Leu	Ser	Leu	Phe	Tyr	Lys	Lys	Val	Tyr	Arg
				645					650					655	
Leu	Ala	Tyr	Leu	Arg	Leu	Asn	Thr	Leu	Cys	Glu	Arg	Leu	Leu	Ser	Glu
			660					665					670		

His Pro Glu Leu Glu His Ile Ile Trp Thr Leu Phe Gln His Thr Leu
 675 680 685
 Gln Asn Glu Tyr Glu Leu Met Arg Asp Arg His Leu Asp Gln Ile Met
 690 695 700
 Met Cys Ser Met Tyr Gly Ile Cys Lys Val Lys Asn Ile Asp Leu Lys
 705 710 715 720
 Phe Lys Ile Ile Val Thr Ala Tyr Lys Asp Leu Pro His Ala Val Gln
 725 730 735
 Glu Thr Phe Lys Arg Val Leu Ile Lys Glu Glu Glu Tyr Asp Ser Ile
 740 745 750
 Ile Val Phe Tyr Asn Ser Val Phe Met Gln Arg Leu Lys Thr Asn Ile
 755 760 765
 Leu Gln Tyr Ala Ser Thr Arg Pro Pro Thr Leu Ser Pro Ile Pro His
 770 775 780
 Ile Pro Arg Ser Pro Tyr Lys Phe Pro Ser Ser Pro Leu Arg Ile Pro
 785 790 795 800
 Gly Gly Asn Ile Tyr Ile Ser Pro Leu Lys Ser Pro Tyr Lys Ile Ser
 805 810 815
 Glu Gly Leu Pro Thr Pro Thr Lys Met Thr Pro Arg Ser Arg Ile Leu
 820 825 830
 Val Ser Ile Gly Glu Ser Phe Gly Thr Ser Glu Lys Phe Gln Lys Ile
 835 840 845
 Asn Gln Met Val Cys Asn Ser Asp Arg Val Leu Lys Arg Ser Ala Glu
 850 855 860
 Gly Ser Asn Pro Pro Lys Pro Leu Lys Lys Leu Arg Phe Asp Ile Glu
 865 870 875 880
 Gly Ser Asp Glu Ala Asp Gly Ser Lys His Leu Pro Gly Glu Ser Lys
 885 890 895
 Phe Gln Gln Lys Leu Ala Glu Met Thr Ser Thr Arg Thr Arg Met Gln
 900 905 910
 Lys Gln Lys Met Asn Asp Ser Met Asp Thr Ser Asn Lys Glu Glu Lys
 915 920 925

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 3853 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 209..250

- (ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 254..289
- (ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 293..505
- (ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 509..514
- (ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 518..520
- (ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 524..658
- (ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 662..691
- (ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 695..748
- (ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 752..781
- (ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 785..829
- (ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 1132..1134
- (ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 1138..1149
- (ix) FEATURE:
 (A) NAME/KEY: CDS
 (B) LOCATION: 833..862

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

GACGGATCGG GAGATCTCCC GATCCCCTAT GGTGACTCT CAGTACAATC TGCTCTGATG	60
CCGCATAGTT AAGCCAGTAT CTGCTCCCTG CTTGTGTGTT GGAGGTCGCT GAGTAGTGCG	120
CGAGCAAAAT TTAAGCTACA ACAAGGCAAG GCTTGACCGA CAATTGCATG AAGAATCTGC	180
TTAGGGTTAG GCGTTTTGCG CTGCTTCG CGA TGT ACG GGC CAG ATA TAC GCG	232
Arg Cys Thr Gly Gln Ile Tyr Ala	
1 5	
TTG ACA TTG ATT ATT GAC TAG TTA TTA ATA GTA ATC AAT TAC GGG GTC	280

Leu	Thr	Leu	Ile	Ile	Asp		Leu	Leu	Ile	Val	Ile	Asn	Tyr	Gly	Val	
	10						1				5					
ATT	AGT	TCA	TAG	CCC	ATA	TAT	GGA	GTT	CCG	CGT	TAC	ATA	ACT	TAC	GGT	328
Ile	Ser	Ser		Pro	Ile	Tyr	Gly	Val	Pro	Arg	Tyr	Ile	Thr	Tyr	Gly	
10				1				5					10			
AAA	TGG	CCC	GCC	TGG	CTG	ACC	GCC	CAA	CGA	CCC	CCG	CCC	ATT	GAC	GTC	376
Lys	Trp	Pro	Ala	Trp	Leu	Thr	Ala	Gln	Arg	Pro	Pro	Pro	Ile	Asp	Val	
		15					20					25				
AAT	AAT	GAC	GTA	TGT	TCC	CAT	AGT	AAC	GCC	AAT	AGG	GAC	TTT	CCA	TTG	424
Asn	Asn	Asp	Val	Cys	Ser	His	Ser	Asn	Ala	Asn	Arg	Asp	Phe	Pro	Leu	
	30					35					40					
ACG	TCA	ATG	GGT	GGA	CTA	TTT	ACG	GTA	AAC	TGC	CCA	CTT	GGC	AGT	ACA	472
Thr	Ser	Met	Gly	Gly	Leu	Phe	Thr	Val	Asn	Cys	Pro	Leu	Gly	Ser	Thr	
45					50					55					60	
TCA	AGT	GTA	TCA	TAT	GCC	AAG	TAC	GCC	CCC	TAT	TGA	CGT	CAA			514
Ser	Ser	Val	Ser	Tyr	Ala	Lys	Tyr	Ala	Pro	Tyr		Arg	Gln			
				65					70			1				
TGA	CGG	TAA	ATG	GCC	CGC	CTG	GCA	TTA	TGC	CCA	GTA	CAT	GAC	CTT	ATG	562
	Arg		Met	Ala	Arg	Leu	Ala	Leu	Cys	Pro	Val	His	Asp	Leu	Met	
	1		1				5					10				
GGA	CTT	TCC	TAC	TTG	GCA	GTA	CAT	CTA	CGT	ATT	AGT	CAT	CGC	TAT	TAC	610
Gly	Leu	Ser	Tyr	Leu	Ala	Val	His	Leu	Arg	Ile	Ser	His	Arg	Tyr	Tyr	
15						20					25					
CAT	GGT	GAT	GCG	GTT	TTG	GCA	GTA	CAT	CAA	TGG	GCG	TGG	ATA	GCG	GTT	658
His	Gly	Asp	Ala	Val	Leu	Ala	Val	His	Gln	Trp	Ala	Trp	Ile	Ala	Val	
30					35					40					45	
TGA	CTC	ACG	GGG	ATT	TCC	AAG	TCT	CCA	CCC	CAT	TGA	CGT	CAA	TGG	GAG	706
	Leu	Thr	Gly	Ile	Ser	Lys	Ser	Pro	Pro	His		Arg	Gln	Trp	Glu	
	1				5					10		1				
TTT	GTT	TTG	GCA	CCA	AAA	TCA	ACG	GGA	CTT	TCC	AAA	ATG	TCG			748
Phe	Val	Leu	Ala	Pro	Lys	Ser	Thr	Gly	Leu	Ser	Lys	Met	Ser			
5					10					15						
TAA	CAA	CTC	CGC	CCC	ATT	GAC	GCA	AAT	GGG	CGG	TAG	CGC	TGT	ACG	GTG	796
	Gln	Leu	Arg	Pro	Ile	Asp	Ala	Asn	Gly	Arg		Arg	Cys	Thr	Val	
	1				5					10		1				
GGA	GGT	CTA	TAT	AAG	CAG	AGC	TCT	CTG	GCT	AAC	TAG	AGA	ACC	CAC	TGC	844
Gly	Gly	Leu	Tyr	Lys	Gln	Ser	Ser	Leu	Ala	Asn		Arg	Thr	His	Cys	
5					10					15		1				
TTA	CTG	GCT	TAT	CGA	AAT	TAATACGACT	CACTATAGGG	AGACCCAAGC								892
Leu	Leu	Ala	Tyr	Arg	Asn											
5					10											
TTGCGCGCGG	TACCACTCTC	TTCCGCATCG	CTGTCTGCGA	GGGCCAGCTG	TTGGGCTCGC											952
GGTTGAGGAC	AAACTCTTCG	CGGTCTTTCC	AGTACTCTTG	GATCGGAAAC	CCGTCGGCCT											1012
CCGAACGGTA	CTCCGCCACC	GAGGGACCTG	AGCGAGTCCG	CATCGACCGG	ATCGGAAAAC											1072
CTCTCGAGGC	GGCCGCTGCA	GTCTAGACGA	ATTCGCGTAC	GATATCGATG	GGCCCTATT											1131

CTA TAG TGT CAC CTA AAT GCTAGAGCTC GCTGATCAGC CTCGACTGTG	1179
Leu Cys His Leu Asn	
1 1	
CCTTCTAGTT GCCAGCCATC TGTTGTTTGC CCCTCCCCCG TGCCTTCCTT GACCCTGGAA	1239
GGTGCCACTC CCACTGTCCT TTCCTAATAA AATGAGGAAA TTGCATCGCA TTGTCTGAGT	1299
AGGTGTCATT CTATTCTGGG GGGTGGGGTG GGGCAGGACA GCAAGGGGGA GGATTGGGAA	1359
GACAATAGCC GAAATGACCG ACCAAGCGAC GCCCAACCTG CCATCACGAG ATTTTCGATTC	1419
CACCGCCGCC TTCTATGAAA GGTTGGGCTT CGGAATCGTT TTCCGGGACG CCGGCTGGAT	1479
GATCCTCCAG CGCGGGGATC TCATGCTGGA GTTCTTCGCC CACCCCAACT TGTTTATTGC	1539
AGCTTATAAT GGTTACAAAT AAAGCAATAG CATCACAAAT TTCACAAATA AAGCATTTTT	1599
TTCACTGCAT TCTAGTTGTG GTTTGTCCAA ACTCATCAAT GTATCTTATC ATGTCTGTAT	1659
ACCGTCGACC TCTAGCTAGA GCTTGGCGTA ATCATGGTCA TAGCTGTTTC CTGTGTGAAA	1719
TTGTTATCCG CTCACAATTC CACACAACAT ACGAGCCGGA AGCATAAAGT GTAAAGCCTG	1779
GGGTGCCTAA TGAGTGAGCT AACTCACATT AATTGCGTTG CGCTCACTGC CCGCTTTCCA	1839
GTCGGGAAAC CTGTCGTGCC AGCTGCATTA ATGAATCGGC CAACGCGCGG GGAGAGGCGG	1899
TTTGCGTATT GGGCGCTCTT CCGCTTCCTC GCTCACTGAC TCGCTGCGCT CGGTGCTTCG	1959
GCTGCGGCGA GCGGTATCAG CTCACTCAAA GGCGGTAATA CGGTTATCCA CAGAATCAGG	2019
GGATAACGCA GGAAAGAACA TGTGAGCAAA AGGCCAGCAA AAGGCCAGGA ACCGTAAAAA	2079
GGCCGCGTTG CTGGCGTTTT TCCATAGGCT CCGCCCCCCT GACGAGCATC AAAAAAATCG	2139
ACGCTCAAGT CAGAGGTGGC GAAACCCGAC AGGACTATAA AGATACCAGG CGTTTCCCCC	2199
TGGAAGCTCC CTCGTGCGCT CTCCTGTTCC GACCCTGCCG CTTACCGGAT ACCTGTCCGC	2259
CTTTCTCCCT TCGGGAAGCG TGGCGCTTTC TCAATGCTCA CGCTGTAGGT ATCTCAGTTC	2319
GGTGTAGGTC GTTCGCTCCA AGCTGGGCTG TGTGCACGAA CCCCCCGTTC AGCCCGACCG	2379
CTGCGCCTTA TCCGGTAACT ATCGTCTTGA GTCCAACCCG GTAAGACACG ACTTATCGCC	2439
ACTGGCAGCA GCCACTGGTA ACAGGATTAG CAGAGCGAGG TATGTAGGCG GTGCTACAGA	2499
GTTCTTGAAG TGGTGGCCTA ACTACGGCTA CACTAGAAGG ACAGTATTTG GTATCTGCGC	2559
TCTGCTGAAG CCAGTTACCT TCGGAAAAAG AGTTGGTAGC TCTTGATCCG GCAAACAAAC	2619
CACCGCTGGT AGCGGTGGTT TTTTGTGTTG CAAGCAGCAG ATTACGCGCA GAAAAAAGG	2679
ATCTCAAGAA GATCCTTTGA TCTTTTCTAC GGGGTCTGAC GCTCAGTGGA ACGAAAACTC	2739
ACGTTAAGGG ATTTTGGTCA TGAGATTATC AAAAAGGATC TTCACCTAGA TCCTTTTAAA	2799
TTAAAAATGA AGTTTTAAAT CAATCTAAAG TATATATGAG TAAACTTGGT CTGACAGTTA	2859
CCAATGCTTA ATCAGTGAGG CACCTATCTC AGCGATCTGT CTATTTTCGT CATCCATAGT	2919

TGCCTGACTC	CCCGTCGTGT	AGATAACTAC	GATACGGGAG	GGCTTACCAT	CTGGCCCCAG	2979
TGCTGCAATG	ATACCGCGAG	ACCCACGCTC	ACCGGCTCCA	GATTTATCAG	CAATAAACCA	3039
GCCAGCCGGA	AGGGCCGAGC	GCAGAAGTGG	TCCTGCAACT	TTATCCGCCT	CCATCCAGTC	3099
TATTAATTGT	TGCCGGGAAG	CTAGAGTAAG	TAGTTCGCCA	GTTAATAGTT	TGCGCAACGT	3159
TGTTGCCATT	GCTACAGGCA	TCGTGGTGTC	ACGCTCGTCG	TTTGGTATGG	CTTCATTCAG	3219
CTCCGGTTCC	CAACGATCAA	GGCGAGTTAC	ATGATCCCCC	ATGTTGTGCA	AAAAAGCGGT	3279
TAGCTCCTTC	GGTCCTCCGA	TCGTTGTCAG	AAGTAAGTTG	GCCGCAGTGT	TATCACTCAT	3339
GGTTATGGCA	GCACTGCATA	ATTCTCTTAC	TGTCATGCCA	TCCGTAAGAT	GCTTTTCTGT	3399
GACTGGTGAG	TACTCAACCA	AGTCATTCTG	AGAATAGTGT	ATGCGGCGAC	CGAGTTGCTC	3459
TTGCCCCGGC	TCAATACGGG	ATAATACCGC	GCCACATAGC	AGAACTTTAA	AAGTGCTCAT	3519
CATTGGAAAA	CGTTCTTCGG	GGCGAAAACT	CTCAAGGATC	TTACCGCTGT	TGAGATCCAG	3579
TTCGATGTAA	CCCACTCGTG	CACCCAACCTG	ATCTTCAGCA	TCTTTTACTT	TCACCAGCGT	3639
TTCTGGGTGA	GCAAAAACAG	GAAGGCAAAA	TGCCGCAAAA	AAGGGAATAA	GGGCGACACG	3699
GAAATGTTGA	ATACTCATAC	TCTTCCTTTT	TCAATATTAT	TGAAGCATTT	ATCAGGGTTA	3759
TTGTCTCATG	AGCGGATACA	TATTTGAATG	TATTTAGAAA	AATAAACAAA	TAGGGGTTCC	3819
GCGCACATTT	CCCCGAAAAG	TGCCACCTGA	CGTC			3853

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 14 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Arg	Cys	Thr	Gly	Gln	Ile	Tyr	Ala	Leu	Thr	Leu	Ile	Ile	Asp
1				5					10				

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 12 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

Leu	Leu	Ile	Val	Ile	Asn	Tyr	Gly	Val	Ile	Ser	Ser
1				5					10		

(2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 71 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

```

Pro Ile Tyr Gly Val Pro Arg Tyr Ile Thr Tyr Gly Lys Trp Pro Ala
 1           5           10           15
Trp Leu Thr Ala Gln Arg Pro Pro Pro Ile Asp Val Asn Asn Asp Val
          20           25           30
Cys Ser His Ser Asn Ala Asn Arg Asp Phe Pro Leu Thr Ser Met Gly
          35           40           45
Gly Leu Phe Thr Val Asn Cys Pro Leu Gly Ser Thr Ser Ser Val Ser
          50           55           60
Tyr Ala Lys Tyr Ala Pro Tyr
 65           70

```

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 2 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

```

Arg Gln
 1

```

(2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 1 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

```

Arg
 1

```

(2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 45 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

```

Met Ala Arg Leu Ala Leu Cys Pro Val His Asp Leu Met Gly Leu Ser
 1           5           10           15
Tyr Leu Ala Val His Leu Arg Ile Ser His Arg Tyr Tyr His Gly Asp
          20           25           30
Ala Val Leu Ala Val His Gln Trp Ala Trp Ile Ala Val
      35           40           45

```

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 10 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

```

Leu Thr Gly Ile Ser Lys Ser Pro Pro His
 1           5           10

```

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 18 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

```

Arg Gln Trp Glu Phe Val Leu Ala Pro Lys Ser Thr Gly Leu Ser Lys
 1           5           10           15

```

Met Ser

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 10 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

```

Gln Leu Arg Pro Ile Asp Ala Asn Gly Arg
 1           5           10

```


(2) INFORMATION FOR SEQ ID NO:15:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

Arg	Cys	Thr	Val	Gly	Gly	Leu	Tyr	Lys	Gln	Ser	Ser	Leu	Ala	Asn
1				5					10					15

(2) INFORMATION FOR SEQ ID NO:16:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 10 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

Arg	Thr	His	Cys	Leu	Leu	Ala	Tyr	Arg	Asn
1				5					10

(2) INFORMATION FOR SEQ ID NO:17:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

Leu
1

(2) INFORMATION FOR SEQ ID NO:18:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 4 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

Cys	His	Leu	Asn
1			

(2) INFORMATION FOR SEQ ID NO:19:

- (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 4026 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 209..250

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 254..289

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 293..505

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 509..514

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 518..520

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 524..658

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 662..691

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 695..748

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 752..781

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 785..829

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 833..862

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1305..1307

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1311..1322

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

GACGGATCGG	GAGATCTCCC	GATCCCCTAT	GGTCGACTCT	CAGTACAATC	TGCTCTGATG	60										
CCGCATAGTT	AAGCCAGTAT	CTGCTCCCTG	CTTGTGTGTT	GGAGGTCGCT	GAGTAGTGCG	120										
CGAGCAAAAT	TTAAGCTACA	ACAAGGCAAG	GCTTGACCGA	CAATTGCATG	AAGAATCTGC	180										
TTAGGGTTAG	GCGTTTTGCG	CTGCTTCG	CGA	TGT	ACG	GGC	CAG	ATA	TAC	GCG	232					
			Arg	Cys	Thr	Gly	Gln	Ile	Tyr	Ala						
			1				5									
TTG	ACA	TTG	ATT	ATT	GAC	TAG	TTA	TTA	ATA	GTA	ATC	AAT	TAC	GGG	GTC	280
Leu	Thr	Leu	Ile	Ile	Asp		Leu	Leu	Ile	Val	Ile	Asn	Tyr	Gly	Val	
	10						1				5					
ATT	AGT	TCA	TAG	CCC	ATA	TAT	GGA	GTT	CCG	CGT	TAC	ATA	ACT	TAC	GGT	328
Ile	Ser	Ser		Pro	Ile	Tyr	Gly	Val	Pro	Arg	Tyr	Ile	Thr	Tyr	Gly	
	10			1				5					10			
AAA	TGG	CCC	GCC	TGG	CTG	ACC	GCC	CAA	CGA	CCC	CCG	CCC	ATT	GAC	GTC	376
Lys	Trp	Pro	Ala	Trp	Leu	Thr	Ala	Gln	Arg	Pro	Pro	Pro	Ile	Asp	Val	
		15					20					25				
AAT	AAT	GAC	GTA	TGT	TCC	CAT	AGT	AAC	GCC	AAT	AGG	GAC	TTT	CCA	TTG	424
Asn	Asn	Asp	Val	Cys	Ser	His	Ser	Asn	Ala	Asn	Arg	Asp	Phe	Pro	Leu	
	30					35					40					
ACG	TCA	ATG	GGT	GGA	CTA	TTT	ACG	GTA	AAC	TGC	CCA	CTT	GGC	AGT	ACA	472
Thr	Ser	Met	Gly	Gly	Leu	Phe	Thr	Val	Asn	Cys	Pro	Leu	Gly	Ser	Thr	
	45				50					55					60	
TCA	AGT	GTA	TCA	TAT	GCC	AAG	TAC	GCC	CCC	TAT	TGA	CGT	CAA			514
Ser	Ser	Val	Ser	Tyr	Ala	Lys	Tyr	Ala	Pro	Tyr		Arg	Gln			
				65				70				1				
TGA	CGG	TAA	ATG	GCC	CGC	CTG	GCA	TTA	TGC	CCA	GTA	CAT	GAC	CTT	ATG	562
	Arg		Met	Ala	Arg	Leu	Ala	Leu	Cys	Pro	Val	His	Asp	Leu	Met	
	1		1			5						10				
GGA	CTT	TCC	TAC	TTG	GCA	GTA	CAT	CTA	CGT	ATT	AGT	CAT	CGC	TAT	TAC	610
Gly	Leu	Ser	Tyr	Leu	Ala	Val	His	Leu	Arg	Ile	Ser	His	Arg	Tyr	Tyr	
	15				20						25					
CAT	GGT	GAT	GCG	GTT	TTG	GCA	GTA	CAT	CAA	TGG	GCG	TGG	ATA	GCG	GTT	658
His	Gly	Asp	Ala	Val	Leu	Ala	Val	His	Gln	Trp	Ala	Trp	Ile	Ala	Val	
	30				35					40					45	
TGA	CTC	ACG	GGG	ATT	TCC	AAG	TCT	CCA	CCC	CAT	TGA	CGT	CAA	TGG	GAG	706
	Leu	Thr	Gly	Ile	Ser	Lys	Ser	Pro	Pro	His		Arg	Gln	Trp	Glu	
	1				5					10		1				
TTT	GTT	TTG	GCA	CCA	AAA	TCA	ACG	GGA	CTT	TCC	AAA	ATG	TCG			748
Phe	Val	Leu	Ala	Pro	Lys	Ser	Thr	Gly	Leu	Ser	Lys	Met	Ser			
	5				10					15						
TAA	CAA	CTC	CGC	CCC	ATT	GAC	GCA	AAT	GGG	CGG	TAG	GCG	TGT	ACG	GTG	796
	Gln	Leu	Arg	Pro	Ile	Asp	Ala	Asn	Gly	Arg		Ala	Cys	Thr	Val	
	1				5					10		1				
GGA	GGT	CTA	TAT	AAG	CAG	AGC	TCT	CTG	GCT	AAC	TAG	AGA	ACC	CAC	TGC	844
Gly	Gly	Leu	Tyr	Lys	Gln	Ser	Ser	Leu	Ala	Asn		Arg	Thr	His	Cys	
	5				10					15		1				

TTA CTG GCT TAT CGA AAT TAATACGACT CACTATAGGG AGACCCAAGC	892
Leu Leu Ala Tyr Arg Asn	
5 10	
TTCGCGCGGG TACCACTCTC TTCCGCATCG CTGTCTGCGA GGGCCAGCTG TTGGGCTCGC	952
GGTTGAGGAC AAACCTCTTCG CGGTCTTTCC AGTACTCTTG GATCGGAAAC CCGTCGGCCT	1012
CCGAACGGTA CTCCGCCACC GAGGGACCTG AGCGAGTCCG CATCGACCGG ATCGGAAAAC	1072
CTCTCGAGGA ACTGAAAAAC CAGAAAGTTA ACTGGTAAGT TTAGTCTTTT TGTCTTTTAA	1132
TTTCAGGTCC CGGATCCGGT GGTGGTGCAA ATCAAAGAAC TGCTCCTCAG TGGATGTTGC	1192
CTTTACTTCT AGGCCTGTAC GGAAGTGTTA CTTCTGCTCT AAAAGCTGCG GAATTGTACC	1252
CGCGGCCGCT GCAGTCTAGA CGAATTCGCG TACGATATCG ATGGGCCCTA TT CTA	1307
Leu	
1	
TAG TGT CAC CTA AAT GCTAGAGCTC GCTGATCAGC CTCGACTGTG CCTTCTAGTT	1362
Cys His Leu Asn	
1	
GCCAGCCATC TGTTGTTTGC CCCTCCCCCG TGCCTTCCTT GACCCTGGAA GGTGCCACTC	1422
CCACTGTCCT TTCCTAATAA AATGAGGAAA TTGCATCGCA TTGTCTGAGT AGGTGTCATT	1482
CTATTCTGGG GGGTGGGGTG GGGCAGGACA GCAAGGGGGA GGATTGGGAA GACAATAGCC	1542
GAAATGACCG ACCAAGCGAC GCCCAACCTG CCATCACGAG ATTTTCGATTC CACCGCCGCC	1602
TTCTATGAAA GGTGGGGCTT CGGAATCGTT TTCCGGGACG CCGGCTGGAT GATCCTCCAG	1662
CGCGGGGATC TCATGCTGGA GTTCTTCGCC CACCCCAACT TGTTTATTGC AGCTTATAAT	1722
GGTTACAAAT AAAGCAATAG CATCACAAAT TTCACAAATA AAGCATTTTTT TTCACTGCAT	1782
TCTAGTTGTG GTTTGTCCAA ACTCATCAAT GTATCTTATC ATGTCTGTAT ACCGTCGACC	1842
TCTAGCTAGA GCTTGGCGTA ATCATGGTCA TAGCTGTTTC CTGTGTGAAA TTGTTATCCG	1902
CTCACAATTC CACACAACAT ACGAGCCGGA AGCATAAAGT GTAAAGCCTG GGGTGCCTAA	1962
TGAGTGAGCT AACTCACATT AATTGCGTTG CGCTCACTGC CCGCTTTCCA GTCGGGAAAC	2022
CTGTCGTGCC AGCTGCATTA ATGAATCGGC CAACGCGCGG GGAGAGGCGG TTTGCGTATT	2082
GGGCGCTCTT CCGCTTCCTC GCTCACTGAC TCGCTGCGCT CGGTCGTTCG GCTGCGGCGA	2142
GCGGTATCAG CTCACTCAAA GGCGGTAATA CGGTTATCCA CAGAATCAGG GGATAACGCA	2202
GGAAAGAACA TGTGAGCAAA AGGCCAGCAA AAGGCCAGGA ACCGTAAAAA GGCCGCGTTG	2262
CTGGCGTTTT TCCATAGGCT CCGCCCCCCT GACGAGCATC ACAAAAATCG ACGCTCAAGT	2322
CAGAGGTGGC GAAACCCGAC AGGACTATAA AGATACCAGG CGTTTCCCCC TGGAAGCTCC	2382
CTCGTGCGCT CTCCTGTTCC GACCCTGCCG CTTACCGGAT ACCTGTCCGC CTTTCTCCCT	2442
TCGGGAAGCG TGGCGCTTTC TCAATGCTCA CGCTGTAGGT ATCTGAGTTC GGTGTAGGTC	2502

GTTGCTCCA	AGCTGGGCTG	TGTGCACGAA	CCCCCGTTC	AGCCCGACCG	CTGCGCCTTA	2562
TCCGGTAACT	ATCGTCTTGA	GTCCAACCCG	GTAAGACACG	ACTTATCGCC	ACTGGCAGCA	2622
GCCACTGGTA	ACAGGATTAG	CAGAGCGAGG	TATGTAGGCG	GTGCTACAGA	GTTCTTGAAG	2682
TGGTGGCCTA	ACTACGGCTA	CACTAGAAGG	ACAGTATTTG	GTATCTGCGC	TCTGCTGAAG	2742
CCAGTTACCT	TCGGAAAAAG	AGTTGGTAGC	TCTTGATCCG	GCAAACAAAC	CACCGCTGGT	2802
AGCGGTGGTT	TTTTTGTTTG	CAAGCAGCAG	ATTACGCGCA	GAAAAAAAGG	ATCTCAAGAA	2862
GATCCTTTGA	TCTTTTCTAC	GGGGTCTGAC	GCTCAGTGGA	ACGAAAATC	ACGTTAAGGG	2922
ATTTTGGTCA	TGAGATTATC	AAAAAGGATC	TTCACCTAGA	TCCTTTTAAA	TTAAAAATGA	2982
AGTTTTAAAT	CAATCTAAAG	TATATATGAG	TAAACTTGGT	CTGACAGTTA	CCAATGCTTA	3042
ATCAGTGAGG	CACCTATCTC	AGCGATCTGT	CTATTTTCGT	CATCCATAGT	TGCCTGACTC	3102
CCCGTCGTGT	AGATAACTAC	GATACGGGAG	GGCTTACCAT	CTGGCCCCAG	TGCTGCAATG	3162
ATACCGCGAG	ACCCACGCTC	ACCGGCTCCA	GATTTATCAG	CAATAAACCA	GCCAGCCGGA	3222
AGGGCCGAGC	GCAGAAGTGG	TCCTGCAACT	TTATCCGCCT	CCATCCAGTC	TATTAATTGT	3282
TGCCGGGAAG	CTAGAGTAAG	TAGTTCGCCA	GTTAATAGTT	TGCGCAACGT	TGTTGCCATT	3342
GCTACAGGCA	TCGTGGTGTC	ACGCTCGTCG	TTTGGTATGG	CTTCATTGAG	CTCCGGTTCC	3402
CAACGATCAA	GGCGAGTTAC	ATGATCCCCC	ATGTTGTGCA	AAAAAGCGGT	TAGCTCCTTC	3462
GGTCCTCCGA	TCGTTGTCAG	AAGTAAGTTG	GCCGCAGTGT	TATCACTCAT	GGTTATGGCA	3522
GCACTGCATA	ATTCTCTTAC	TGTCATGCCA	TCCGTAAGAT	GCTTTTCTGT	GACTGGTGAG	3582
TACTCAACCA	AGTCATTCTG	AGAATAGTGT	ATGCGGCGAC	CGAGTTGCTC	TTGCCCCGGC	3642
TCAATACGGG	ATAATACCGC	GCCACATAGC	AGAACTTTAA	AAGTGCTCAT	CATTGGAAAA	3702
CGTTCTTCGG	GGCGAAAACT	CTCAAGGATC	TTACCGCTGT	TGAGATCCAG	TTGATGTAA	3762
CCCACTCGTG	CACCCAACCTG	ATCTTCAGCA	TCTTTTACTT	TCACCAGCGT	TTCTGGGTGA	3822
GCAAAAACAG	GAAGGCAAAA	TGCCGCAAAA	AAGGGAATAA	GGGCGACACG	GAAATGTTGA	3882
ATACTCATAC	TCTTCCTTTT	TCAATATTAT	TGAAGCATTT	ATCAGGGTTA	TTGTCTCATG	3942
AGCGGATACA	TATTTGAATG	TATTTAGAAA	AATAAACAAA	TAGGGGTTCC	GCGCACATTT	4002
CCCCGAAAAG	TGCCACCTGA	CGTC				4026

(2) INFORMATION FOR SEQ ID NO:20:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 14 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

Arg Cys Thr Gly Gln Ile Tyr Ala Leu Thr Leu Ile Ile Asp
 1 5 10

(2) INFORMATION FOR SEQ ID NO:21:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 12 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

Leu Leu Ile Val Ile Asn Tyr Gly Val Ile Ser Ser
 1 5 10

(2) INFORMATION FOR SEQ ID NO:22:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 71 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

Pro Ile Tyr Gly Val Pro Arg Tyr Ile Thr Tyr Gly Lys Trp Pro Ala
 1 5 10 15
 Trp Leu Thr Ala Gln Arg Pro Pro Pro Ile Asp Val Asn Asn Asp Val
 20 25 30
 Cys Ser His Ser Asn Ala Asn Arg Asp Phe Pro Leu Thr Ser Met Gly
 35 40 45
 Gly Leu Phe Thr Val Asn Cys Pro Leu Gly Ser Thr Ser Ser Val Ser
 50 55 60
 Tyr Ala Lys Tyr Ala Pro Tyr
 65 70

(2) INFORMATION FOR SEQ ID NO:23:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

Arg Gln
 1

(2) INFORMATION FOR SEQ ID NO:24:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

Arg
1

(2) INFORMATION FOR SEQ ID NO:25:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 45 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

Met	Ala	Arg	Leu	Ala	Leu	Cys	Pro	Val	His	Asp	Leu	Met	Gly	Leu	Ser
1				5					10					15	
Tyr	Leu	Ala	Val	His	Leu	Arg	Ile	Ser	His	Arg	Tyr	Tyr	His	Gly	Asp
			20					25					30		
Ala	Val	Leu	Ala	Val	His	Gln	Trp	Ala	Trp	Ile	Ala	Val			
			35				40					45			

(2) INFORMATION FOR SEQ ID NO:26:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 10 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

Leu	Thr	Gly	Ile	Ser	Lys	Ser	Pro	Pro	His
1				5					10

(2) INFORMATION FOR SEQ ID NO:27:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 18 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

Arg Gln Trp Glu Phe Val Leu Ala Pro Lys Ser Thr Gly Leu Ser Lys
 1 5 10 15

Met Ser

(2) INFORMATION FOR SEQ ID NO:28:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 10 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

Gln Leu Arg Pro Ile Asp Ala Asn Gly Arg
 1 5 10

(2) INFORMATION FOR SEQ ID NO:29:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

Ala Cys Thr Val Gly Gly Leu Tyr Lys Gln Ser Ser Leu Ala Asn
 1 5 10 15

(2) INFORMATION FOR SEQ ID NO:30:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 10 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

Arg Thr His Cys Leu Leu Ala Tyr Arg Asn
 1 5 10

(2) INFORMATION FOR SEQ ID NO:31:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

Leu
1

(2) INFORMATION FOR SEQ ID NO:32:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 4 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

Cys His Leu Asn
1

(2) INFORMATION FOR SEQ ID NO:33:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 4249 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 209..250

- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 254..289

- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 293..505

- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 509..514

- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 518..520

- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 524..658

- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 662..691

- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 695..748

- (ix) FEATURE:

(A) NAME/KEY: CDS
(B) LOCATION: 752..781

(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION: 785..829

(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION: 833..862

(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION: 1528..1530

(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION: 1534..1545

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

GACGGATCGG GAGATCTCCC GATCCCCTAT GGTGCGACTCT CAGTACAATC TGCTCTGATG	60
CCGCATAGTT AAGCCAGTAT CTGCTCCCTG CTTGTGTGTT GGAGGTCGCT GAGTAGTGCG	120
CGAGCAAAAT TTAAGCTACA ACAAGGCAAG GCTTGACCGA CAATTGCATG AAGAATCTGC	180
TTAGGGTTAG GCGTTTTGCG CTGCTTCG CGA TGT ACG GGC CAG ATA TAC GCG	232
Arg Cys Thr Gly Gln Ile Tyr Ala	
1 5	
TTG ACA TTG ATT ATT GAC TAG TTA TTA ATA GTA ATC AAT TAC GGG GTC	280
Leu Thr Leu Ile Ile Asp Leu Leu Ile Val Ile Asn Tyr Gly Val	
10 1 5	
ATT AGT TCA TAG CCC ATA TAT GGA GTT CCG CGT TAC ATA ACT TAC GGT	328
Ile Ser Ser Pro Ile Tyr Gly Val Pro Arg Tyr Ile Thr Tyr Gly	
10 1 5 10	
AAA TGG CCC GCC TGG CTG ACC GCC CAA CGA CCC CCG CCC ATT GAC GTC	376
Lys Trp Pro Ala Trp Leu Thr Ala Gln Arg Pro Pro Pro Ile Asp Val	
15 20 25	
AAT AAT GAC GTA TGT TCC CAT AGT AAC GCC AAT AGG GAC TTT CCA TTG	424
Asn Asn Asp Val Cys Ser His Ser Asn Ala Asn Arg Asp Phe Pro Leu	
30 35 40	
ACG TCA ATG GGT GGA CTA TTT ACG GTA AAC TGC CCA CTT GGC AGT ACA	472
Thr Ser Met Gly Gly Leu Phe Thr Val Asn Cys Pro Leu Gly Ser Thr	
45 50 55 60	
TCA AGT GTA TCA TAT GCC AAG TAC GCC CCC TAT TGA CGT CAA	514
Ser Ser Val Ser Tyr Ala Lys Tyr Ala Pro Tyr Arg Gln	
65 70 1	
TGA CGG TAA ATG GCC CGC CTG GCA TTA TGC CCA GTA CAT GAC CTT ATG	562
Arg Met Ala Arg Leu Ala Leu Cys Pro Val His Asp Leu Met	
1 1 5 10	
GGA CTT TCC TAC TTG GCA GTA CAT CTA CGT ATT AGT CAT CGC TAT TAC	610
Gly Leu Ser Tyr Leu Ala Val His Leu Arg Ile Ser His Arg Tyr Tyr	

15	20	25	
CAT GGT GAT GCG GTT TTG GCA GTA CAT CAA TGG GCG TGG ATA GCG GTT His Gly Asp Ala Val Leu Ala Val His Gln Trp Ala Trp Ile Ala Val 30 35 40 45			658
TGA CTC ACG GGG ATT TCC AAG TCT CCA CCC CAT TGA CGT CAA TGG GAG Leu Thr Gly Ile Ser Lys Ser Pro Pro His Arg Gln Trp Glu 1 5 10 1			706
TTT GTT TTG GCA CCA AAA TCA ACG GGA CTT TCC AAA ATG TCG Phe Val Leu Ala Pro Lys Ser Thr Gly Leu Ser Lys Met Ser 5 10 15			748
TAA CAA CTC CGC CCC ATT GAC GCA AAT GGG CGG TAG GCG TGT ACG GTG Gln Leu Arg Pro Ile Asp Ala Asn Gly Arg Ala Cys Thr Val 1 5 10 1			796
GGA GGT CTA TAT AAG CAG AGC TCT CTG GCT AAC TAG AGA ACC CAC TGC Gly Gly Leu Tyr Lys Gln Ser Ser Leu Ala Asn Arg Thr His Cys 5 10 15 1			844
TTA CTG GCT TAT CGA AAT TAATACGACT CACTATAGGG AGACCCAAGC Leu Leu Ala Tyr Arg Asn 5 10			892
TTTCGCGCGGG TACCACTCTC TTCCGCATCG CTGTCTGCGA GGGCCAGCTG TTGGGCTCGC			952
GGTTGAGGAC AAACCTCTTCG CGGTCTTTCC AGTACTCTTG GATCGGAAAC CCGTCGGCCT			1012
CCGAACGGTA CTCCGCCACC GAGGGACCTG AGCGAGTCCG CATCGACCGG ATCGGAAAAC			1072
CTCTCGAGGA ACTGAAAAAC CAGAAAGTTA ACTGGTAAGT TTAGTCTTTT TGTCTTTTTA			1132
TTTCAGGTCC CGGATCTGAG TTAGGGCGGG ACATGGGCGG AGTTAGGGGC GGGACTATGG			1192
TTGCTGACTA ATTGAGATGC ATGCTTTGCA TACTTCTGCC TGCTGGGGAG CCTGGGGACT			1252
TTCCACACCT GGTTGCTGAC TAATTGAGAT GCATGCTTTG CATACTTCTG CCTGCTGGGG			1312
AGCCTGGGGA CTTTCCACAC CCTAACTGAC ACACATTCCA CAGCTGGTTC TTTCAGATCC			1372
GGTGGTGGTG CAAATCAAAG AACTGCTCCT CAGTGGATGT TGCCTTTACT TCTAGGCCTG			1432
TACGGAAGTG TTAATTCTGC TCTAAAAGCT GCGGAATTGT ACCCGCGGCC GCTGCAGTCT			1492
AGACGAATTC GCGTACGATA TCGATGGGCC CTATT CTA TAG TGT CAC CTA AAT Leu Cys His Leu Asn 1 1			1545
GCTAGAGCTC GCTGATCAGC CTCGACTGTG CTTTCTAGTT GCCAGCCATC TGTTGTTTGC			1605
CCCTCCCCCG TGCCTTCCTT GACCCTGGAA GGTGCCACTC CCACTGTCCT TTCCTAATAA			1665
AATGAGGAAA TTGCATCGCA TTGTCTGAGT AGGTGTCATT CTATTCTGGG GGGTGGGGTG			1725
GGGCAGGACA GCAAGGGGGA GGATTGGGAA GACAATAGCC GAAATGACCG ACCAAGCGAC			1785
GCCCAACCTG CCATCACGAG ATTTGATTC CACCGCCGCC TTCTATGAAA GGTGTTGGCTT			1845
CGGAATCGTT TTCCGGGACG CCGGCTGGAT GATCCTCCAG CGCGGGGATC TCATGCTGGA			1905

GTTCTTCGCC	CACCCCAACT	TGTTTATTGC	AGCTTATAAT	GGTTACAAAT	AAAGCAATAG	1965
CATCACAAAT	TTCACAAATA	AAGCATTTTT	TTCACGTCAT	TCTAGTTGTG	GTTTGTCCAA	2025
ACTCATCAAT	GTATCTTATC	ATGTCTGTAT	ACCGTCGACC	TCTAGCTAGA	GCTTGGCGTA	2085
ATCATGGTCA	TAGCTGTTTC	CTGTGTGAAA	TTGTTATCCG	CTCACAAATC	CACACAACAT	2145
ACGAGCCGGA	AGCATAAAGT	GTAAAGCCTG	GGGTGCCTAA	TGAGTGAGCT	AACTCACATT	2205
AATTGCGTTG	CGCTCACTGC	CCGCTTTCCA	GTCGGGAAAC	CTGTCGTGCC	AGCTGCATTA	2265
ATGAATCGGC	CAACGCGCGG	GGAGAGGCGG	TTTGCGTATT	GGGCGCTCTT	CCGCTTCCTC	2325
GCTCACTGAC	TCGCTGCGCT	CGGTCGTTTC	GCTGCGGCGA	GCGGTATCAG	CTCACTCAAA	2385
GGCGGTAATA	CGGTTATCCA	CAGAATCAGG	GGATAACGCA	GGAAAGAACA	TGTGAGCAAA	2445
AGGCCAGCAA	AAGGCCAGGA	ACCGTAAAAA	GGCCGCGTTG	CTGGCGTTTT	TCCATAGGCT	2505
CCGCCCCCCT	GACGAGCATC	ACAAAAATCG	ACGCTCAAGT	CAGAGGTGGC	GAAACCCGAC	2565
AGGACTATAA	AGATACCAGG	CGTTTCCCCC	TGGAAGCTCC	CTCGTGCGCT	CTCCTGTTCC	2625
GACCCTGCCG	CTTACCGGAT	ACCTGTCCGC	CTTTCTCCCT	TCGGGAAGCG	TGGCGCTTTC	2685
TCAATGCTCA	CGCTGTAGGT	ATCTCAGTTC	GGTGTAAGTC	GTTCGCTCCA	AGCTGGGCTG	2745
TGTGCACGAA	CCCCCGTTC	AGCCCGACCG	CTGCGCCTTA	TCCGGTAACT	ATCGTCTTGA	2805
GTCCAACCCG	GTAAGACACG	ACTTATCGCC	ACTGGCAGCA	GCCACTGGTA	ACAGGATTAG	2865
CAGAGCGAGG	TATGTAGGCG	GTGCTACAGA	GTTCTTGAAG	TGGTGGCCTA	ACTACGGCTA	2925
CACTAGAAGG	ACAGTATTTG	GTATCTGCGC	TCTGCTGAAG	CCAGTTACCT	TCGGAAAAAG	2985
AGTTGGTAGC	TCTTGATCCG	GCAAACAAAC	CACCGCTGGT	AGCGGTGGTT	TTTTTGTTTG	3045
CAAGCAGCAG	ATTACGCGCA	GAAAAAAAGG	ATCTCAAGAA	GATCCTTTGA	TCTTTTCTAC	3105
GGGGTCTGAC	GCTCAGTGGA	ACGAAAACCTC	ACGTTAAGGG	ATTTTGGTCA	TGAGATTATC	3165
AAAAAGGATC	TTCACCTAGA	TCCTTTTAAA	TTAAAAATGA	AGTTTTAAAT	CAATCTAAAG	3225
TATATATGAG	TAAACTTGGT	CTGACAGTTA	CCAATGCTTA	ATCAGTGAGG	CACCTATCTC	3285
AGCGATCTGT	CTATTTTCGTT	CATCCATAGT	TGCCTGACTC	CCCGTCGTGT	AGATAACTAC	3345
GATACGGGAG	GGCTTACCAT	CTGGCCCCAG	TGCTGCAATG	ATACCGCGAG	ACCCACGCTC	3405
ACCGGCTCCA	GATTTATCAG	CAATAAACCA	GCCAGCCGGA	AGGGCCGAGC	GCAGAAGTGG	3465
TCCTGCAACT	TTATCCGCCT	CCATCCAGTC	TATTAATTGT	TGCCGGGAAG	CTAGAGTAAG	3525
TAGTTCGCCA	GTTAATAGTT	TGCGCAACGT	TGTTGCCATT	GCTACAGGCA	TCGTGGTGTC	3585
ACGCTCGTCG	TTTGGTATGG	CTTCATTCAG	CTCCGGTTCC	CAACGATCAA	GGCGAGTTAC	3645
ATGATCCCCC	ATGTTGTGCA	AAAAAGCGGT	TAGCTCCTTC	GGTCCTCCGA	TCGTTGTCAG	3705
AAGTAAGTTG	GCCGCAGTGT	TATCACTCAT	GGTTATGGCA	GCACTGCATA	ATTCTCTTAC	3765

TGTCATGCCA TCCGTAAGAT GCTTTTCTGT GACTGGTGAG TACTCAACCA AGTCATTCTG	3825
AGAATAGTGT ATGCGGCGAC CGAGTTGCTC TTGCCCCGGCG TCAATACGGG ATAATACCGC	3885
GCCACATAGC AGAACTTTAA AAGTGCTCAT CATTGGAAAA CGTTCTTCGG GCGGAAAAC	3945
CTCAAGGATC TTACCGCTGT TGAGATCCAG TTCGATGTAA CCCACTCGTG CACCCAACTG	4005
ATCTTCAGCA TCTTTTACTT TCACCAGCGT TTCTGGGTGA GCAAAAACAG GAAGGCAAAA	4065
TGCCGCAAAA AAGGGAATAA GGGCGACACG GAAATGTTGA ATACTCATA TCTTCCTTTT	4125
TCAATATTAT TGAAGCATTT ATCAGGGTTA TTGTCTCATG AGCGGATACA TATTTGAATG	4185
TATTTAGAAA AATAAACAAA TAGGGGTTCC GCGCACATTT CCCCAGAAAAG TGCCACCTGA	4245
CGTC	4249

(2) INFORMATION FOR SEQ ID NO:34:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 14 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

Arg	Cys	Thr	Gly	Gln	Ile	Tyr	Ala	Leu	Thr	Leu	Ile	Ile	Asp
1				5					10				

(2) INFORMATION FOR SEQ ID NO:35:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 12 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

Leu	Leu	Ile	Val	Ile	Asn	Tyr	Gly	Val	Ile	Ser	Ser
1				5					10		

(2) INFORMATION FOR SEQ ID NO:36:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 71 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:

Pro	Ile	Tyr	Gly	Val	Pro	Arg	Tyr	Ile	Thr	Tyr	Gly	Lys	Trp	Pro	Ala
1				5					10					15	

Trp Leu Thr Ala Gln Arg Pro Pro Pro Ile Asp Val Asn Asn Asp Val
 20 25 30
 Cys Ser His Ser Asn Ala Asn Arg Asp Phe Pro Leu Thr Ser Met Gly
 35 40 45
 Gly Leu Phe Thr Val Asn Cys Pro Leu Gly Ser Thr Ser Ser Val Ser
 50 55 60
 Tyr Ala Lys Tyr Ala Pro Tyr
 65 70

(2) INFORMATION FOR SEQ ID NO:37:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 2 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

Arg Gln
 1

(2) INFORMATION FOR SEQ ID NO:38:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 1 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

Arg
 1

(2) INFORMATION FOR SEQ ID NO:39:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 45 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:

Met Ala Arg Leu Ala Leu Cys Pro Val His Asp Leu Met Gly Leu Ser
 1 5 10 15
 Tyr Leu Ala Val His Leu Arg Ile Ser His Arg Tyr Tyr His Gly Asp
 20 25 30
 Ala Val Leu Ala Val His Gln Trp Ala Trp Ile Ala Val
 35 40 45

(2) INFORMATION FOR SEQ ID NO:40:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 10 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:

Leu Thr Gly Ile Ser Lys Ser Pro Pro His
 1 5 10

(2) INFORMATION FOR SEQ ID NO:41:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 18 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:

Arg Gln Trp Glu Phe Val Leu Ala Pro Lys Ser Thr Gly Leu Ser Lys
 1 5 10 15
 Met Ser

(2) INFORMATION FOR SEQ ID NO:42:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 10 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:

Gln Leu Arg Pro Ile Asp Ala Asn Gly Arg
 1 5 10

(2) INFORMATION FOR SEQ ID NO:43:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 15 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:

Ala Cys Thr Val Gly Gly Leu Tyr Lys Gln Ser Ser Leu Ala Asn
 1 5 10 15

(2) INFORMATION FOR SEQ ID NO:44:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 10 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: protein

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:

Arg	Thr	His	Cys	Leu	Leu	Ala	Tyr	Arg	Asn
1				5					10

(2) INFORMATION FOR SEQ ID NO:45:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: protein

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:45:

Leu
1

(2) INFORMATION FOR SEQ ID NO:46:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 4 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: protein

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:46:

Cys	His	Leu	Asn
1			